

Neuropilin-1: a checkpoint target with unique implications for cancer immunology and immunotherapy

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ABSTRACT

Checkpoint blockade immunotherapy established a new paradigm in cancer treatment; for certain patients curative treatment requires immune reinvigoration. Despite this monumental advance, only 20%-30% of patients achieve an objective response to standard of care immunotherapy. necessitating the consideration of alternative targets. Optimal strategies will not only stimulate CD8+ T cells, but concomitantly modulate immunosuppressive cells in the tumor microenvironment (TME), most notably regulatory T cells (T_{reg} cells). In this context, the immunoregulatory receptor Neuropilin-1 (NRP1) is garnering renewed attention as it reinforces intratumoral T_{req} cell function amidst inflammation in the TME. Loss of NRP1 on T_{ren} cells in mouse models restores antitumor immunity without sacrificing peripheral tolerance. Enrichment of NRP1+ T_{req} cells is observed in patients across multiple malignancies with cancer, both intratumorally and in peripheral sites. Thus, targeting NRP1 may safely undermine intratumoral T_{rea} cell fitness, permitting enhanced inflammatory responses with existing immunotherapies. Furthermore, NRP1 has been recently found to modulate tumor-specific CD8+ T cell responses. Emerging data suggest that NRP1 restricts CD8+ T cell reinvigoration in response to checkpoint inhibitors, and more importantly, acts as a barrier to the long-term durability of CD8⁺ T cell-mediated tumor immunosurveillance. These novel and distinct regulatory mechanisms present an exciting therapeutic opportunity. This review will discuss the growing literature on NRP1-mediated immune modulation which provides a strong rationale for categorizing NRP1 as both a key checkpoint in the TME as well as an immunotherapeutic target with promise either alone or in combination with current standard of care therapeutic regimens.

INTRODUCTION

Immuno-oncology opportunities beyond PD1 and CTLA4

The unprecedented success of checkpoint blockade immunotherapy has brought immune reactivation to the forefront of next-generation cancer therapeutic strategies, demonstrating that reinvigoration and/or prevention from immune exhaustion of CD8⁺ T cells is essential for a clinical response. In particular, the first checkpoint blockade immunotherapies offered as standard of

care for patients with cancer, targeting either cytotoxic T-lymphocyte associated protein 4 (CTLA4) via ipilimumab¹⁻⁵ or the programmed cell death protein 1 (PD1) pathway via nivolumab, pembrolizumab or atezolizumab,⁶⁻¹⁰ achieved overall response rates of up to 40%.

Despite these successes, the majority of patients fail to achieve objective clinical response.⁹ 10 Numerous explanations for immunotherapy resistance have been posited, including low inhibitory receptor (IR) ligand expression, 11 low prevalence of immunogenic neoepitopes, 12-14 high tumor burden to inflammatory response ratio, ¹⁵ and immune exclusion. 16 Consequently, significant efforts in the field have centered on evaluating combinatorial strategies. In fact, combination of ipilimumab and nivolumab in advanced melanoma has demonstrated modest clinical benefit over nivolumab alone. extending median overall survival beyond 3 years. 1718 Continued evaluation of new targets and therapeutic combinations with standard of care immunotherapies is required to maximize the proportion of patients who mount a durable response.

Beyond CLTA4 and PD1, clinical investigation of additional IRs holds promise for bridging the gap in response rates. Major emerging targets include lymphocyte activation gene 3 (LAG3), 19 20 T cell immunoreceptor with Ig and ITIM domains (TIGIT), 21 22 and T-cell immunoglobulin and mucin-domain containing 3 (TIM3),²³ among others. Although these receptors regulate antitumor immunity through different signaling pathways, the outcome on CD8⁺ T cell function is comparable and primarily mediated during the effector response.²⁴ Thus, immunomodulatory receptors that impact unique T cell functions, subtypes, or differentiation stages may pose significant combinatorial therapeutic advantages.



