

511 ACTIVITY OF ANTI-CD47 ANTIBODIES IS ENHANCED THROUGH FC OPTIMIZATION

^{1,2}Juan C Osorio*, ²Patrick Smith, ²David A Knorr, ²Jeffrey Ravetch. ¹Memorial Sloan Kettering Cancer Center, New York, NY, USA; ²The Rockefeller University, New York, NY, USA

Background The CD47/SIRPα axis plays a crucial role in cancer immunosurveillance.^{1–2} While anti-CD47 antibodies have shown promise in several preclinical models,^{1–8} results from early phase clinical trials have shown limited clinical benefit,^{9–11} suggesting that the sole blockade of CD47 by the antibody Fab domain might not be sufficient for effective tumor control. A critical question that remains to be answered is whether interactions between the antibody Fc and Fc gamma receptors (FcγRs) also contribute to their antitumor activity.¹² Our study aims to investigate the role of the Fc domain in the in vivo antitumor activity of anti-CD47 antibodies using immunocompetent species-matched models, overcoming limitations of previous studies conducted in immunocompromised models or with interspecies differences between mouse (m) and human (h) CD47, SIRPα and FcγRs.

Methods We modified the Fc domain of the anti-mCD47 antibody MIAP301 to generate antibodies with varying affinity to mFcγRs: 1) MIAP301-mIgG2a Fc, binding to preferentially to activating mFcγRs, 2) MIAP301-mIgG1 Fc, binding to the inhibitory mFcγRIIB, and 3) MIAP301-mIgG1-D265A Fc, which lacks binding to any mFcγRs. We evaluated the antitumor activity of these antibodies in MC38 and B16 tumor models in immunocompetent C57BL/6J mice and mice lacking activating FcγRs.¹⁵ Additionally, we generated a mouse humanized for the expression of hCD47, hSIRPα and hFcγRs by CRISPR/Cas9-mediated gene-targeting strategy, and by backcrossing to our hFcγR mice.¹⁶ We used this model to compare the antitumor activities of the anti-hCD47 antibody Magrolimab (5F9-hIgG4).¹⁴ and an Fc-optimized variant that enhances binding for all the activating hFcγRs (5F9-GAALIE) generated in the lab.

Results The MIAP301-mIgG2a Fc variant led to the most significant reduction in tumor burden when compared to the control or other Fc variants in WT mice in both MC38 and B16 models. This therapeutic effect was abrogated in mice lacking activating FcγRs (figure 1). The CD47/SIRPα/FcγR humanized mice recapitulate the expression profile of CD47 and SIRPα found in human cells (figure 2). Furthermore, increasing dosing concentrations of both 5F9-hIgG4 and 5F9-GAALIE antibodies led to on-target anemia and thrombocytopenia in hCD47/hSIRPα/hFcγR mice, recapitulating results from clinical trials (figure 3). Intratumoral administration of the Fc optimized 5F9-GAALIE results in enhanced long-term antitumor immunity, abscopal antitumor effect, and minimal on-target toxicity when compared to 5F9-hIgG4 or control alone or in combination with PD-1 blockade (figure 4).

Conclusions The antitumor activities of anti-CD47 antibodies require interactions with activating FcγRs, highlighting the

