

Albumin and interferon- β fusion protein serves as an effective vaccine adjuvant to enhance antigen-specific CD8+ T cell-mediated antitumor immunity

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ABSTRACT

Background Type I interferons (IFN) promote dendritic cells maturation and subsequently enhance generation of antigen-specific CD8 +T cell for the control of tumor. Using type I interferons as an adjuvant to vaccination could prove to be a potent strategy. However, type I interferons have a short half-life. Albumin linked to a protein will prolong the half-life of the linked protein.

Methods In this study, we explored the fusion of albumin to IFN β (Alb-IFN β) for its functional activity both in vitro and in vivo. We determined the half-life of Alb-IFN β following treatment in the serum, tumor, and tumor draining lymph nodes in both wild type and FcRn knockout mice. We characterized the ability of Alb-IFN β to enhance antigen-specific CD8+ T cells using ovalbumin (OVA) or human papillomavirus (HPV) E7 long peptides. Next, we evaluated the therapeutic antitumor effect of coadministration of AlbIFN β with antigenic peptides against HPVE7 expressing tumor and the treatment's ability to generate HPVE7 antigen specific CD8+ T cells. The contribution of the antitumor effect by lymphocytes was also examined by an antibody depletion experiment. The ability of Alb-IFN β to serve as an adjuvant was tested using clinical grade therapeutic protein-based HPV vaccine, TACIN.

Results Alb-IFN β retains biological function and does not alter the biological activity of IFN β . In addition, Alb-IFN β extends half-life of IFN β in serum, lymph nodes and tumor. The coadministration of Alb-IFN β with OVA or HPVE7 antigenic peptides enhances antigen-specific CD8 +T cell immunity, and in a TC-1 tumor model results in a significant therapeutic antitumor effect. We found that CD8 +T cells and dendritic cells, but not CD4 +T cells, are important for the observed antitumor therapeutic effect mediated by Alb-IFN β . Finally, Alb-IFN β served as a potent adjuvant for TA-CIN for the treatment of HPV antigen expressing tumors.

Conclusions Overall, Alb-IFN β serves as a potent adjuvant for enhancement of strong antigen-specific CD8 +T cell antitumor immunity, reduction of tumor burden, and increase in overall survival. Alb-IFN β potentially can serve as an innovative adjuvant for the development of vaccines for the control of infectious disease and cancer.

Key messages

What is already known on this topic

► Interferon- β (IFN β) is a major class of immune cytokine that can contribute to the efficacy of anticancer therapies; however the short half-life and inability to target key immune system tissues make IFN β less clinically practical. Albumin (alb) is a ubiquitous plasma protein with a known extended half-life in vivo.

What this study adds

► This study describes a novel chimeric protein comprised of IFN β fused to alb (Alb-IFN β). The data presented here shows that fusing albumin to IFN β can enhance the anti-tumor immune response, extend the half-life of IFN β in vivo, and route it to lymph nodes and tumor.

How this study may affect research, practice, or policy

► This study describes a novel molecule Alb-IFN β , which pending further research may be a prime candidate for clinical translation to deliver as an adjuvant therapy to patients with cancer.

INTRODUCTION

Type I interferons are a major class of immune cytokines that also can be used as potent antiviral agents for the treatment of viruses such as hepatitis C.^{1,2} Beyond inducing antiviral immune responses, these cytokines, including both interferon- α (IFN α) and interferon- β (IFN β), elicit a plethora of signals. For example, type I interferons contribute to the efficacy of anticancer therapies and have shown to mediate antineoplastic effects against many different malignancies.³ Type I interferons also intervene in different stages of cancer immunoeediting, including the elimination of malignant cells via the immune system, the establishment of an

equilibrium between the immune system and malignant cells, as well as during the phase in which neoplastic cell variants escape due to compromised immune systems.³ Most importantly, type I interferons have been shown to trigger dendritic cell (DC) maturation and migration toward lymph nodes (LNs), both of which are important in cross-priming cytotoxic immune responses.³ During antigen presentation by plasmacytoid DCs (pDCs), a high level of type I interferon secretion is observed in a concentrated area of T cells within lymphatic tissues associated with cancer. Recently, pDCs have been shown to traffic to tumor tissues and secrete chemokines such as C-X-C motif ligand 9 (CXCL9) and CXCL10, which recruit T cells to the tumor.⁴ Type I interferons can also directly increase the cytotoxicity and survival of CD8 +T cells.⁵

In recent years, immunotherapy has begun to emerge as a potentially promising strategy for cancer treatment. Additionally, many anticancer therapies that rely on type I interferon signaling have shown success in clinical use.^{3,6} In many instances, administering type I interferons can lead to antiviral and antiproliferative bioactivities.⁷ These cytokines even possess immunostimulatory functions.⁷ As an adjuvant to standard cancer immunotherapies, type I interferons have already demonstrated improvements in disease-free survival as well as overall survival. For example, several randomized trials using IFN α as an adjuvant in both low-dose and high-dose regimens have suggested effectiveness in improving survival outcomes of melanoma patients.^{8–10} Unfortunately, type I interferons have a short 2–3 hours long plasma half-life and require weekly injections when administered as an adjuvant,^{10,11} which significantly reduces its applicability in clinical settings.

Albumin is a ubiquitous plasma protein that is known for its long half-life in vivo and ability to thus increase the half-life of molecules that are associated with it.^{12,13} This is achieved via transcytotic recycling of albumin's neonatal Fc receptor (FcRn).¹⁴ Due to its circulation pattern as a plasma protein physiologically, albumin is able to drain into the lymphatic tissues. Albumin binding has been shown to be effective for directing immunostimulatory molecules, including vaccine constructs, to the LNs in order to elicit potent immune responses.^{15,16} Albumin has low immunogenicity and is easy to construct, express, and purify, therefore it is an advantageous drug carrier.¹⁷ Albumin thus serves as a prime candidate to deliver cytokines and other biological cargo preferentially toward the LNs. Due to the ability of IFN β to promote DC expansion¹⁸ and albumin's ability to traffic toward LNs and extend half-life, we reason that a fusion between albumin and IFN β may have a profound impact on cross-priming cytotoxic immune responses and may generate large pDC populations for antigen presentation. Strategies that expand cross-presenting DC populations also have the potential to be efficacious in the treatment of cancer.

We generated fusion protein albumin-IFN β (Alb-IFN β) by genetically fusing albumin to IFN β . In this study, we evaluated the therapeutic potential of Alb-IFN β to

modulate immune cell phenotypes and improve anti-tumor responses. We show that Alb-IFN β does not alter or impede the biological activity of IFN β , as our novel molecule is able to generate DCs in vitro from bone marrow (BM) cells. The half-life of Alb-IFN β is indeed longer than that of IFN β alone, suggesting efficacy in the fusion strategy to albumin. In addition, in vivo distribution studies of Alb-IFN β show preferential accumulation of our fusion protein in the tumor-draining LNs (tdLNs). More importantly, cross-presenting DCs generated by Alb-IFN β in vivo are functional and able to generate potent antigen-specific T and B cell responses to both the ovalbumin (OVA) and human papillomavirus (HPV) E7 antigens. We also found that knocking out basic leucine zipper ATF-like transcription factor 3 (*Batf3*), which plays a crucial role in the development, expansion, and function of cross-presenting pDCs,^{19,20} reduced the antitumor effects of Alb-IFN β . Furthermore, administering Alb-IFN β as an adjuvant to a clinical grade therapeutic HPV protein-based vaccine, TA-CIN, for the treatment of HPV-associated TC-1 tumors resulted in a significant reduction in TC-1 tumor burden, improved overall survival, and upregulation of E7-specific CD8 +T cell and DC activities. We also show that the antitumor immunity elicited by our fusion protein is both CD8- and DC-dependent and CD4-independent. In response to Alb-IFN β , we observed an upregulation in CXCL9 and CXCL10 expressions in the tdLNs, which are chemoattractants secreted by DCs to recruit T cells to the tumor. Our results strongly support that Alb-IFN β makes an excellent adjuvant to immunotherapies with strong therapeutic and clinical translation implications because it is able to enhance immunological responses mediated by DCs.

MATERIALS AND METHODS

Cells

As previously described, TC-1 cells express the HPV16 E6 and E7 proteins.²¹ Cells were grown in RPMI 1640 media, supplemented with 10% (v/v) fetal bovine serum, 50 units/mL of penicillin/streptomycin, 2 mM L-glutamine, 1 mM sodium pyruvate, 2 mM non-essential amino acids, and 0.1% (v/v) 2-mercaptoethanol at 37°C with 5% CO₂. For the BMDC isolation and culture, the tibias and femurs were removed from C57BL/6 mice under sterile condition. Both ends of the bone were cut-off and BM was flushed out by 26-gage syringes with complete RPMI medium. Following red blood cell lysis and washing, BM cells were suspended with complete RPMI medium and seeded in 6-well culture plates with 29 ng/mL of granulocyte-macrophage colony-stimulating factor (GM-CSF) for 5 days.²² For the DC activation experiments, BMDCs matured in GM-CSF were treated with 0.1 μ M of Alb-IFN β , 0.1 μ M of IFN β , or 1 μ g/mL of lipopolysaccharide (LPS) (as positive control) for 16 hours.

Generation of Alb-IFN β protein constructs

For the generation of pcDNA3- Alb-IFN β , mouse IFN β was first amplified via PCR with a cDNA template of the mouse IFN β (pUNO1-mIFN β) plasmid from Invivogen (San Diego, CA 92121 USA) and the following primers: 5' AAAGAATTCATCAACTATAAGCAGCTC-3' and 5- AAAC TTAAGTCAGTTTTGGAAGTTTCT-3'. The amplified product was then cloned into the EcoRI/Afl II sites of pcDNA3-Alb.²³ The plasmid constructs were confirmed by DNA sequencing. Alb-IFN β proteins were expressed using Expi293F expression system kit (Thermo Fisher Scientific, Waltham, Massachusetts, USA) according to manufacturer's instructions. Expi293F cells were transfected with Alb-IFN β . Proteins were purified by HiTrap Albumin column (GE Healthcare Life Sciences, Marlborough, Massachusetts, USA).