The broad mechanism of action for checkpoint blockade therapy is reversal of intrinsic T cell inhibition via IRs. Such agents interfere with ligand binding to IRs expressed on CD8+ T cells in the tumor microenvironment (TME).²⁵ Under normal physiology, IRs tune the immune response to pathogenic challenge by directly attenuating the T cell receptor (TCR) signaling cascade. As a consequence, IR expression limits off-target activation, minimizes host tissue damage, facilitates immune resolution, and enhances T cell memory formation.²⁵ However in cancer, chronic antigen exposure sustains prolonged expression of multiple IRs, leading to extreme dysfunction marked by reduced effector function, that is, cytokine production and T cell proliferation in response to stimulation. ²⁶ Thus, IR blockade in patients with cancer leads to improved antitumor immunity.

Furthermore, the cell intrinsic impact of IR blockade on the function of immunosuppressive cells in the TME, most notably T $_{\rm reg}$ cells, warrants additional consideration. 27 Whereas CTLA4 28 , TIGIT, $^{29\,30}$ and TIM3 $^{31\,32}$ mark more suppressive T_{reg} cells, the roles of PD1 and LAG3 are equivocal and may be context-specific. 33-36 In gastric cancer, a recent paper demonstrated that anti-PD1 therapy reinvigorates PD1⁺ intratumoral T_{reg} cells in patients who have hyperprogression under treatment.³⁴ Therefore, it is critical for the field to consider the balance between promoting CD8⁺ T cell activation and enforcing T_{reg} cell suppression as a consequence of checkpoint blockade administration. Careful selection of therapeutic targets with this perspective in mind may elucidate superior clinical strategies for individual malignancies and perhaps even individual patients based on the prevalence of intratumoral T_{reg} cells compared with CD8⁺ T cells.

Recently Neuropilin-1 (NRP1) has begun to garner significant interest in the immune-oncology field as a novel target for its potent dual function: augmentation of T_{reg} cell suppression and restriction of durable CD8⁺ T cell responses. Though initially considered a T_{reg} cell marker,³⁷ new research indicates that NRP1 is not only required for intrinsic T_{reg} cell stability in the TME, ³⁸ ³⁹ but it also substantially inhibits CD8⁺ T cell antitumor function. 40 It is also highly expressed on myeloid subpopulations, but its function in this context is less well characterized. These preclinical observations justify further investigation of NRP1 antagonism in combination with established immunotherapies. This review will highlight the initial findings for immunomodulatory function via NRP1 and discuss emerging findings that NRP1 functions as a key immune modulator in the TME with attractive therapeutic opportunity.

Basic NRP1 biology

Neuropilins (NRPs) are single-pass transmembrane, non-tyrosine kinase surface glycoproteins found in all vertebrates and are highly conserved across species. Two homologous NRP isoforms are known to exist, namely NRP1 and NRP2, encoded by distinct *neuropilin* genes (*Nrp1* and *Nrp2*) which arose due to a gene duplication

event. He both NRPs were originally discovered as neuronal adhesion molecules participating in Semaphorin-mediated axonal guidance. They were later found to be fundamentally involved in vascular biology, with NRP1 required for normal embryonic vascular development and NRP2 involved in the formation of small lymphatic vessels and capillaries. Usualis sover the past decade have revealed NRPs are multifunctional proteins participating in a variety of biological processes beyond the nervous and vascular development, with NRP1 having a major role in immunity and tumorigenesis.