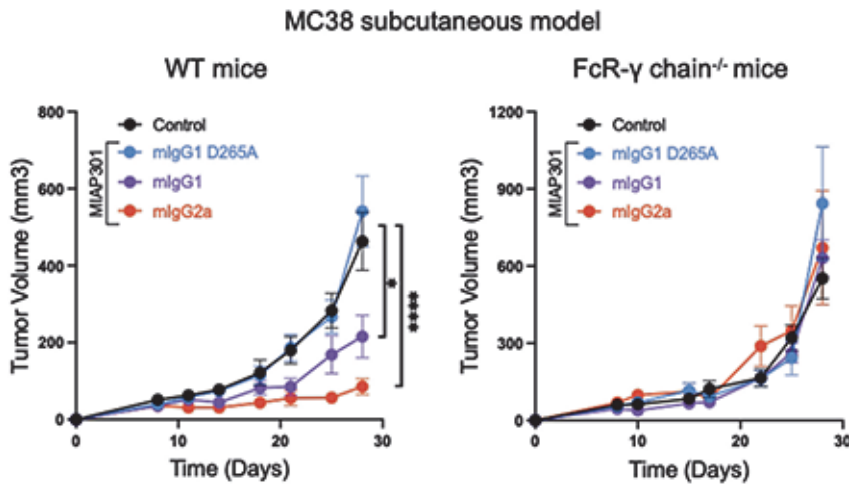
importance of Fc optimization in the development of effective anti-CD47 therapies.

Acknowledgements We thank Maria L. Baez, Alessandra E. Marino, and Carlo M. Sevilla for their excellent technical assistance. We also thank all the members of the J.V.R. Laboratory of Molecular Genetics and Immunology for helping discussions and sharing experiment materials.

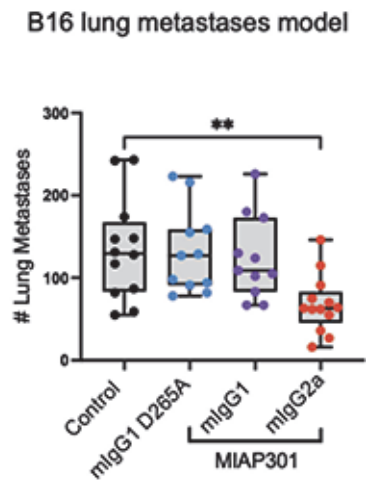
REFERENCES

1. Willingham SB, Volkmer JP, Gentles AJ, et al. The CD47-signal regulatory protein alpha (SIRPα) interaction is a therapeutic target for human solid tumors. *Proc Natl Acad Sci U S A* 2012;**109**:6662–7.
2. Adams S, van der Laan LJ, Vernon-Wilson E, et al. Signal-regulatory protein is selectively expressed by myeloid and neuronal cells. *J Immunol* 1998;**161**:1853–9.
3. Chao MP, Alizadeh AA, Tang C, et al. Anti-CD47 antibody synergizes with rituximab to promote phagocytosis and eradicate non-Hodgkin lymphoma. *Cell* 2010;**142**:699–713.
4. Majeti R, Chao MP, Alizadeh AA, et al. CD47 is an adverse prognostic factor and therapeutic antibody target on human acute myeloid leukemia stem cells. *Cell* 2009;**138**:286–99.
5. Jaiswal S, Jamieson CH, Pang WW, et al. CD47 is upregulated on circulating hematopoietic stem cells and leukemia cells to avoid phagocytosis. *Cell* 2009;**138**:271–85.
6. Weiskopf K, Jahchan NS, Schnorr PJ, et al. CD47-blocking immunotherapies stimulate macrophage-mediated destruction of small-cell lung cancer. *J Clin Invest* 2016;**126**:2610–20.
7. Liu X, Pu Y, Cron K, et al. CD47 blockade triggers T cell-mediated destruction of immunogenic tumors. *Nat Med* 2015;**21**:1209–15.
8. Liu Q, Wen W, Tang L, et al. Inhibition of SIRPα in dendritic cells potentiates potent antitumor immunity. *Oncoimmunology* 2016;**5**:e1183850.
9. Bouwstra R, van Meerten T, Bremer E. CD47-SIRPα blocking-based immunotherapy: Current and prospective therapeutic strategies. *Clin Transl Med* 2022;**12**:e943.
10. Uger R, Johnson L. Blockade of the CD47-SIRPα axis: a promising approach for cancer immunotherapy. *Expert Opin Biol Ther* 2020;**20**:5–8.
11. Jalil AR, Andrechak JC, Discher DE. Macrophage checkpoint blockade: results from initial clinical trials, binding analyses, and CD47-SIRPα structure-function. *Antib Ther* 2020;**3**:80–94.
12. Zhao XW, Matlung HL, Kuijpers TW, van den Berg TK. On the mechanism of CD47 targeting in cancer. *Proc Natl Acad Sci U S A* 2012;**109**:E2843; author reply E4–5.
13. Zhao XW, Kuijpers TW, van den Berg TK. Is targeting of CD47-SIRPα enough for treating hematopoietic malignancy? *Blood* 2012;**119**:4333–4; author reply 4–5.
14. Takai T, Li M, Sylvestre D, Clynes R, Ravetch JV. FcR gamma chain deletion results in pleiotropic effector cell defects. *Cell* 1994;**76**:519–29.
15. Smith P, DiLillo DJ, Bournazos S, Li F, Ravetch JV. Mouse model recapitulating human Fcγ receptor structural and functional diversity. *Proc Natl Acad Sci U S A* 2012;**109**:6181–6.
16. Advani R, Flinn I, Popplewell L, et al. CD47 Blockade by Hu5F9-G4 and Rituximab in Non-Hodgkin's Lymphoma. *N Engl J Med* 2018;**379**:1711–21.