Mice

Female C57BL/6 mice aged 6–8-weeks were purchased from Charles Rivers Laboratories (Frederick, Maryland, USA). All mice were maintained under specific pathogen-free conditions at the Johns Hopkins University School of Medicine Animal Facility (Baltimore, Maryland, USA). All animal procedures were performed according to protocols approved by the Johns Hopkins Institutional Animal Care and Use Committee. Recommendations for the proper use and care of laboratory animals were closely followed. For tumor inoculation, 2×10^5 TC-1 cells in 50 μ L of PBS were subcutaneously (s.c.) injected into 6–8 weeks old female C57BL/6 mice. Tumor volume was measured by digital calipers and greatest length and width were determined. Tumor volumes were calculated by the formula: tumor volume = (length \times width²)/2. For CD4⁺ and CD8⁺ T cell depletion, 100 μ g of anti-mouse CD8⁺ depleting antibody or 200 μ g of anti-mouse CD4⁺ depleting antibody were administered to tumor-bearing mice for 3 days via intraperitoneal injection. The depletion was maintained through the experiment by giving depleting antibody once a week. CD4⁺ or CD8⁺ T cell depletion level were evaluated by flow cytometry on blood.²⁴ Batf3^{-/-} and FcRN^{-/-} mice were acquired from Jackson Laboratories.

Mice vaccination

Naïve or tumor-bearing C57BL/6 mice were s.c. vaccinated in the left rear flank twice at 1-week intervals with either 100 μ g of OVA protein,^{25 26} 10 μ g of HPV16 E7 long peptide (amino acids 43–62),^{27 28} or 25 μ g of HPV vaccine TA-CIN²⁹ in combination with or without either IFN β (Prospecbio, East Brunswick NJ) or Alb-IFN β twice at 1-week interval treatments. Vaccine doses were determined by previous experiments.^{25–29} One week after the final vaccination, peripheral blood mononuclear cells (PBMCs) were collected for flow cytometric analysis.

Flow cytometry analyses

Peripheral blood samples from naïve and TC-1 tumor-bearing mice were collected into 100 μ L of PBS containing

0.5 mM EDTA. Following red blood cell lysis and washing, PBMCs were collected and stained with Zombie Aqua to determine the cell viability. Fc receptors were blocked by anti-mouse CD16/CD32 antibody. To analyze OVA- and E7-specific T cells, Fc receptor blocked PBMCs were stain with PE-conjugated SIINFEKL (OVA) peptide or HPV16 E7aa49-57 peptide loaded H-2D^b E7 tetramer and FITC-conjugated anti-mouse CD8 α antibody (Biolegend). To determine immune response of vaccinated tumor-bearing mice, TC-1 tumor-bearing mice were injected with either IFN β or Alb-IFN β followed by E7 long peptide as an adjuvant. One week after last vaccination, PBMCs were subsequently collected and stained for FITC-conjugated anti-mouse Ki67 antibodies, PE-conjugated HPV16 E7aa49-57 peptide-loaded H-2Db E7 tetramer, APC/Fire 750-conjugated anti-mouse CD8 α antibodies, APC/Fire 750-conjugated anti-mouse CD11c antibodies, APC/Fire 750-conjugated anti-mouse I-A/I-E major histocompatibility II (MHC-II) antibodies, and PE-CF594 APC/Fire 750-conjugated anti-mouse CD86 antibodies for the presence of E7-specific CD8 +T cells and DCs. To evaluate biological function between Alb-IFN β and IFN β in vitro, FcRn-blocked BMDCs were stained with BV421-conjugated anti-mouse CD11c antibodies, APC/Fire 650-conjugated anti-mouse I-A/I-E (MHC II) antibodies, PerCP-conjugated anti-mouse CD40 and PE-CF594-conjugated anti-mouse CD86. FACS analysis was performed using CytoFLEX S (Beckman) and data were analyzed by FlowJo software.

Adaptive T cell transfer and tracking

C57BL/6 mice were s.c. injected with 5×10^5 TC-1 cells for 10 days (after the tumor reached 8 to 10 mm in diameter). A 10 μ g of IFN β or 50 μ g of Alb-IFN β was intravenously injected into tumor-bearing mice through retro orbital sinus. One day after treatment with Alb-IFN β or IFN β , 5×10^6 luciferase-expressing E7-specific T cells were intravenously injected into the tumor-bearing mice via tail vein. Luciferase-expressing E7-specific T cells were generated as previously described.³⁰ For tracking the E7-specific T cell in tumor-bearing mice, 75 mg/kg of D-Luciferin was given to the mice via intraperitoneal injection and imaged by the IVIS Spectrum in vivo imaging system series 2000 (PerkinElmer) on day 1 and day 4 after T cell transfer. Total photon counts were quantified in the tumor site by using Living Image 2.50 software (PerkinElmer).

ELISA

For the half-life experiment, naïve or TC-1 tumor-bearing C57BL/6 mice were intravenously injected with 10 μ g of IFN β or 50 μ g of Alb-IFN β . Serum were collected in EDTA-free Eppendorf tube on 3, 24 and 48 hours post-treatment. Tumor and LNs were harvested 16 hours post-treatment and then minced into 1–2 mm pieces and lysis by RIPA buffer (Cell Signaling Technology, Massachusetts, USA). IFN β was measured by IFN beta Mouse ELISA Kit (Thermo Fisher Scientific) according to the manufacturer's instructions. For the OVA and HPV16 L2 antibody

detection, 1 $\mu\text{g}/\text{mL}$ of mouse OVA or HPV16 L2 protein in PBS was coated on BRANDplates microplates overnight at 4°C. After 16 hours, the plates were washed, blocked with eBioscience™ ELISA/ELISPOT Diluent (Thermo Fisher Scientific), and added diluted serum for 2 hours at room temperature. Non-vaccinated mice serum was used as control. Goat anti-mouse IgG-HRP secondary antibody was added at 1:5000 dilution for 1 hour, followed by TMB substrate. The OD at 450 nm was determined by 800 TS Absorbance Reader (BioTek Instruments).

Quantitative real-time PCR

RNA was isolated from tumors treated with either PBS, IFN β , or Alb-IFN β , or from adjusted muscle tissues (as a normal tissue control) and reverse transcribed to cDNA. Gene expression levels were measured by quantitative real-time PCR (qRT-PCR) using SYBER Green with CXCL9-specific or CXCL10-specific primers. Briefly, total RNA was extracted by the Direct-zol RNA Kits (Zymo Research) following the manufacturer's instructions. A 1 μL of RNA was converted to cDNA by the iScript Reverse Transcription Supermix (Bio-Rad Laboratories). A 1 μL of cDNA was used as template for qRT-PCR using SsoAdvanced Universal SYBR Green Supermix (Bio-Rad Laboratories), and qRT-PCR was performed using CFX96 Touch Real-Time PCR Detection System (Bio-Rad Laboratories). Primer for RT-PCR experiments to detect mouse CXCL9 expression, forward 5'- GAGCAGTGTGGAGTTC-GAGG-3'; reverse 5'- TCCGGATCTAGGCAGGTTTG-3', mouse CXCL10 forward 5'- GCCGTCATTTTCTGCCT-CAT-3'; reverse 5'- GCTTCCCTATGGCCCTCATT-3', 18S rRNA, forward 5'- GTAACCCGTTGAACCCCAT-3'; reverse 5'- CCATCCAATCGGTAGTAGCG-3' (Integrated DNA Technologies). All data shown are normalized to the internal control gene 18S rRNA.

Statistical analysis

The statistical analysis was performed using GraphPad Prism V.6 software and data were interpreted as means with SD. Kaplan-Meier survival plots are used to estimate the survival percentage and tumor-free rate. Long rank tests were used to compare the survival time between treatment groups. Comparison between individual data points were used to analyze in the t-test and p value smaller than 0.05 is considered statistically significant, * $p \leq 0.05$, ** $p \leq 0.01$, *** $p \leq 0.001$, **** $p \leq 0.0001$, ns=not significant.

RESULTS

IFN β fused to albumin retains biological function and does not alter the biological activity of IFN β

To bypass short half-life limitations posed by IFN β , a genetic fusion protein consisting of albumin and IFN β was produced and purified (figure 1A). In many instances, cytokine function can be altered when it is fused to a carrier protein. Thus, we determined whether the fusion of albumin to IFN β (Alb-IFN β) would affect

the biological function of IFN β . Through in vitro titration experiments, the expression of H-2Kb and PD-L1 on TC-1 cells increased as the cells were treated with increasing concentrations of IFN β or Alb-IFN β (figure 1B–C). BMDCs were also treated with either IFN β alone or Alb-IFN β , with BMDCs treated with LPS serving as a positive control and untreated BMDCs serving as a negative control. H-2Kb (figure 1E) and PD-L1 (figure 1G) expressions were comparable between Alb-IFN β -treated BMDCs and BMDCs treated with IFN β alone. Similarly, BMDCs treated with Alb-IFN β also expressed similar levels of CD40 (figure 1D) and CD86 (figure 1F) compared with IFN β alone. The addition of Alb-IFN β failed to increase PD-1 or LAG3 expression using both TC-1 and BMDCs (online supplemental figure 1). Taken together, our data suggest that Alb-IFN β retains similar biological function compared with IFN β alone.

The linkage of albumin to IFN β extends half-life and increases IFN β in serum, LNs and tumor in vivo

Next, we sought to better understand the underlying trafficking mechanism of Alb-IFN β in vivo. Albumin is known to increase the half-life of molecules that are associated with it through transcytotic recycling of the FcRn.^{12–14} Specifically, albumin fusion to IFN β has been shown to increase the half-life of IFN β by more than fivefold.³¹ Thus, we suspected FcRn to play a role in our albumin fusion strategy. To determine whether Alb-IFN β has an extended half-life, we intravenously injected Alb-IFN β or IFN β into C57BL/6 mice. We found that levels of IFN β were significantly higher at every time point when mice were treated with Alb-IFN β as compared with mice treated with IFN β alone (figure 2A). We also found that levels of IFN β significantly decreased in FcRn knockout mice (figure 2A), suggesting the importance of FcRn in extending the half-life of Alb-IFN β . Previous studies have also suggested that due to the circulation pattern of albumin in the plasma, albumin fusion proteins are able to preferentially traffic toward the draining LNs.^{15,16} From our experiments, we found that Alb-IFN β was present at higher levels in LNs (figure 2B), and targets to both the tumors and the tumor draining LNs (tdLNs) more efficiently than IFN β alone (figure 2C). Thus, we were able to confirm the notion that an albumin fusion strategy increases the half-life of conjugated IFN β and is somewhat reliant on the presence of FcRn. We also showed that an albumin fusion strategy allows IFN β to be targeted to both tumors and tdLNs at higher levels than being treated with IFN β alone.