Protein structure and binding ligands

The NRP1 protein consists of a long N-terminal extracellular domain, followed by a transmembrane region and a very short cytosolic tail of 43–44 amino acids (figure 1). The large extracellular domain is comprised of three major segments, namely, two CUB (complement C1r/C1s, Uegf, bone morphogenetic protein 1) domains (denoted 'a1/ a2') and two coagulation factor V/VIII domains (denoted FV/VIII or 'b1/b2'), followed by a MAM (homologous to meprin protease, A5 antigen, receptor tyrosine phosphatase µ and K) domain (denoted 'c'). The CUB domains are required for the binding of the Semaphorin group of ligands, while the FV/VIII domains are responsible for vascular endothelial growth factor (VEGF) binding as well as the docking site for different ligands (such as Semaphorins and VEGFs). 47 48 The MAM domain is known to mediate homodimerization or heterodimerization of the receptors. Although it was initially thought that NRP1 was a non-signaling receptor due to its very short cytosolic tail, a conserved PDZ (PSD-95/Dlg/ZO-1 homology) domain-binding motif (SEA) at the c-terminus was later found to bind to the GAIP Interacting Protein C-terminus/synectin, which mediates intracellular signaling⁴⁹ and receptor internalization.⁵⁰

NRP1 is capable of binding to a broad repertoire of ligands, accounting for its diverse biological functions. First identified as the coreceptor for the secreted class III Semaphorins, such as Semaphorin 3A (Sema3A), ⁵¹ NRP1 was later shown to bind to a variety of growth factors, most notably VEGF₁₆₅, as well as others including transforming growth factor beta, ⁵² ⁵³ platelet-derived growth factors C and D, ⁵⁴ ⁵⁵ and c-Met. ⁵⁶ Interestingly, recent studies have revealed that NRP1 also binds to extracellular microRNA/AGO2 complexes and facilitates their internalization, ⁵⁷ adding another mechanism by which NRP1 modulates cellular functions.

Physiological consequence of genetic deletion

Nrp1-null mice are embryonic lethal due to severe developmental defects, 43 58 mainly manifested in the neuronal and cardiovascular systems. While the signals conveyed by Semaphorin and VEGF through NRP1 are responsible for the neuronal and vascular defects, respectively, both signals are required for normal heart development. 59 Studies utilizing advanced genetic tools including a Nrp1 variant knock-in mouse strain (Nrp1-sema) in which the

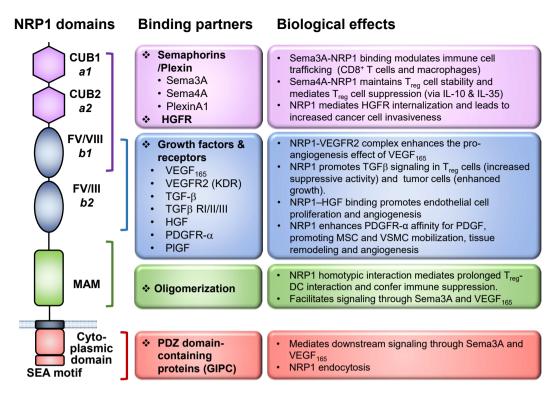


Figure 1 NRP1 structure and ligands. Graphical depiction of NRP1 structure to scale with key functional domains annotated. The associated ligands for each domain are listed and grouped into classes, whereas the A1 and A2 domains primarily facilitate Sema interactions, the B1 and B2 domains contribute to NRP1 binding a number of growth factors. The MAM domain uniquely functions to oligomerize NRP1 in *trans*. The cytoplasmic tail contains a C-terminal SEA motif which serves as a docking site for PDZ domain-containing proteins. Lastly, the key biological functions of the interactions referenced are listed, with specific emphasis on known immune-related and cancer cell biology functions. DC, dendritic cell; NRP-1, Neuropilin-1; Sema, Semaphorin; Sema-3A, Semaphorin-3A; TGFβ, transforming growth factor beta; VEGF, vascular endothelial growth factor; PDGFR, platelet-derived growth factor receptor; PIGF, placenta growth factor; VSMC, vascular smooth muscle cell; MSC, mesenchymal stem cell; HGF, hepatocyte growth factor; GIPC, GAIP Interacting Protein C-terminus.