A.

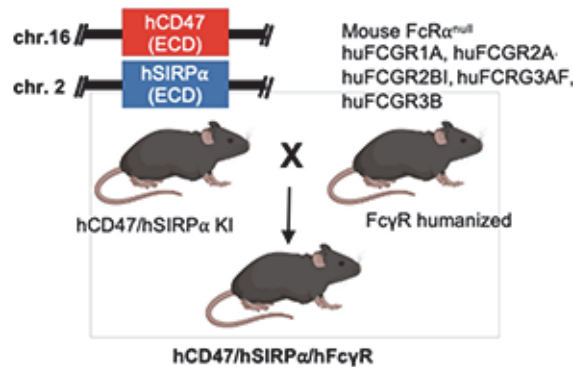


B.

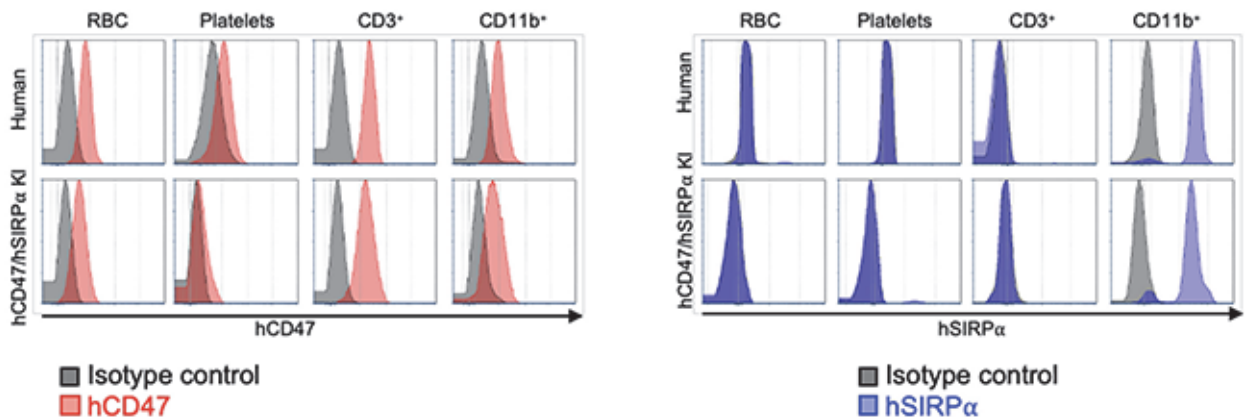


Abstract 511 Figure 1 Engagement of activating FcγRs enhances in vivo antitumor activity of antibodies blocking mouse CD47. (A) Average growth ± SEM of sq. MC38 tumors in WT (right) or FcR-γ chain mice (left), treated with Fc variants of anti-mCD47 ab (MIAP301) or control (50ug IT, d. 8,10,14 and 18). (B) Average of lung metastases (Min to Max) of WT mice inoculated IV with B16 tumor cells, treated with anti-mCD47 ab Fc variants (MIAP301) or control (20 mg/Kg IP, d. 1,4,7 and 11).

