

MULTIPLEX IMMUNOFLUORESCENCE IMAGING OF ARMORED CAR T CELLS IN PANCREATIC DUCTAL ADENOCARCINOMA MOUSE XENOGRAPTS CONFIRMS POTENT ANTI-TUMOR RESPONSE

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Background Chimeric antigen receptor (CAR) T-cells are a promising modality for the treatment of solid tumors, however their functionality to date has been hampered by the immunosuppressive tumor microenvironment (TME). Armoring CAR T cells to overcome TME may improve clinical outcomes. We established a subcutaneous BxPC3 xenograft model of pancreatic ductal adenocarcinoma (PDAC), that exhibits hallmark characteristics of human PDAC. In this model, CAR T cells regressed tumors; by contrast, armored CAR T cells designed to resist TME, mediated an apparent tumor volume increase. We utilized MACSima™ Imaging Cycling Staining (MICS) system to elucidate the mechanism of apparent tumor volume increase post armored CAR T cell treatment, and the interactions between tumor, stroma and immune cells.

Methods BxPC3 bearing NSG mice were treated with CAR T cells with or without armor, or untransduced T cell control (UTD) at high or low CAR T cell/mouse dose (2×10^6 or 1×10^6) or left untreated. FFPE sections from tumors harvested 7 days post treatment were imaged on MICS by serial immunofluorescence for 29 protein markers and nuclear stain DAPI (ROI of 3mm by 5 mm; $\sim 14.4 \text{ mm}^2$). Multiplex image processing and segmentation, unsupervised UMAP clustering, and target quantitation were performed on MACS® iQ view software.

Results MICS of BxPC3 xenograft tissues revealed PDAC-characteristic morphology, with distinct tumor cell clusters (Panc-cytokeratin, Cytokeratin 7, MSLN, p53) and mucin vacuoles (CA125), surrounded by dense, activated stroma (Vimentin and α SMA). UMAP dimension reduction for 29 markers yielded distinct tumor and stroma clusters in each treatment group. Following CAR treatment, tumor and stroma were infiltrated by CD4+ and CD8+ human T cells. In the high CAR dose groups, there were 14.7% CD8+ tumor-infiltrating lymphocytes (TILs) in CAR alone, and 51.6% for armored CAR. Armored CAR T cells disrupted the tumor/stroma morphology and mediated a dramatic reduction in tumor cells to 1.8%, as compared to 62% for UTD control and 29.3% for CAR T alone. The armored CD8+ TILs were highly proliferative with high abundance of mitochondria: 28% of armored TILs co-expressed Ki67 and TOM22, versus 2.6% in the CAR T alone group, and 0.6% in UTD.

Conclusions Using MICS technology, we therefore delineate the mechanism of action of the armored CAR-T cells in PDAC xenograft model, and rule out tumor progression. Tumor-activated armored CAR T cells manifest high proliferative and metabolic activity, superior tumor penetration and killing.

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