Semaphorin binding was disrupted without affecting the VEGF binding, as well as an endothelial cell conditional *Nrp1* knockout (*Tie2*^{Cre}*Nrp1*^{L/L}) strain,⁵⁹ indicate that NRP1 acts as a receptor 'hub' for sorting signals from diverse ligands. This canonical mode of action is key to determining the context and cell-type specific function of NRP1.

NRP1 in the immune system

Myeloid cells

Whereas NRP1 is ubiquitously expressed across multiple tissue and cell types within the central nervous and vasculature systems in both human and mouse, NRP1 expression in the immune system is more restricted and regulated. NRP1 was identified as a marker for human dendritic cells (DCs), known as the blood DC antigen 4 (BDCA4, or CD304), which is expressed in all plasmacytoid DCs (pDCs). Known as the 'professional' type I interferon (IFN) producing cell type, human pDCs showed reduced IFN- α release on virus infection when treated with anti-NRP1 antibody, suggesting an immunoregulatory role of NRP1 in the context of antivirus immunity. Other antigen-presenting cells (APCs) that express NRP1 include monocytes and macrophages, in particular

several types of tissue-resident macrophages, such as microglia and adipose tissue macrophages (ATMs). 62-65 Monocyte/macrophage NRP1 expression is generally considered proangiogenic and anti-inflammatory, thereby contributing to tissue remodeling and wound healing. 66 In a recent report, the NRP1+ ATMs were found to confer protection against obesity and metabolic syndrome by maintaining glucose homeostasis. 63

T cells

Within the adaptive immune compartment, NRP1 has been known as a marker for murine thymically derived T_{reg} (tT_{reg}) cells, although this does not extend to human T_{reg} cells (to be discussed later). Conversely, NRP1 is expressed at very low levels on resting CD4⁺ helper T cells and CD8⁺ T cells. In a mouse model resistant to the pathogenesis of experimental autoimmune encephalitis (EAE), NRP1 is upregulated in the EAE-tolerant CD4⁺ T cells (both the Foxp3⁺ and Foxp3⁻ compartments) and functionally contributes to their suppressive phenotype. Other T cell subsets where NRP1 expression is reported include the T follicular helper cells ⁶⁸ and IL-17 expressing invariant natural killer T cells cells. Lastly, NRP1 is constitutively expressed on human thymic epithelial cells

(TECs) and upregulated on immature thymocytes during TEC-thymocyte contact, which is subsequently blocked by Sema3A to direct thymocyte migration.⁷⁰

Myeloid-T cell interactions

NRP1 is also expressed by conventional DCs isolated from human peripheral blood, where it may promote early T cell priming by mediating the formation of immunological synapse between DCs and T cells via homotypic interactions. 71 As T_{reg} cells preferentially express NRP1, T_{reg} cells gain advantage over conventional CD4⁺ T cells by engaging DCs longer at immune synapses through NRP1-NRP1 interactions in the absence of inflammation or foreign antigen exposure, a mechanism that maintains immune tolerance at homeostatic state.⁷² Moreover, in the context of transplantation, intercellular membrane protein transfer from APCs to T cells, called trogocytosis, sensitizes CD4⁺ T cells to inhibitory signals through several known NRP1 ligands, such as Sema3A and VEGF.⁷³ Furthermore, VEGF, known as an immunosuppressive cytokine, can inhibit lipopolysaccharide-induced DC maturation in a NRP1-dependent manner.⁷⁴

To summarize, it is evident that NRP1 mediates important immunoregulatory functions including self-tolerance and immune homeostasis, resembling the activity of some known IRs. However, NRP1 is distinguished from classical IRs by exerting such impacts on a variety of cell types, in particular immunosuppressive cells. These features underscore why examining the function of NRP1 in the context of cancer may be a key next step for improving immunotherapy.