Coadministration Alb-IFN β and antigenic peptides enhances antigen-specific CD8+ T cell immunity in vivo

To evaluate the ability of Alb-IFN β to promote CD8+ T cell responses to an exogenously-derived antigen, we vaccinated C57BL/6 mice with either OVA protein or E7 peptide (amino acids 43–62). It has been well documented that long E7 peptide is better than short peptides at inducing robust T cell responses, and it is capable of

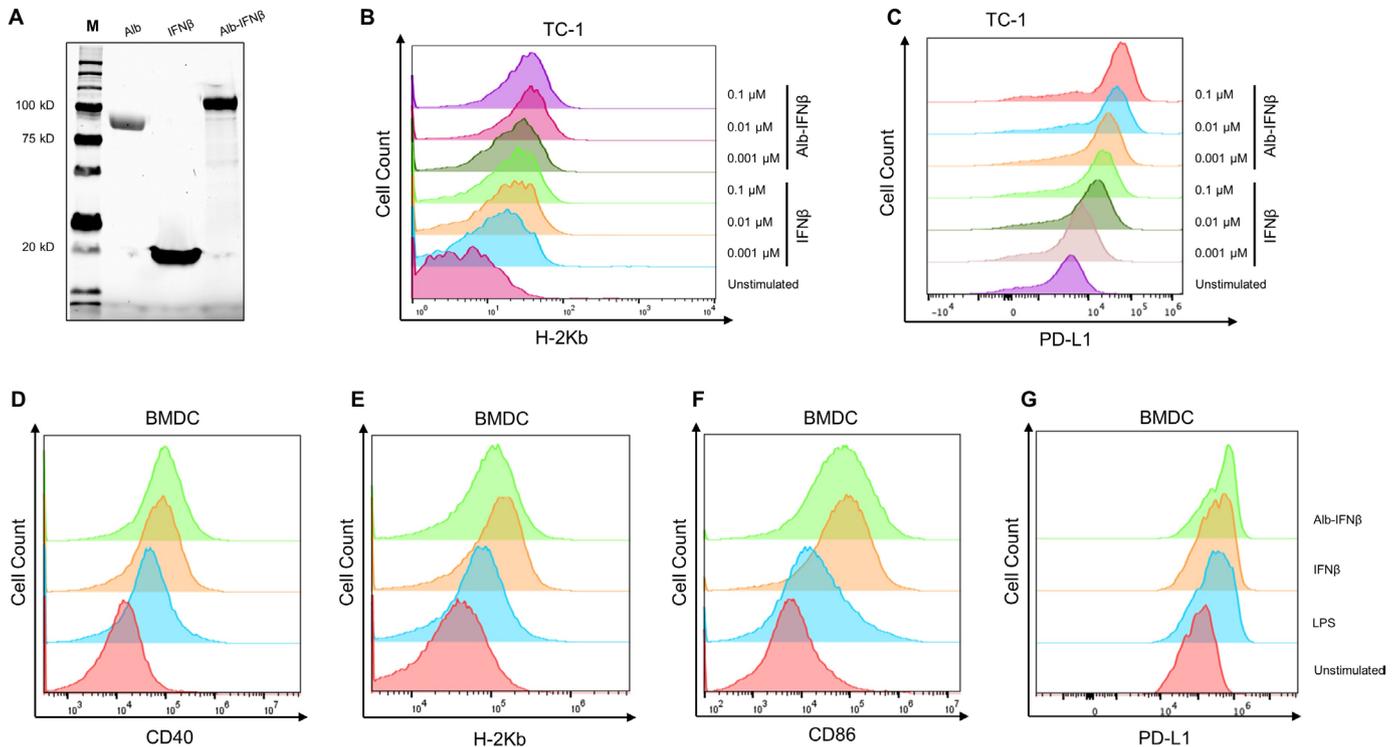


Figure 1 Characterization of biological activity of Alb-IFN β compared with IFN β using TC-1 cells or BMDCs. (A) SDS-PAGE analysis for purified albumin (Alb), IFN β , and Alb-IFN β . (M: molecular weight markers, as measured in kD). TC-1 cells were treated with 0.1 μ M, 0.01 μ M, or 0.001 μ M of either IFN β or Alb-IFN β for 24 hours. The TC-1 cells were subsequently harvested and stained for (B) H-2Kb or (C) PD-L1 expression and analyzed by flow cytometry. Representative flow cytometry images of H-2Kb expression on TC-1 cells treated with either IFN β or Alb-IFN β . (D–G) BMDCs were treated with 0.1 μ M of either IFN β or Alb-IFN β for 24 hours. BMDCs treated with lipopolysaccharide (LPS) as positive control. Shown here are representative flow cytometry images of (D) C40 expression, (E) H-2Kb expression, (F) CD86, and (G) PD-L1 expression on BMDCs treated with either IFN β or Alb-IFN β . BMDCs, bone marrow-dendritic cells. SDS-PAGE, sodium dodecyl sulfate–polyacrylamide gel electrophoresis

delivering specific cargo to antigen presenting cells.³² Additionally, OVA protein was used rather than OVA long peptide in order to demonstrate that our treatment strategy can be used with various vaccination platforms, such as protein vaccines. We simultaneously administered these antigens with or without either IFN β or Alb-IFN β to C57BL/6 mice twice at 1-week intervals (figure 3A). One week after the second vaccination, PBMCs were analyzed for the presence of OVA-specific and E7-specific CD8 +T cells by tetramer staining. Mice that received coadministration of Alb-IFN β and OVA induced the highest number of OVA-specific CD8 +T cells compared with the other treatment groups (figure 3B–C). In addition, mice treated with coadministration of E7 long peptide and Alb-IFN β similarly developed the most robust E7-specific CD8 +T cells (figure 3D–E). Finally, significantly higher titers of OVA-specific IgG2a/IgG1a antibodies were detected in the sera of mice vaccinated with both Alb-IFN β and the OVA antigen compared with mice vaccinated with IFN β and OVA or OVA alone (online supplemental figure 2). Our results suggest that coadministration of Alb-IFN β with antigen enhances antigen-specific CD8 +T cell mediated and humoral immune responses in vivo compared with coadministration of IFN β alone.

Coadministration of Alb-IFN β and E7 peptide generates a potent therapeutic antitumor effect against E7 expressing TC-1 tumor

We next looked at the antitumor properties of Alb-IFN β coadministered with E7 antigen. We treated C57BL/6 mice bearing HPV16 E7-positive TC-1 tumors with either E7 alone, Alb-IFN β alone, E7 with IFN β , or E7 with Alb-IFN β twice at 1-week intervals (figure 4A). Tumor-bearing mice administered with the E7 antigen with Alb-IFN β showed the smallest tumor volume compared with the other groups (figure 4B). Consistently, tumor-bearing mice administered with E7 with Alb-IFN β survived twice as long compared with mice treated with the other treatment groups (figure 4C). Clinically, interferon has led to the development of many side effects and potentially toxic at high doses, especially when registered with multiple different treatments.^{33,34} While the doses of IFN β and Alb-IFN β used in this study were low and all tumor-bearing mice were only treated twice, there may still be toxicity concerns. One important side effect as a result of IFN β -induced toxicity is weight loss.^{35–38} In our study, we did not observe any significant weight loss in tumor-bearing mice administered with any combination of the treatments (figure 4D). It seems that coadministration of Alb-IFN β

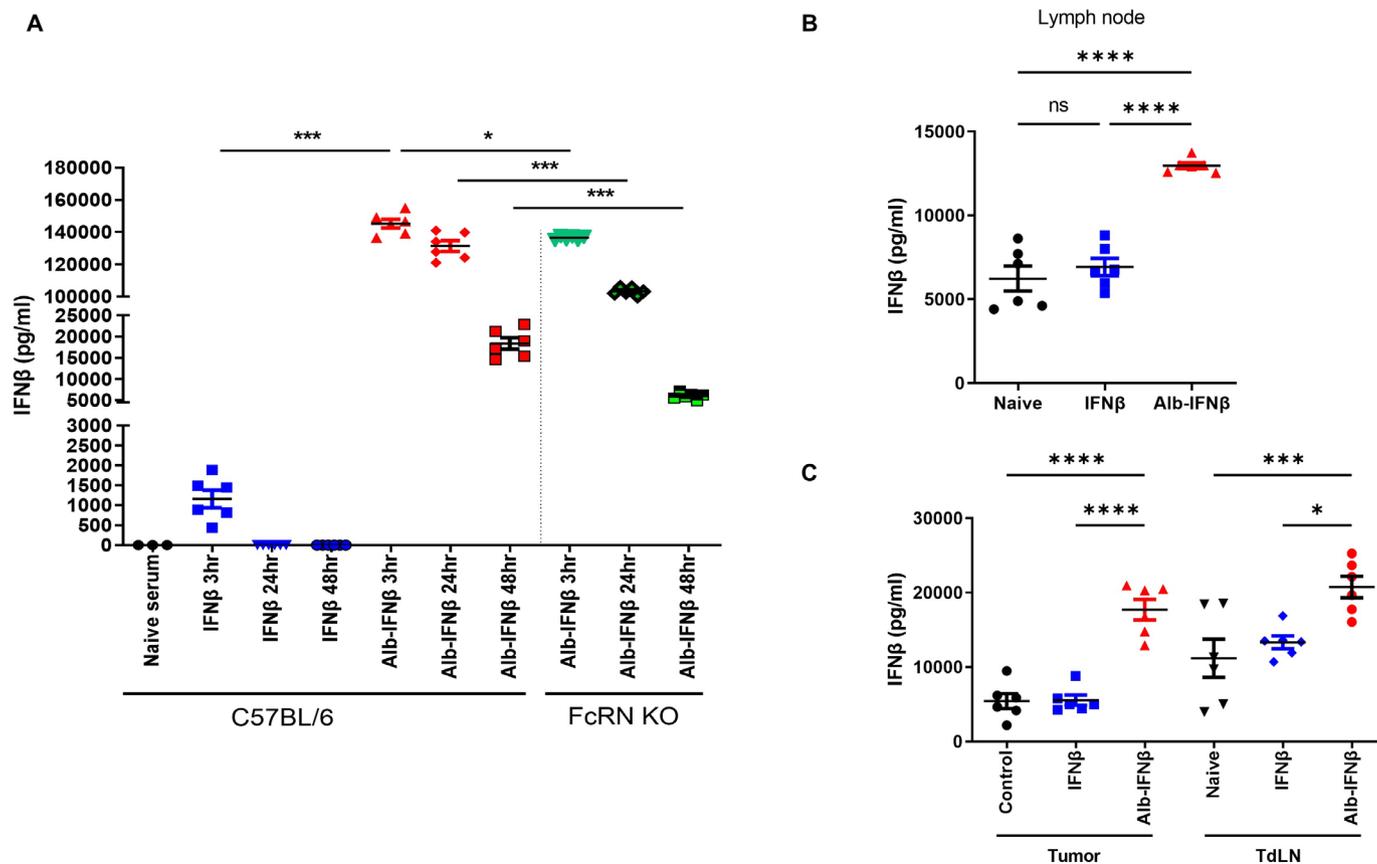


Figure 2 Analysis of Alb-IFN β half-life within the C57BL/6 or FcRn knockout mice. (A) Naive C57BL/6 or FcRN KO mice (6 mice per group) were intravenously injected with either IFN β or Alb-IFN β . Sera were collected 3, 24, and 48 hours post injection. An IFN β ELISA assay was performed. (B) Naive C57BL/6 mice (6 mice per group) were injected with either IFN β or Alb-IFN β . LNs were collected 16 hours post injection. IFN β concentration in LN protein extracts were detected by ELISA. Untreated mice LN lysates served as control. (C) TC-1 tumor-bearing mice (six per group) were treated with either IFN β or Alb-IFN β , tumors and tdLNs were harvested 16 hours post injection. IFN β concentration in tumor lysates or LN lysate were detected by ELISA. Untreated mice tumor or LN lysate served as control. * $P < 0.05$, *** $p < 0.001$, **** $p < 0.0001$. ns, not significant; tdLNs, tumor-draining lymph nodes.

and E7 peptide does not induce any noticeable toxic side effects and was able to suppress tumor growth and prolong survival rates of tumor-bearing mice better than tumor bearing mice treated with IFN β and E7 peptide.