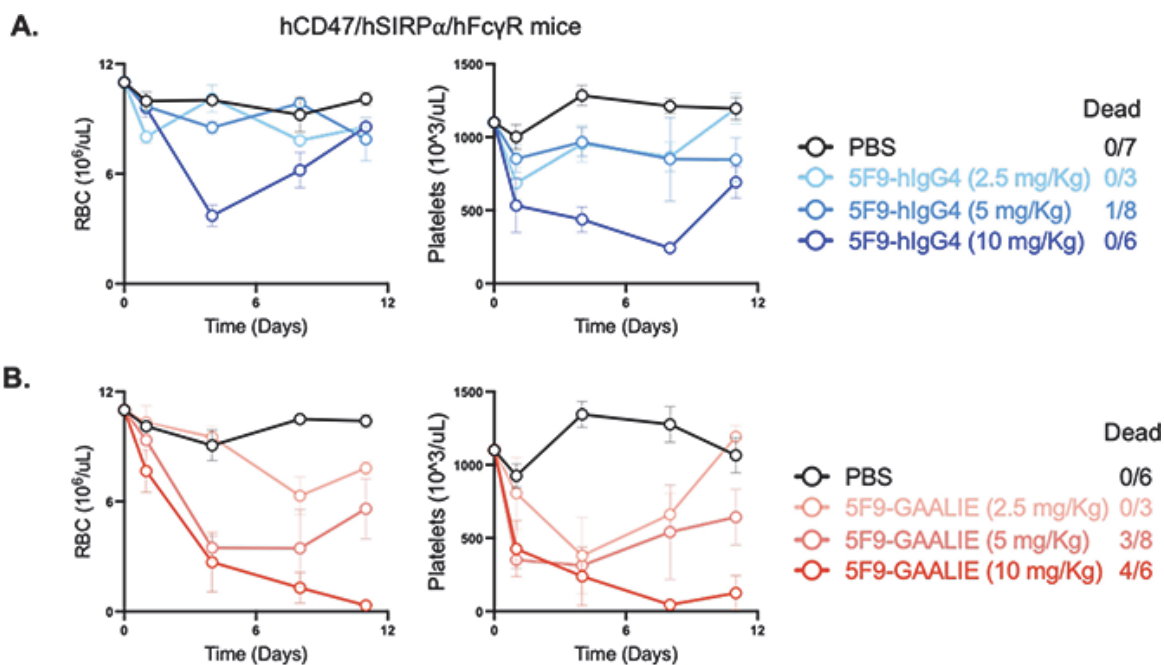
A.



