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Supplementary Materials

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3 **Methods**

4 **Antibodies used and immunohistochemistry**

5 The following primary antibodies were used as following: anti-B7-H4 (clone HPA054200,
6 1:400; Sigma–Aldrich, Ronkonkoma, NY, USA), anti-LAG3 (clone D2G40, 1:100; Cell
7 Signaling Technology, CST, Beverly, Massachusetts), anti-PD-L1 (clone E1L3N, 1:200
8 dilution; CST), anti-B7-H3 (clone D9M2 L, 1:300; CST), anti-IDO1 (clone D5J4E; 1:400;
9 CST), anti-VISTA (clone D1L2G, 1:500; CST), anti-TIM-3 (clone D5D5R, 1:400; CST),
10 anti-ICOS (clone D1K2T, 1:1600; CST), and anti-OX40 (ab119904, 1:1600; Abcam,
11 Cambridge, UK).

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13 **Digital image analysis**

14 The immune checkpoint PD-L1 is expressed by both tumor cells and tumor-associated
15 immune cells and were assessed in combination with both compartments. In contrast, B7-H3
16 is expressed only by tumor cells whereas IDO1, VISTA, TIM-3, ICOS, and OX40 are
17 expressed by tumor-associated immune cells and were assessed only in the tumor cells or
18 tumor-associated immune cells compartment. B7-H4 and LAG3 were all negative in the
19 tissue samples. All cell nuclei in the tumour bulk were identified in haematoxylin channel
20 images after stain deconvolution [1] using a Mask-RCNN-based instance segmentation model.
21 [2] The positive nuclei were further recognized according to the intensities in the DAB
22 channel image, and the density of the positive nuclei in the tumor bulk was calculated. Based

1 on a computational pathology analysis, the expression of the immune checkpoints was
2 digitally defined and quantified as the number of positive expression in combined areas (per
3 mm²), including tumor cells and/or tumor-associated immune cells.

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5 **Random forest for ICS construction**

6 Random forest is a complex nonlinear model and built using the R package
7 “randomForestSRC 3.1.0. As the number of trees increased, the error rate gradually stabilized
8 between 0.42 and 0.44 (**Supplementary Figure S2A**). We used an ensemble of 177 estimator
9 trees and Gini entropy to evaluate the quality of a split at each node of a tree. The variable
10 performance measures were generated based on the Gini index of the random forest model
11 (**Supplementary Figure S2B**). Eventually, the top four immune checkpoints, OX40, B7-H3,
12 ICOS, and TIM-3, were selected for the model (**Supplementary Figure S2B**). The
13 out-of-bag (OOB) Brier and OOB continuous rank probability score (CRPS) were
14 consistently used to evaluate the prediction accuracy of the model. The OOB Brier and OOB
15 CRPS gradually increased as the OS time increased (**Supplementary Figure S2C and S2D**).
16 **Supplementary Figure S2E** shows a decision tree in the random forest. A nonlinear
17 relationship existed between the four selected genes and mortality (**Supplementary Figure**
18 **S2F, S2G, S2H and S2I**). The model was constructed using the discovery set of our FFPE
19 cases (n = 212), and then, we grouped the NB patients into low- and high-risk groups based
20 on the ICS scores using a minimum *P* value.

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1 The model was further validated in the GSE85047 dataset. We used the “predict.rfsrc”
2 function of the R package “randomForestSRC” to confirm the prediction value of the ICS in
3 the validation set.

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5 **References**

- 6 1 Ruifrok AC, Katz RL, Johnston DA. Comparison of quantification of histochemical
7 staining by hue-saturation-intensity (HSI) transformation and color-deconvolution. *Appl*
8 *Immunohistochem Mol Morphol* 2003; 11: 85-91.
- 9 2 Jung H, Lodhi B, Kang J. An automatic nuclei segmentation method based on deep
10 convolutional neural networks for histopathology images. *BMC Biomed Eng* 2019; 1: 24.

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