Coadministration of Alb-IFN β with HPV E7 peptides results in enhanced E7-specific CD8 $^{+}$ T cell immune responses and DC activity

Because albumin conjugation has been shown to extend the half-life of the IFN β (figure 2), we explored whether a single s.c. administration of Alb-IFN β with E7 peptide in tumor-bearing mice can elicit E7-specific CD8 $^{+}$ T cell-mediated immune responses and enhanced DC activity. DCs are known for their potent ability to cross present exogenous antigens to cytotoxic CD8 $^{+}$ T cells. While we have demonstrated that fusion of albumin to IFN β does not impede the ability of IFN β to expand DCs, we further examined whether coadministration of Alb-IFN β with E7 antigen was superior in expanding DCs and promoting cytotoxic T cell responses to E7 antigens in vivo compared with coadministration of IFN β with E7 antigen. TC-1 tumor bearing C57BL/6 mice were vaccinated with either

E7 alone, Alb-IFN β alone, IFN β with E7, or Alb-IFN β with E7. PBMCs were then collected from the mice for analysis. There was a significantly higher amount of E7-specific CD8 $^{+}$ T cells in mice treated with Alb-IFN β and E7 compared with all other treatment conditions in the TC-1 tumor-bearing mice, indicating potent expansion of E7-specific CD8 $^{+}$ T cells following coadministration with Alb-IFN β with E7 antigen (figure 5A–B). Moreover, mice treatment with Alb-IFN β and E7 also had the highest levels of DC activation marker CD86 (figure 5C). In order to characterize the immune cell proliferation, we next examined the proliferative marker Ki67 following treatment.³⁹ The advantage of using Ki67 for lymphocyte proliferative assays is to indicate the function of E7 specific T cells. Alb-IFN β and E7 vaccinated mice also exhibited the highest proliferation activity of E7-specific CD8 $^{+}$ T cells in tumor-bearing mice compared with all other vaccination regimens (figure 5D). DCs in Alb-IFN β and E7 vaccinated mice also had significantly higher Ki67 proliferative expression than other treatment conditions (figure 5E). Thus, our data suggest that Alb-IFN β is able

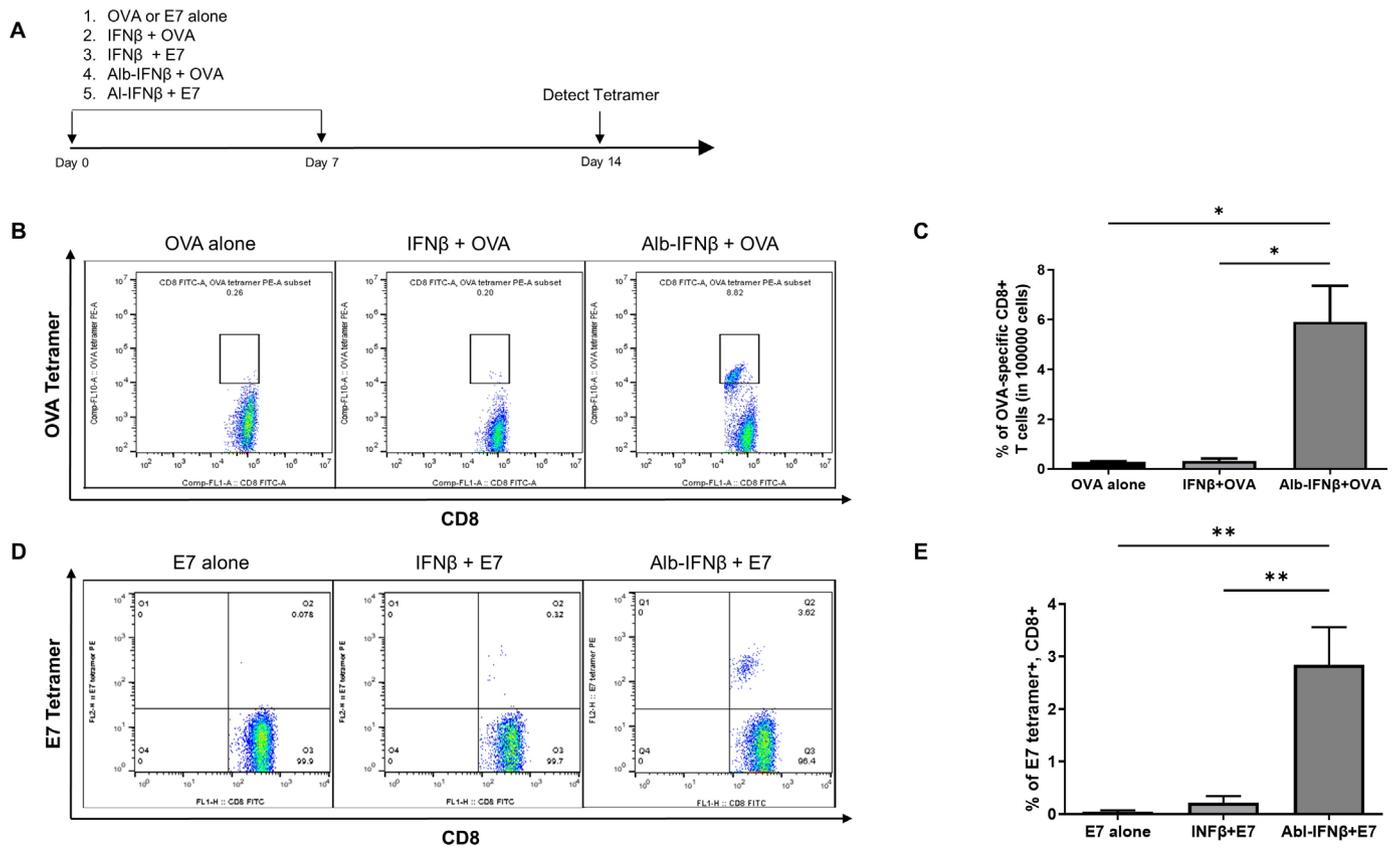


Figure 3 Characterization of the antigen specific CD8 +T cell immune response in mice treated with Alb-IFN β mixed with OVA or E7. (A) Schematic illustration of the experiment. Briefly, naive C57BL/6 mice (five per group) were subcutaneously vaccinated with antigen (either OVA protein or E7 long peptide (amino acids 43–62)) alone, or in combination with either IFN β or Alb-IFN β s.c. once a week at day 0 and 7. Seven days after the final vaccination, PBMCs were collected from previously-treated mice and stained with either PE-conjugated H-2 Kb OVA (SIINFEKL) or PE-conjugated HPV16 E7aa49–57 peptide-loaded H-2D^p E7 tetramer and FITC-conjugated anti-mouse CD8 α antibodies, followed by flow cytometry analyses. (B) Representative flow cytometric images of OVA tetramer staining. (C) Bar graph summary of OVA tetramer staining. (D) Representative flow cytometric images of E7 tetramer staining. (E) Bar graph summary of E7 tetramer staining. *P<0.05, **p<0.01. OVA, ovalbumin; PBMCs, peripheral blood mononuclear cells; s.c, subcutaneously.

to generate and expand more potent antigen-specific cytotoxic T cell and DCs compared with IFN β when coadministered tumor antigen in tumor-bearing mice.

CD8 +T cells and cross presenting DCs, but not CD4 +T cells, are important for the observed antitumor therapeutic effect mediated by Alb-IFN β coadministered with E7 peptides

After determining Alb-IFN β elicits antitumor immunity through the expansion of both DCs and E7-specific CD8 +T cells when coadministered with E7 antigen, we wanted to test for the subset of lymphocytes that are essential for Alb-IFN β to produce its effects. We first depleted either CD4 +or CD8+T cells in TC-1 tumor-bearing mice (n=5) by injecting them with either 200 μ g of anti-mouse CD4 antibodies or 100 μ g of anti-mouse CD8 antibodies daily for 3 days, followed by Alb-IFN β and E7 vaccination (figure 6A). Our results suggest that CD8 +T cell depletion completely abolished the anti-tumor effects generated by Alb-IFN β and E7 vaccination (figure 6B) and decreases the survival rate of Alb-IFN β and E7 vaccinated tumor-bearing mice (figure 6C). However, the tumor

volume and survival rates of Alb-IFN β and E7 -treated tumor-bearing mice did not significantly change by depletion of CD4 +T cells (figure 6D–E). Furthermore, *Batf3* is known to play an important role in the development, expansion, and function of cross-presenting DCs.¹⁵ Thus, we perform the anti-tumor experiment in *Batf3* KO mice to determine whether it affected antitumor effect of Alb-IFN β and E7 vaccination. *Batf3* KO mice treated with Alb-IFN β and E7 vaccination apparently reduced their ability to control the tumor progression (figure 6F) and generated fewer E7-specific CD8 +T cells (figure 6G). Taken together, our results suggest both E7-specific CD8 +T cells and cross-presenting DCs are important for Alb-IFN β to properly elicit potent antitumor responses when coadministered with E7 antigen.

