

POSTER PRESENTATION

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Single-cell motility and gene expression signature as predictors of the overall survival of act in melanoma patients

Melisa Martinez-Paniagua^{1*}, Cara Haymaker², Jay R Adolacion³, Laszlo Radvanyi⁴, Patrick Hwu², Badrinath Roysam¹, Chantale Bernatchez², Navin Varadarajan¹

From 30th Annual Meeting and Associated Programs of the Society for Immunotherapy of Cancer (SITC 2015) National Harbor, MD, USA. 4-8 November 2015

Background

Adoptive cell therapy (ACT) of tumor infiltrating lymphocytes (TIL) has shown the ability to induce complete regression of stage IV melanoma. Two challenges associated TIL ACT are that (i) T cells are capable of different anti-tumor effector functions including cytotoxicity, cytokine secretion, and homing to target tissues, and currently no methodologies exist that can inform on all of these functions for a given (single) cell, and (ii) the relative contributions of each of these effector functions to the overall anti-tumor effect remains un-quantified.

Methods

Here, we evaluated a set of four infusion products from patients treated with TIL ACT: two Complete Responders (CR) and two patients with Progressive Disease (PD) with high percentages of CD8⁺ T cell population and with established autologous primary tumor cell lines.

Results

Flow cytometry analysis of the tumor reactive TIL showed no difference in the frequency of TIL secreting IFN- γ or degranulating in response to the autologous tumor. Utilizing Timelapse Imaging Microscopy In Nanowell Grids (TIMING), we demonstrate that while the frequencies of individual TIL participating in killing of autologous tumor cells is equivalent across the donors, TIL from CR patients demonstrated a longer duration of conjugation prior to killing the tumor cell (CR 211 \pm 7 min vs PD 131 \pm 5 min). Furthermore, both in the presence and absence of tumors cells, the

motility of the TIL was significantly higher (CR 4.7 \pm 0.2 μ m vs PD 2.9 \pm 0.1 μ m). Multiplexed single-cell gene-expression profiling of the TIL, classified based on activated functional states (CD69/CD107a/IFN- γ) demonstrated that the functional classification was not useful in clustering cells from different donors but rather, clustering of cells was dictated by the donor regardless of function. Lastly, we demonstrate that a core signature of the activated TIL consisting of CD37, CD44, IL7R, CD28, CD27, IFNGR1, TIM-3, MAPK1 and MTOR were up-regulated in CR patients and the levels of TGFB1, BCL2, IL2RG, PDRM1 and IL12RB2 were up-regulated in PD patients.

Conclusions

In summary, our results demonstrate that single cell profiling may be useful in determining not only the transcriptional signatures of TIL with favorable clinical responses but also uncover simple physical attributes like basal motility that might serve as a surrogate marker for activated/functional TIL.

Authors' details

¹University of Houston, Houston, TX, USA. ²UT MD Anderson Cancer Center, Houston, TX, USA. ³University of Houston, University of the Philippines, Houston, TX, USA. ⁴Moffit Cancer Center, Tampa, FL, USA.

Published: 4 November 2015

doi:10.1186/2051-1426-3-S2-P33

Cite this article as: Martinez-Paniagua *et al*: Single-cell motility and gene expression signature as predictors of the overall survival of act in melanoma patients. *Journal for ImmunoTherapy of Cancer* 2015 **3**(Suppl 2): P33

¹University of Houston, Houston, TX, USA Full list of author information is available at the end of the article