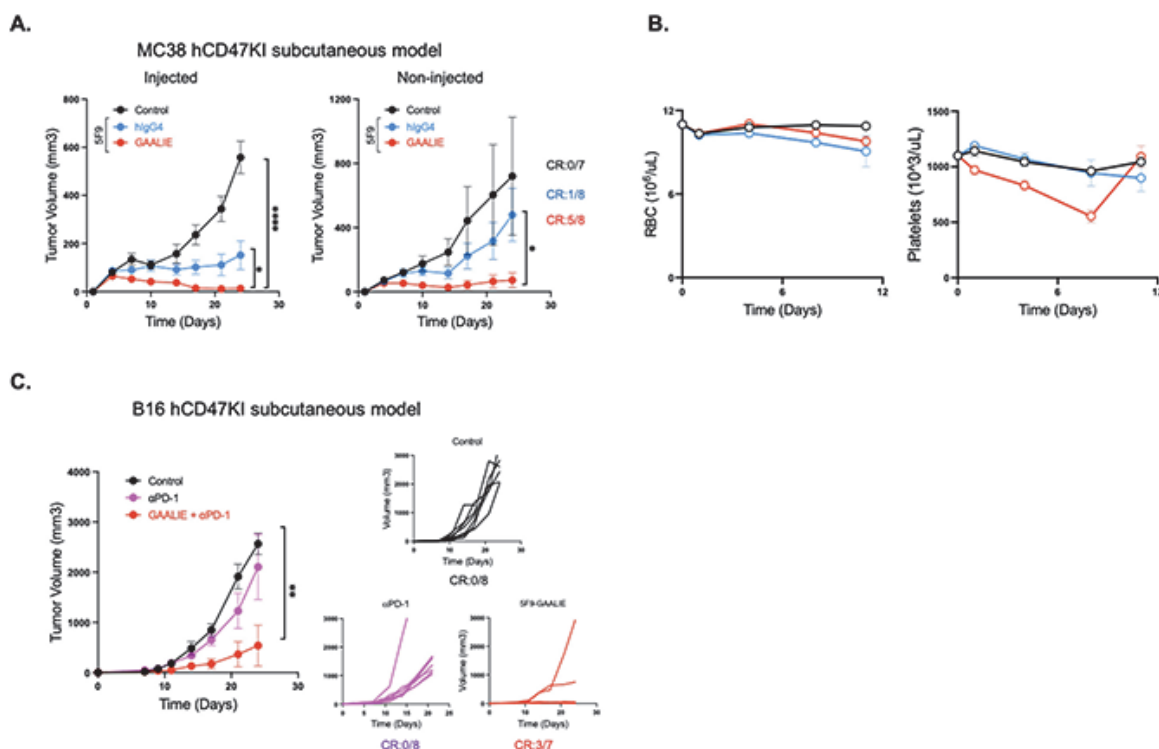
B.



Abstract 511 Figure 2 Generation of humanized mouse model for CD47, SIRPα, and FcγRs. (A) Schematic drawing showing the generation of the hCD47/hSIRPα/hFcγR mice. (B) Flow cytometry analysis of hCD47 and hSIRPα in red blood cells (RBC), platelets, CD3+ and CD11b+ leucocytes isolated from peripheral blood of human (top) and hCD47/hSIRPα KI mice (bottom).



Abstract 511 Figure 3 Humanized mouse recapitulates toxicity profile of fully humanized anti-CD47 antibodies. (A) Peripheral RBC and platelet count from hCD47/hSIRP α /hFc γ R mice after treatment with increasing doses of 5F9-hlgG4 or (B) 5F9-GAALIE ab (2.5, 5, and 10 mg/Kg d. 0, 4, 8 and 11)



Abstract 511 Figure 4 Fc-optimized humanized anti-CD47 Ab promotes effective in vivo antitumor activity (A) Average growth \pm SEM of injected (left) and non-injected contralateral (right) sq. MC38 hCD47 KI tumors in hCD47/hSIRP α /hFc γ R, treated with 5F9-hlgG4, 5F9-GAALIE ab or control (50ug IT, d. 7, 9, and 12). (B) Serial measurements of peripheral RBC and platelets from hCD47/hSIRP α /hFc γ R mice after treatment with 5F9-hlgG4 or 5F9-GAALIE (d. 0 indicates the day when treatment was started). (C) Average growth \pm SEM of sq. B16 hCD47 KI tumors in hCD47/hSIRP α /hFc γ R, treated with 5F9-hlgG4, 5F9-GAALIE ab alone (50ug IT, d. 8,10,14 and 18), or in combination with anti-PD1 ab (RMP1-14, 200 ug IP d. 8, 10, 14 and 18), or control.

<http://dx.doi.org/10.1136/jitc-2023-SITC2023.0511>