Treatment with Alb-IFN β increased antigen-specific CD8+ T lymphocytes in the tumor microenvironment

To understand how Alb-IFN β affects antigen-specific CD8 +T cells trafficking to the tumor microenvironment (TME), tumor-bearing mice were treated with either

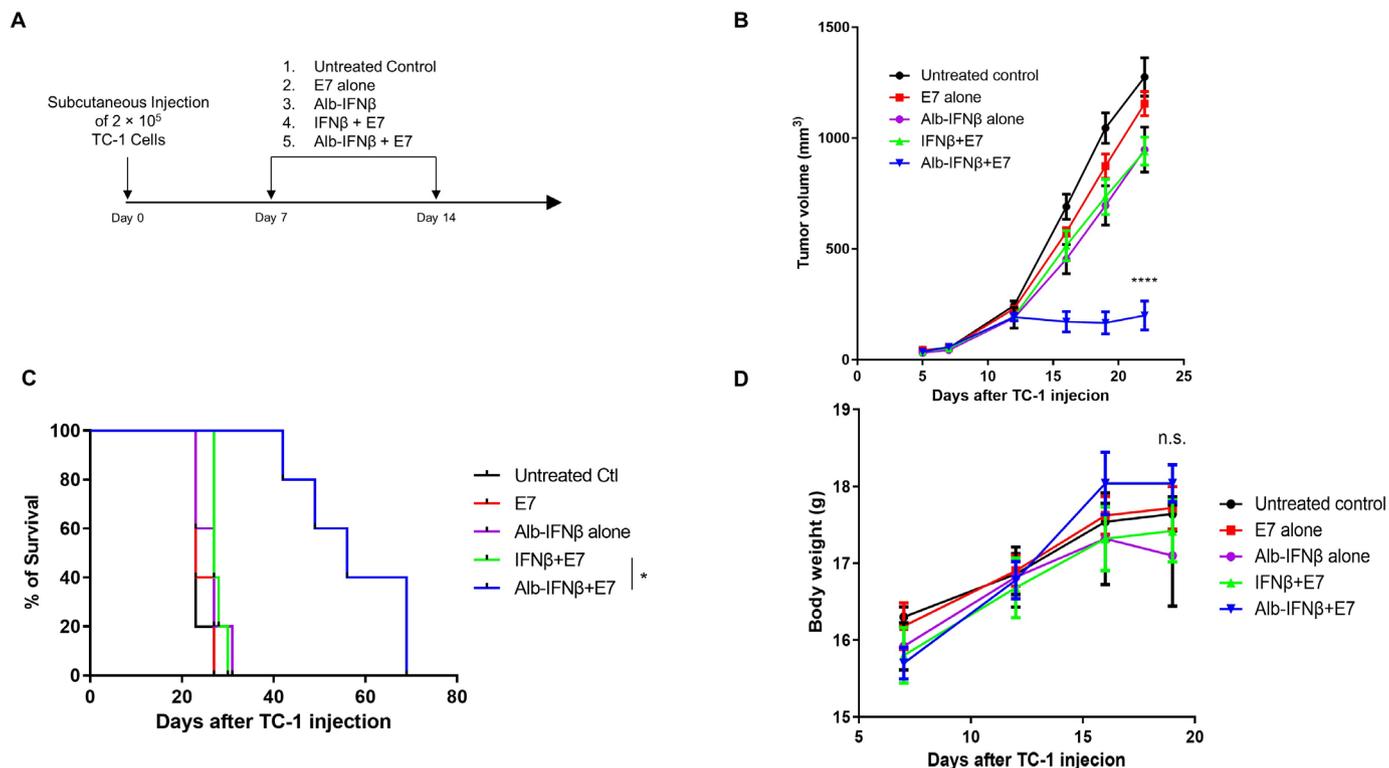


Figure 4 Characterization of therapeutic antitumor effect generated by Alb-IFN β in conjunction with E7 vaccination. (A) Schematic illustration of the experiment. Briefly, C57BL/6 mice (five per group) were s.c. injected with 2×10^5 TC-1 cells on day 0. These TC-1 tumor-bearing mice were then vaccinated with either E7 alone, Alb-IFN β alone, IFN β with E7, or Alb-IFN β with E7 on days 7 and 14 after TC-1 tumor inoculation. A group of untreated TC-1 tumor-bearing mice served as the control group. (B) Tumor growth curve of TC-1 tumor-bearing mice. (C) Kaplan-Meier survival curve of TC-1 tumor-bearing mice. (D) Change in body weights of TC-1 tumor-bearing mice treated with the various therapeutic approaches. * $P < 0.05$, **** $p < 0.0001$. n.s., not significant; s.c., subcutaneously.

Alb-IFN β , IFN β , or PBS control followed by adoptive transfer of luciferase-expressing E7-specific CD8 +T cells (see online supplemental figure 3). By day 4, E7-specific CD8 +T cells were highly accumulated in the tumor area of mice administered with Alb-IFN β compared with IFN β (online supplemental figure 3). In comparison, tumor bearing mice administered with IFN β did not demonstrated impact to the number of E7-specific CD8 +T cell in the tumor compared with untreated group. Taken together, our data indicated that administration of Alb-IFN β facilitates tumor infiltration of E7-specific CD8 +T lymphocytes in the TME.

Treatment with Alb-IFN β leads to increased levels of chemokines in tumors and increased CD8+ T cell activity and DC activation in the tDLNs

Cross-presenting DCs have been shown to secrete chemokines such as CXCL9 and CXCL10. These chemokines are then able to recruit T cells to the TME, thus mounting an antitumor immune response.⁴ To test whether Alb-IFN β can promote the expression of these chemokines in the tumors, we analyzed DC activation in the TME and changes in chemokine expression following Alb-IFN β treatment. The levels of CXCL10 and CXCL9 were significantly higher in tumors treated with Alb-IFN β compared mice treated with to IFN β (online supplemental figure

4A-B). Within the tDLNs, mice treated with Alb-IFN β exhibited significantly higher CD8 +T cell proliferative activity in the tDLNs of tumor-bearing mice compared with untreated control mice. However, there is no significant difference between mice treated with Alb-IFN β or IFN β (online supplemental figure 4C). Additionally, Alb-IFN β was able to induce higher DC activation in the tDLNs compared with IFN β (online supplemental figure 4D). Thus, our experiments showed that Alb-IFN β treatment successfully increases chemokine expression, which also increases CD8 +T cell activity and DC maturation in the tDLNs of tumor bearing mice.

Alb-IFN β serves as a potent adjuvant for HPV protein based therapeutic vaccine, TA-CIN, for the treatment of HPV antigen expressing tumors

Tissue Antigen-Cervical Intraepithelial Neoplasia (TA-CIN) is a candidate therapeutic HPV protein vaccine comprised of a fusion of full length HPV16 L2, E6, and E7 proteins.⁴⁰ It is administered as a filterable protein aggregate to promote uptake by antigen presenting cells. This protein vaccine has been shown to induce both E7-specific CD8 +T cell-mediated antitumor and HPV L2-specific neutralizing antibody responses in preclinical models.^{16 40 41} However, the clinical efficacy of TA-CIN alone may not be as effective as intended, probably due to the lack of immunogenic

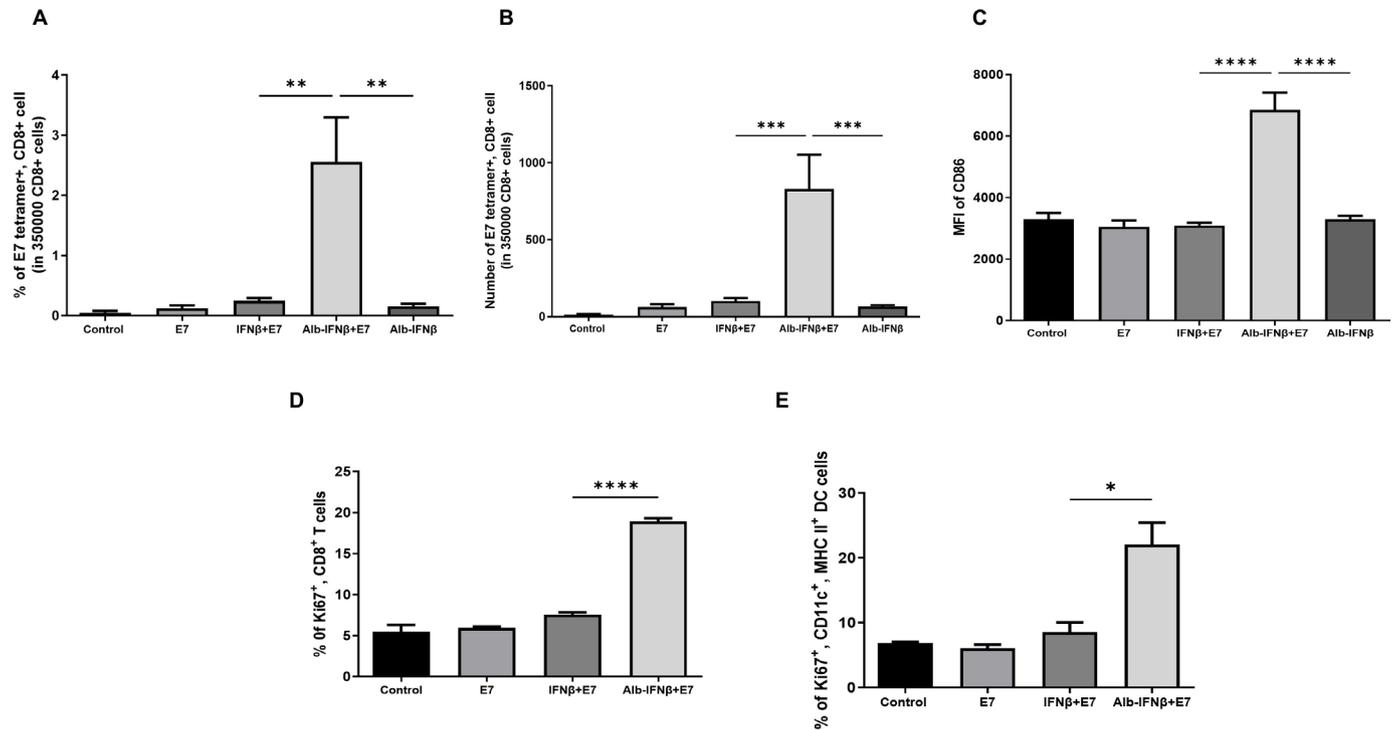


Figure 5 Characterization of E7-specific CD8 +T cells and DCs in PBMCs following treatment with Alb-IFN β and E7 vaccination. (A–C) PBMCs were collected from the previously treated mice in figure 4 on 1 week after last vaccination (five per group), and stained with PE-conjugated HPV16 E7aa49–57 peptide-loaded H-2D^p E7 tetramer. (A) Bar graph summary of the percentages of E7 tetramer and CD8 +T cells in tumor-bearing mice administered with either E7, Alb-IFN β , IFN β with E7, or Alb-IFN β with E7. (B) Bar graph summary of the total number of E7 tetramer and CD8 +T cells in tumor-bearing. (C) Bar graph summary of the mean fluorescence intensity of CD86 in CD11c+ and I-A+/I-E +DCs. (D, E) PBMCs were collected from mice (five per group) treated with E7 alone, IFN β with E7, or Alb-IFN β with E7, which were independent from the mice used in A–C, and subsequently stained with FITC-conjugated anti-mouse Ki67 antibodies, APC/Fire 750-conjugated anti-mouse CD8 α antibodies, APC/Fire 750-conjugated anti-mouse CD11c antibodies, and APC/Fire 750-conjugated anti-mouse I-A/I-E antibodies. (D) Bar graph summary of the percentages of proliferative Ki67 + and CD8+T cells. (E) Bar graph summary of percentage of proliferative Ki67+, CD11c+, and I-A+/I-E +DCs. *P<0.05, **p<0.01, ***p<0.001, ****p<0.0001. DC, dendritic cell; PBMCs, peripheral blood mononuclear cells.

