Whole Exome DNA Sequencing

These same tissue blocks were used to extract tumor DNA for sequencing, which determined the presence of somatic mutations in ccRCC tumors. Sequence reads were aligned to the reference human genome with the Burrows-Wheeler Aligner (BWA)¹, and insertion/deletion realignment and quality score recalibration were performed with the Genome Analysis ToolKit (GATK)². Tumor specific mutations were identified with Strelka³ and MuTect⁴, and were annotated to determine genic context (ie, non-synonymous, missense, splicing) using ANNOVAR⁵ and summarized using in-house Perl and R scripts. A summarized outline of the above methods is provided in the Supplementary Material, along with the entire scripts utilized for this analysis.

RNA-seq Immune Cell Expression and IMmotion150 Angiogenesis and T-effector scores

A subset of patients in the IF cohort (n = 92) had tumor samples that had previously undergone bulk RNA sequencing of macro-dissected tumor samples. The TruSeq RNA Exome kit (Illumina) for 50 million 100–base pair paired-end reads was utilized, and RNA sequence reads were aligned to the human reference genome in a splice-aware fashion using Spliced Transcripts Alignment to a Reference (STAR) ⁶, allowing for accurate alignments of sequences across introns. Aligned sequences were assigned to exons using the HTseq package⁷ to generate initial counts by region. Normalization, expression modeling, and difference testing were performed using DESeq2 ⁸. A summarized outline of the above methods is provided in the Supplementary Material, along with the entire scripts utilized for this analysis.

RNA sequencing data was analyzed for cell-type enrichment using the xCell bioinformatic pipeline⁹. xCell uses a compendium of validated gene expression signatures for 64 individual cell-types derived from thousands of expression profiles. Single-sample gene set enrichment analysis scores were adjusted for spillover compensation to generate an adjusted enrichment score for each cell type within

the specimen, which is referred to as the xCell score⁹. xCell scores were generated for each of the 64 cell-types for each tumor specimen. Notably, the tumor samples utilized for RNA sequencing were almost entirely composed of tumor tissue, by volume, and therefore are expected to most closely

resemble the tumor-core zone from the IF analysis. Relevant comparisons were made between xCell

cell-types and corresponding mIF marked cell types.

Angiogenesis and T-effector scores were generated from the RNA-seq data per the

IMmotion150 gene signatures, in a manner according to the original publication¹⁰. Angiogenesis: VEGFA,

KDA, ESM1, PECAM1, ANGPTL4, and CD34. T-effector: CD8A, EOMES, PRF1, IFNG, and CD274.

- 1. Li H, Durbin R. Fast and accurate short read alignment with Burrows-Wheeler transform. *Bioinformatics*. 2009;25(14):1754-1760.
- 2. DePristo MA, Banks E, Poplin R, et al. A framework for variation discovery and genotyping using next-generation DNA sequencing data. *Nat Genet.* 2011;43(5):491-498.
- Saunders CT, Wong WS, Swamy S, Becq J, Murray LJ, Cheetham RK. Strelka: accurate somatic small-variant calling from sequenced tumor-normal sample pairs. *Bioinformatics*. 2012;28(14):1811-1817.
- 4. Cibulskis K, Lawrence MS, Carter SL, et al. Sensitive detection of somatic point mutations in impure and heterogeneous cancer samples. *Nat Biotechnol.* 2013;31(3):213-219.
- 5. Wang K, Li M, Hakonarson H. ANNOVAR: functional annotation of genetic variants from highthroughput sequencing data. *Nucleic Acids Res.* 2010;38(16):e164.
- 6. Dobin A, Davis CA, Schlesinger F, et al. STAR: ultrafast universal RNA-seq aligner. *Bioinformatics* (Oxford, England). 2013;29(1):15-21.
- 7. Anders S, Pyl PT, Huber W. HTSeq--a Python framework to work with high-throughput sequencing data. *Bioinformatics (Oxford, England).* 2015;31(2):166-169.
- 8. Love MI, Huber W, Anders S. Moderated estimation of fold change and dispersion for RNA-seq data with DESeq2. *Genome biology*. 2014;15(12):550.
- 9. Aran D, Hu Z, Butte AJ. xCell: digitally portraying the tissue cellular heterogeneity landscape. *Genome Biol.* 2017;18(1):220.
- 10. McDermott DF, Huseni MA, Atkins MB, et al. Clinical activity and molecular correlates of response to atezolizumab alone or in combination with bevacizumab versus sunitinib in renal cell carcinoma. *Nat Med.* 2018;24(6):749-757.