adjuvants in the formulation of the protein vaccine.^{42,43} Thus, we sought to overcome TA-CIN immunogenic deficiencies by combining it with Alb-IFN β treatment. Tumor-bearing mice were administered either TA-CIN or TA-CIN in combination with Alb-IFN β (figure 7A). TC-1 tumor-bearing mice receiving TA-CIN treatment in combination with Alb-IFN β had significantly lower tumor growth compared with mice that were vaccinated with TA-CIN alone (figure 7B). Additionally, combination treatment of TA-CIN and Alb-IFN β was also more effective in prolonging the survival of tumor-bearing mice than TA-CIN alone or untreated tumor-bearing mice (figure 7C). Combination treatment of Alb-IFN β and TA-CIN also induced higher levels of E7-specific CD8 +T cells in tumor-bearing mice (figure 7D). Significantly higher level of anti-L2 IgG antibodies were similarly detected in the combination group compare to TA-CIN alone (figure 7E). Taken together, we show that Alb-IFN β is able to enhance TA-CIN-elicited antitumor effects to suppress tumor growth and we believe that Alb-IFN β serves as a potentially potent immunologic adjuvant.

DISCUSSION

In this study, we evaluated the therapeutic potential of Alb-IFN β in combination with antigens to modulate immune cells and improve antigen-specific antitumor responses. Our data shown Alb-IFN β not only retains similar biological activity compared with IFN β in vitro but is able to generate potent antigen-specific T and B cell responses to OVA and HPV16 proteins. Additionally, vaccination of Alb-IFN β and HPV16 antigens in tumor-bearing mice resulted in a significant reduction in tumor burden and better overall survival. The antitumor immune responses generated by Alb-IFN β and HPV16 antigens vaccination were found to be CD8-dependent and DC-dependent and CD4-independent. One possible explanation for why CD4 +T cells are not as important for the antitumor effect is that it has been documented that mice vaccinated with IFN α and OVA antigens generate OVA specific CD8 independent of CD4 or CD40.⁴⁴ This is likely because Alb-interferons cause maturation of DCs and also provide a third signal to enhance CD8 proliferation.⁴⁵ We also observed a significant increase of the antigen-specific CD8 +T cells in the tumor location was observed in tumor bearing mice treated with Alb-IFN β . Alb-IFN β also

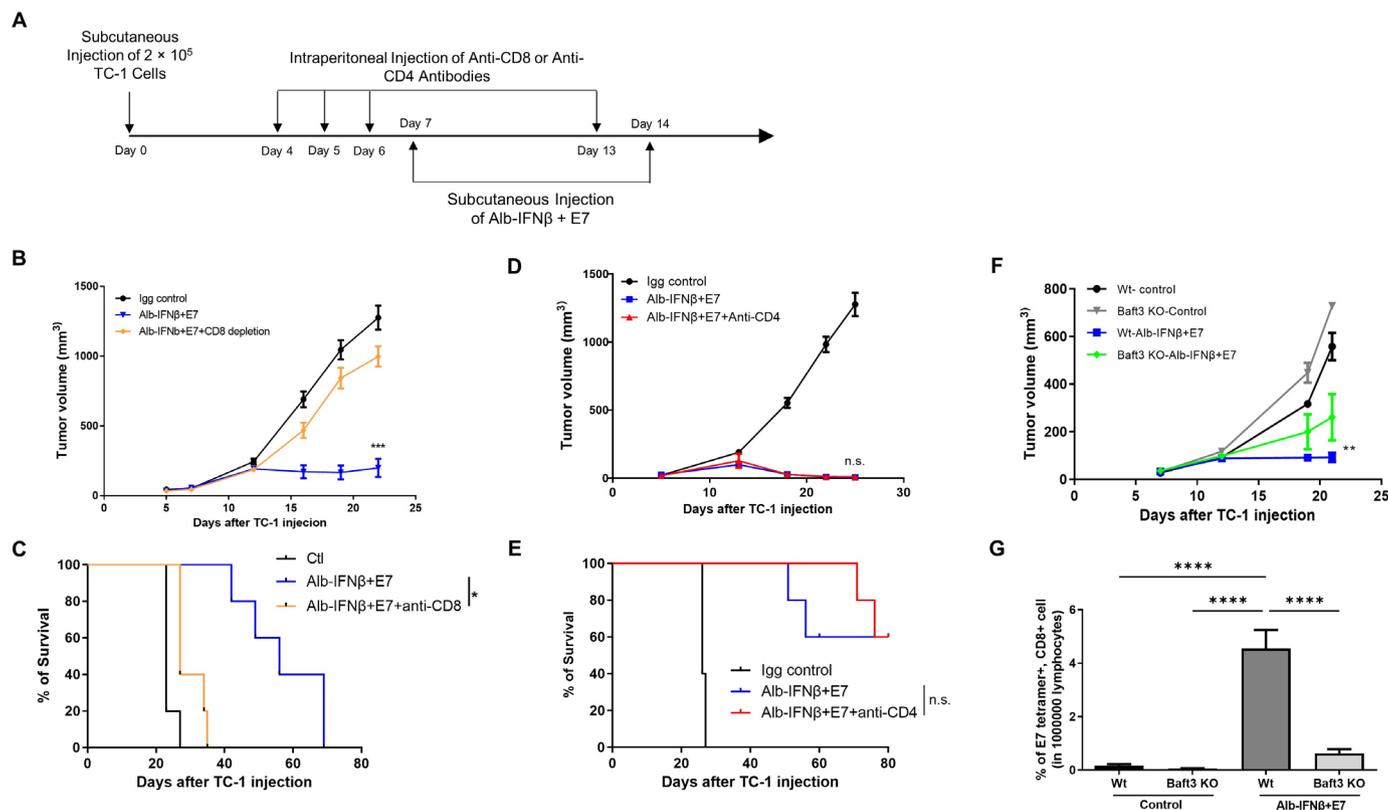


Figure 6 Determination of the role of CD8 T cells, CD4 T cells, or dendritic cells on therapeutic antitumor immunity generated by Alb-IFN β and E7 vaccination. (A) Schematic illustration of the experiment. To deplete CD4 +or CD8 +T cells in TC-1 tumor-bearing mice (five per group), C57BL/6 mice received either 200 μ g of anti-mouse CD4 antibodies or 100 μ g of anti-mouse CD8 antibodies daily by intraperitoneal injection for three continuous days prior to Alb-IFN β treatment. Control mice received the same dose of mouse IgG isotype antibodies. (B) Tumor growth curve of CD8 +T cell-depleted mice. (C) Kaplan-Meier survival of CD8 +T cell-depleted mice. (D) Tumor growth curve of CD4 +T cell-depleted mice. (E) Kaplan-Meier survival of CD4 +T cell-depleted mice. To determine the significance of DC cells capable of cross presentation for the antitumor effect in TC-1 tumor-bearing mice, Baft3 KO mice were used. (F) Tumor growth curve of Baft3 KO mice (G) Bar graph summary of the percentages of E7 tetramer and CD8 +T cells in Baft3 KO or control tumor-bearing mice administered with Alb-IFN β with E7. * $P < 0.05$, ** $p < 0.01$, *** $p < 0.001$, **** $p < 0.0001$. DC, dendritic cell; n.s, not significant.

accumulates in the tLNs and facilitates the expansion of antigen-specific CD8 +CTLs in the TME. We suggested Alb-IFN β can increase CD8 +T cell activities and promote DC maturation in the tLNs possibly through inducing an upregulation of CXCL9 and CXCL10. An assessment of Alb-IFN β used in combination with a clinical drug TA-CIN showed superior antitumor effects compared with TA-CIN alone, therefore suggesting Alb-IFN β as an effective immunologic adjuvant. The therapeutic potential of Alb-IFN β lead us to believe that it should be further investigated for clinical translation.

Alb-IFN β holds immense therapeutic potential as a novel immunotherapy. With Alb-IFN β we could possibly improve treatment schedules while limiting any side effects to generate potent antigen-specific antitumor responses. The linkage of Albumin to IFN β not only extends half-life and but also leads to the targeting of IFN β to the LNs and tumor in vivo and thereby serves as a potent adjuvant for vaccination. Thus, Alb-IFN β can bypass shortcomings posed by the weekly administrations and increased dosages of IFN β thereby limiting potential side effects in the clinic. Although PEGylated IFN α and

IFN β have also demonstrated increased half-lives in vivo, Alb-IFN β could likely better target the interperitoneal lymph nodes based on its natural circulation. Therefore, Alb-IFN β has a higher chance of contacting immune cells in LNs, and subsequently enhanced the DC cross-presentation. However, future studies comparing the half-life and effectiveness of PEGylated IFN β and Alb-IFN β should be considered.

Anticancer immunotherapies harness the immune system to develop a response toward tumors. Immunotherapy can include checkpoint inhibitors, adoptive cell therapies, and cancer vaccines, among other approaches.⁴⁶⁻⁴⁷ There are many cancer vaccine delivery methods, including through intratumoral and localized mucosal routes.⁴⁸⁻⁵¹ However, these delivery methods are invasive, therefore limiting the number of participants willing to take part in clinical settings.⁵²⁻⁵⁴ An alternative approach is to target tLNs, which are known to accumulate tumor antigens that can be used to prime antitumor T cell responses.⁵⁵ In our study, we show that Alb-IFN β is able to target the tLNs (figure 2). With our albumin-fusion targeting strategy, we can locally

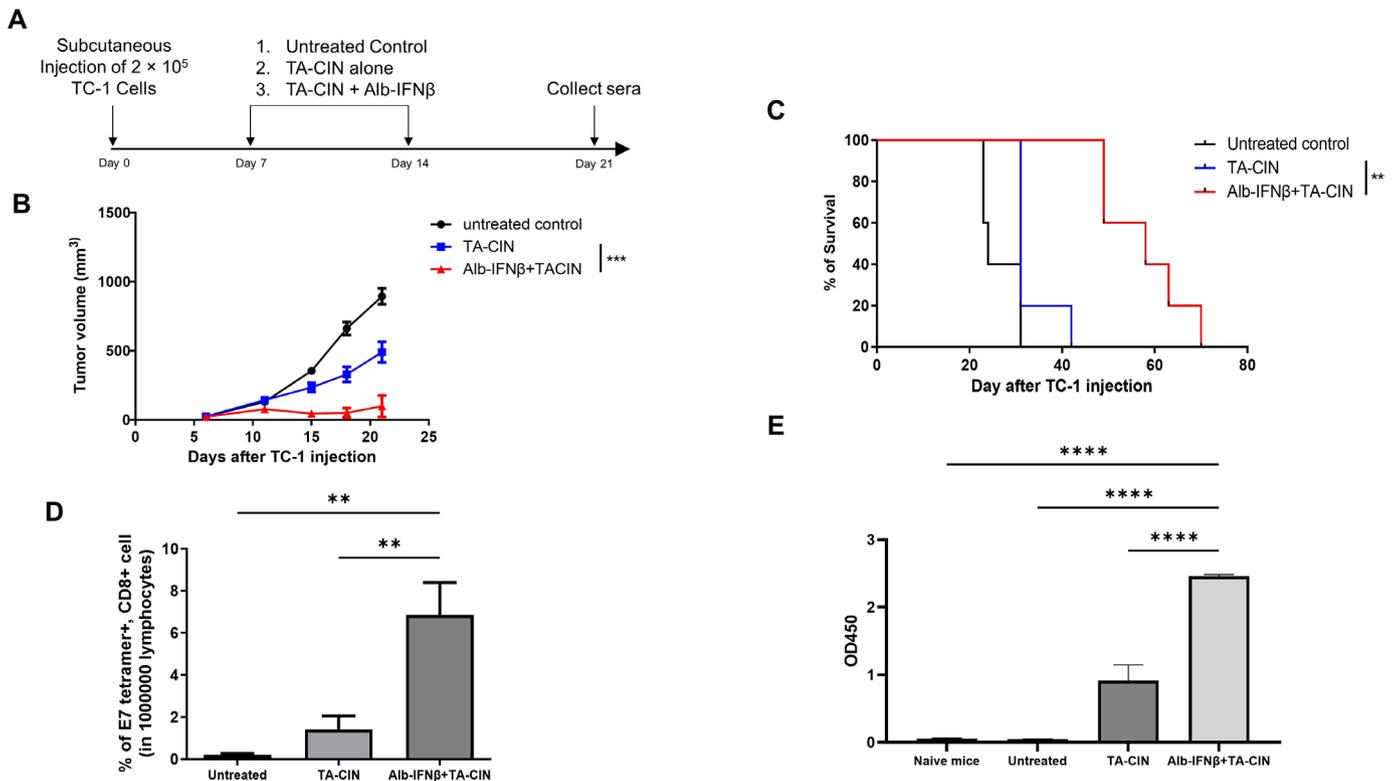


Figure 7 Characterization of therapeutic anti TC-1 tumor effect generated by Alb-IFN β in combination with TA-CIN. (A) Schematic illustration of the experiment. C57BL/6 mice (five per group) were s.c. injected with 2×10^5 TC-1 cells on day 0. 7 days after TC-1 injection, TA-CIN alone or in combination with Alb-IFN β was s.c. injected into the mice every week for 2 weeks. Untreated TC-1 tumor-bearing mice served as control. (B) Tumor growth curve of TC-1 tumor-bearing mice. (C) Kaplan-Meier survival curve of TC-1 tumor-bearing mice. (D) Bar graph summary of the percentage of E7 tetramer and CD8 +T cells in tumor-bearing mice administered with either TA-CIN or Alb-IFN β and TA-CIN. Untreated mice served as control. (E) Bar graph summary of the ELISA assay to detect mouse anti-L2 IgG2A antibodies in vaccinated mice. Sera from vaccinated and untreated mice were collected on day 21. Sera from naïve mice served as negative control. The sera were diluted by 1000-fold. **p<0.01, ***p<0.001, ****p<0.0001. s.c, subcutaneously. TA-CIN, Tissue Antigen-Cervical Intraepithelial Neoplasia

administer a less-invasive procedure that can target therapeutic vaccines toward the tDLNs in order to elicit potent, local antigen-specific antitumor responses within the TME (online supplemental figure S2). Alb-IFN β therefore provides immense clinical opportunities to deliver antigens to DCs at the TME and expand robust cytotoxic immune responses. Additionally, in the current study we observed increased luciferase expressing antigen specific CD8 +T cells in the TME following treatment with Alb-IFN β . There are at least two reasons that may account for the observed phenomenon. First, it may be attributed to the trafficking of the antigen specific CD8 +T cells to the location of the tumor (as implied by the study with CXCL9 and CXCL10). Second is that it may be due to the proliferation of antigen specific CD8 +T cells at the tumor location (as suggested by the characterization of Ki67).

In the current study, we have found that both *Batf3* is an important factor for the ability of Alb-IFN β to control tumor progression. IFN β enhances cross-presenting DC maturation, whereas *Batf3* is crucial to the development, expansion, and functioning of cross-presenting DCs.^{15 56-58} Thus, we used *Batf3* KO mice to study the role of this gene in the ability of Alb-IFN β to expand

cross-presenting DCs. We show that *Batf3* KO mice administered with Alb-IFN β were less capable to control tumor growth progression and generated fewer E7-specific CD8 +T cells (figure 6). Of note, *Batf3* KO mice treated with Alb-IFN β still were able to control tumor growth compared with untreated mice were, suggesting that although *Batf3* alone is important for Alb-IFN β effect it is not the only contributing factor. Other factors may also contribute to the ability of Alb-IFN β to control tumor in addition to *Batf3*.

In our study, we found that FcRn is an important mediator for the ability of albumin to extend the half-life of IFN β . When administering Alb-IFN β to FcRn KO mice, we noticed a shorter half-life of Alb-IFN β compared with Alb-IFN β in C57BL/6 mice (figure 2). However, despite a significant decrease in the half-life of Alb-IFN β in FcRn KO mice, the half-life was still longer than the half-life of IFN β in C57BL/6 mice. Thus, although FcRn can extend the half-life of IFN β linked to albumin, some other factors may also contribute to the prolonged half-life mediated by albumin. It is of interest to further explore the other possible mechanisms that account for the prolongation of half-life of the protein fused to albumin.



We have observed tumor-bearing mice treated with Alb-IFN β resulted in more tumor antigen specific CD8 +T cells in the tumor location (online supplemental figure S2). At least two reasons may account for the observed phenomenon. One is that the antigen specific T cells may be preferentially attracted to the tumor location in tumor-bearing mice treated with Alb-IFN β . Alternatively, the other reason is that tumor-bearing mice treated with Alb-IFN β results in enhanced proliferation of antigen specific CD8 +T cells in the tumor location. Indeed, our data from the characterization of CXCL9 and CXCL10 in the tumor location appears to be higher (online supplemental figure S3). In fact, type I interferons have been shown to induce CXCL10 and CXCL9 production in DCs and subsequently enhance their ability to stimulate CD8 +effector T cells.^{59,60} However, other IFN-modulated chemokines/cytokines couple be involved, potentially in a different manner in the tumor or LN.^{61–64} Thus, further exploration on how other chemokines/cytokines may be impacted by Alb-IFN β should be considered.

Alb-IFN β may serve as a protein based adjuvant that can be used to enhance protein based vaccines. It would be important to further test whether Alb-IFN β can be used as an adjuvant to improve vaccine efficacy of other types of protein based vaccines or other forms of vaccine, such as DNA/RNA based, cell based, or vector based vaccines. This information would create the opportunity for wide application of Alb-IFN β to enhance vaccine potency. For clinical translation, it will be important to further characterize the toxicity generated by Alb-IFN β . The understanding of the ability of Alb-IFN β to enhance vaccine potency as well as the toxicity associated with the coadministration of Alb-IFN β will be critical for the assessment whether Alb-IFN β will serve as a better adjuvant compared with other adjuvants. Such information will be critical for the development of vaccines against infections and cancers.

Contributors S-HT, BL, LL and YJK contributed to the conduction of the experiments. S-HT, MAC, EF and YJK contributed to the original draft of the manuscript. LF contributed to the editing of the manuscript. T-CW and C-FH supervised and conceptualized the study, and interpreted the data. C-FH is the guarantor of the work.

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Competing interests T-CW is a cofounder of and has an equity ownership interest in Papivax. Also, T-CW owns Papivax Biotech stock and is a member of Papivax Biotech's Scientific Advisory Board.

Patient consent for publication Not applicable.

Ethics approval Not applicable.

Provenance and peer review Not commissioned; externally peer reviewed.

Data availability statement Data are available on reasonable request. All data and materials are available from the corresponding author on written request.

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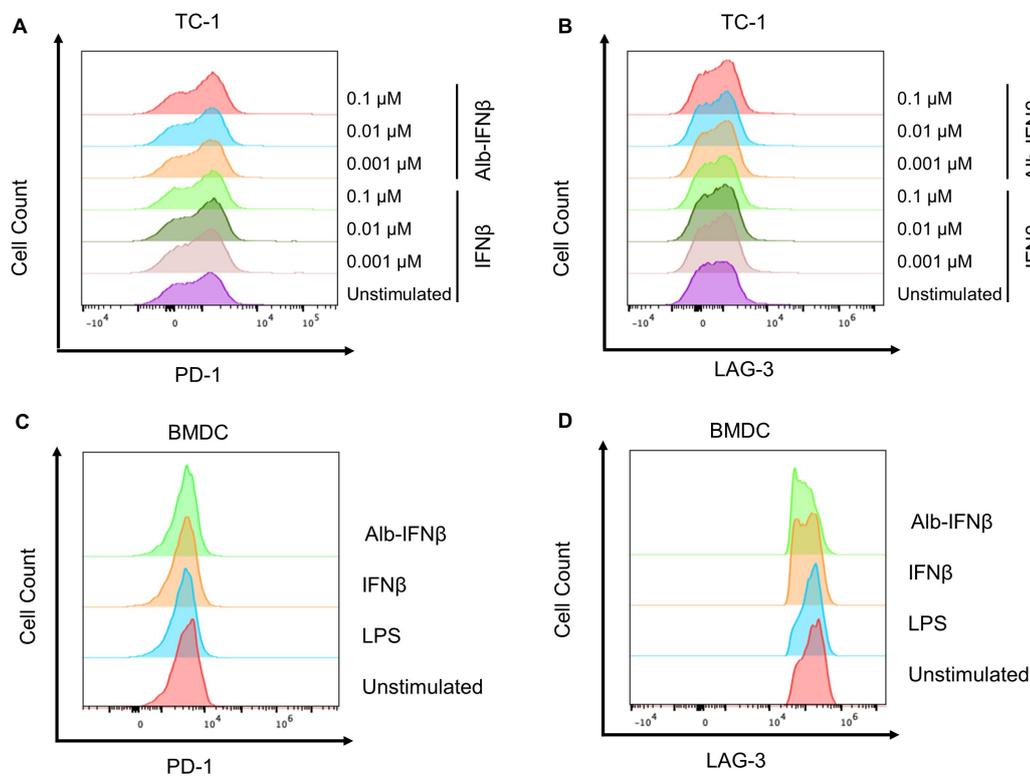
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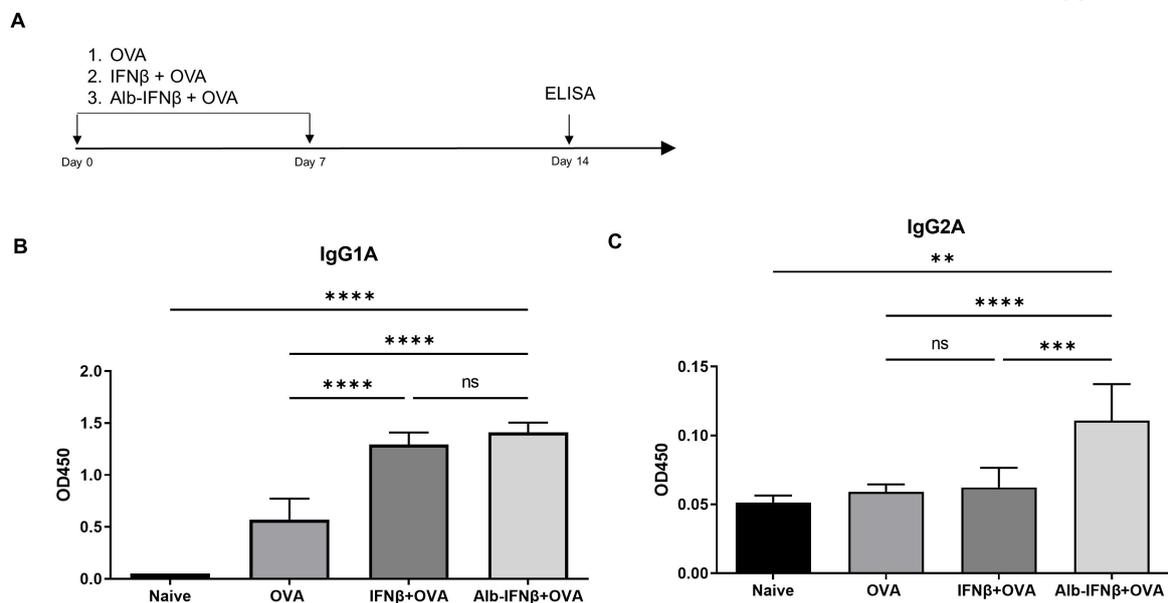
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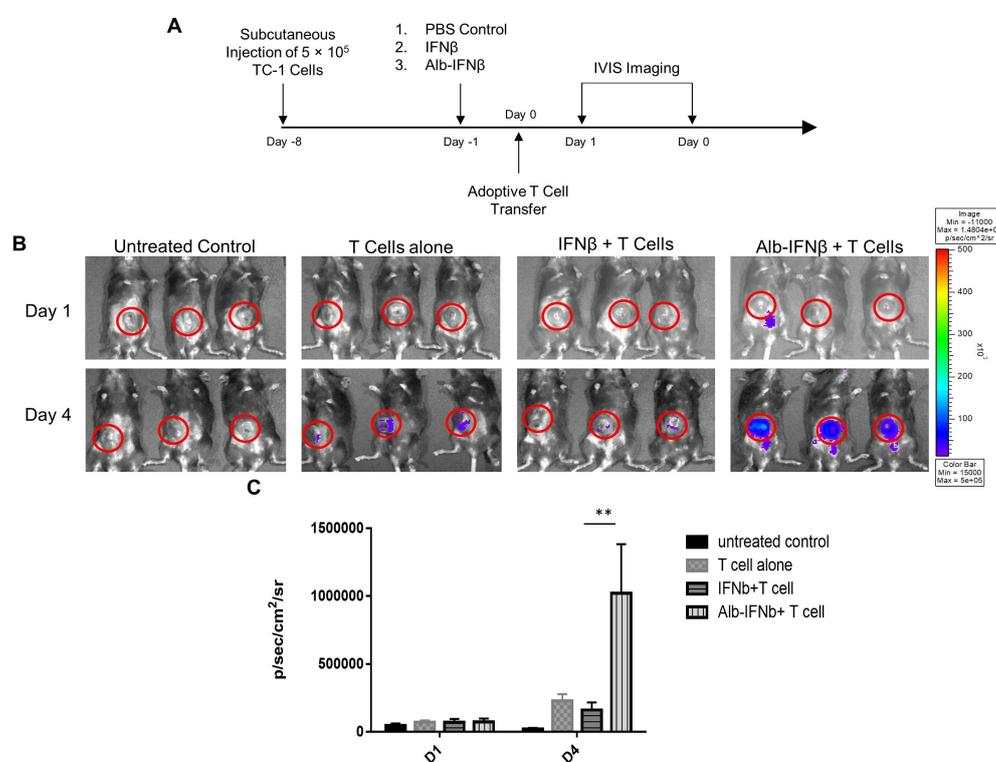


Supplemental Figure 1
Characterization of biological activity of Alb-IFN β compared to IFN β using TC-1 cells and BMDCs. TC-1 cells were treated with 0.1 μM , 0.01 μM , or 0.001 μM of either IFN β or Alb-IFN β for 24 hrs. The TC-1 cells were subsequently harvested and analyzed with flow cytometry for the expression of **(A)** PD-1 and **(B)** LAG-3 expression. BMDCs were treated with 0.1 μM of either IFN β or Alb-IFN β for 24 hrs. BMDCs treated with lipopolysaccharide (LPS) as positive control. Shown here are representative flow cytometry images of **(C)** PD-1 and **(D)** LAG-3 expression.

Supplemental Figure 2

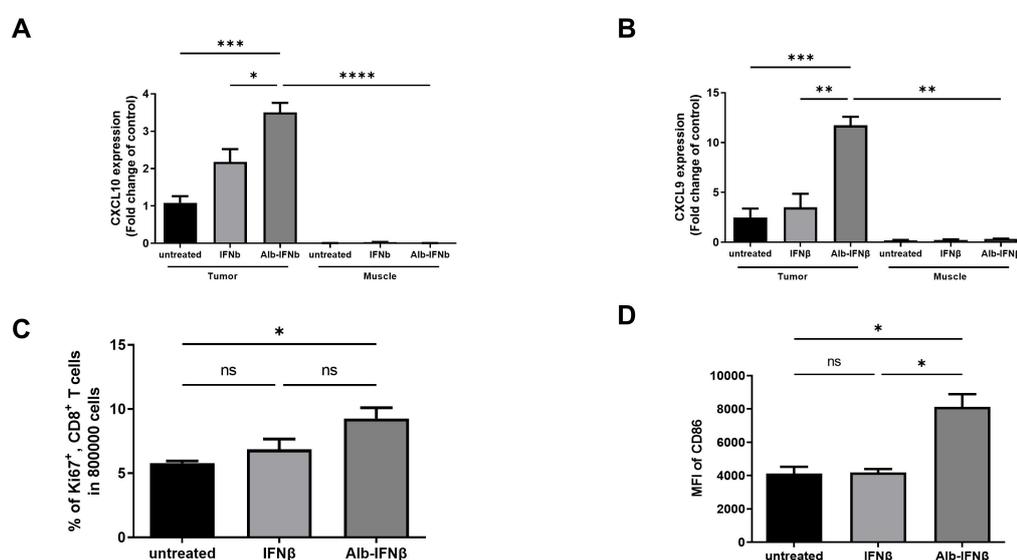
**Supplemental Figure 2. Co-administration of Alb-IFN β and OVA leads to enhanced OVA specific IgG2A antibody response.**

To detect mouse anti-OVA IgG1 or IgG2A antibodies in vaccinated mice, an ELISA assay was performed. Sera from vaccinated and untreated mice were collected on day 25. **(A)** Schematic illustration of the experiment (same as in **Figure 2**). Sera from naïve mice served as negative control. The sera were diluted 1000 \times fold. **(B)** ELISA results of IgG1 antibodies. **(C)** ELISA results of IgG2A antibodies. * p <0.05, ** p <0.01, *** p <0.001, **** p < 0.0001, ns not significant.



Supplemental Figure 3 Characterization of E7-specific CD8+ T cell infiltration in tumors following injection with Alb-IFN β . (A)

Schematic illustration of the experiment. Briefly, C57BL/6 mice (3 per group) were s.c. injected with 5×10^5 TC-1 cells 8 days prior to the adoptive transfer of luciferase-expressing E7-specific T cells. Tumor-bearing mice were i.v. injected with either PBS, IFN β , or Alb-IFN β one day before T cell transfer. On day 0, mice were i.v. adoptive transferred E7-specific T cells in indicated groups, and imaged by IVIS spectrum on days 1 and 4 after T cell transfer. (B) IVIS imaging of the mice. Tumor regions are indicated by red circles. (C) Bar graph displaying the mean fluorescence signals showing T cell infiltration into the tumor regions.



Supplemental Figure 4 Characterization of chemokine levels in tumors and immune cell activity in tumor draining lymph nodes following injection with Alb-IFN β . Tumor-bearing mice ($n = 3$) were i.v. injected with either PBS, IFN β , or Alb-IFN β . Three days after injection, tumors and tdLNs were harvested for quantitative real-time PCR (qRT-PCR) or flow cytometric analyses. **(A)** qRT-PCR data of CXCL10 expression in tumors. **(B)** qRT-PCR data of CXCL9 expression in tumors. **(C-E)** Cells were isolated from tdLNs and stained for proliferative CD8⁺ T cells, or DC activation. **(C)** Bar graph summary of percentage of proliferative CD8⁺ T cells in tdLNs. **(D)** Bar graph summary of mean fluorescence intensity of CD86 in CD11c⁺ and I-A⁺/I-E⁺ DCs. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, **** $p < 0.0001$, ns not significant.