NRP1 expression and function in the mouse and human TME

The established association between NRP1 and cancer is supported by three key observations: (1) elevated NRP1 expression was reported in malignant cells of multiple human cancer types; ⁴⁵ (2) NRP1 is abundantly expressed within the TME, including both the stromal and immune compartments ⁷⁵; and (3) NRP1 expression is generally associated with poor clinical prognosis. ⁷⁶ From the perspective of tumor cells, NRP1 supports tumor cell growth via multiple axes, including cell survival, neoangiogenesis, and metastasis. ⁴⁵ ⁷⁶ ⁷⁷

At the tumor–immune interface, NRP1 promotes immune evasion by orchestrating multiple inhibitory processes within the immune compartments of the TME (figure 2). On the one hand, the physiological roles of NRP1 in DCs and macrophages, which contribute to an anti-inflammatory phenotype of these cells, are 'hijacked' by tumors to promote tumor angiogenesis and tumor-associated immunosuppression. On the other hand, NRP1 has a direct impact on adaptive antitumor immunity, which is achieved by impinging on both cell-extrinsic and cell-intrinsic inhibitory pathways regulating intratumoral T_{reg} cells and CD8⁺ T cells. This review will focus on the latter topic, in particular observations substantiated by data derived from human patients and preclinical mouse models in order to

better contextualize the importance of NRP1 on these two lymphocytic populations that are heavily regulated by immune-based therapies.

NRP1 IN T_{reg} cells: the Guardian of Intratumoral T_{reg} cell stability

NRP1 expression in human tissues and immune cell subsets mirrors that found in mouse models. The most notable exception is T_{reg} cells. NRP1 was initially considered a marker for tT_{reg} (or natural) cells in mice, ³⁷ in part due to its apparent coexpression with Helios. ^{78–80} Indeed, the murine Nrp1 gene is a direct target of Foxp3-mediated transcriptional regulation, demonstrated by ectopic expression and chromatin immunoprecipitation experiments. 81-83 However, subsequent investigation revealed that NRP1 is not expressed by human peripheral T_{reg} cells in blood or lymph nodes. 84 Instead, healthy donor T_{reg} cells upregulate NRP1 on in vitro activation, 84 indicating that immune processes may regulate NRP1 expression in vivo. Though NRP1 regulation may have species-specific determinants, results discussed below suggest that its impact on T_{reg} cell phenotype and function remains conserved.

In the context of cancer, $T_{_{\rm reg}}$ cell expression of NRP1 potentiates immune suppression through at least two parallel pathways: T_{reg} cell recruitment to the tumor by acting as a coreceptor for VEGF,85 and maintaining tumor-specific T_{reg 38 39} cell stability via Semaphorin-4A (Sema4a) ligation. Initial analysis of the effects of T cell-restricted Nrp1 deletion in tumors utilized $Cd4^{Cre}Nrp1^{L/L}$ mice. Though bulk Nrp1-deficient T_{reg} cells retained equal in vitro suppressive function, the proportion and function of intratumoral CD8⁺ T cells was dramatically increased. This led to enhanced tumorfree survival and reduced tumor growth kinetics in both implantable and spontaneous tumor models. As NRP1 is known to act as a VEGFR2 coreceptor in the context of angiogenesis, $^{86\,87}$ it was demonstrated that NRP1 $^{^+}$ $T_{_{\rm reg}}$ cells migrated along a VEGF gradient in vitro. Additionally, in vivo reduction in tumor growth in $\mathit{Cd4}^{\mathsf{Cre}}\mathit{Nrp1}^{\mathsf{L/L}}$ mice could be recapitulated in wildtype mice by administering Vegf^{/-} fibrosarcoma tumors subcutaneously in contrast to VEGF replete tumors. These observations highlight the importance of T_{reg} cell chemotaxis in response to VEGF as a critical component of T_{reg} cellestablished immune suppression in tumor models.

A complimentary finding from our group revealed that beyond cell trafficking, NRP1 was essential for maintaining intratumoral T_{reg} cell function and phenotype. ³⁸ Disruption of the NRP1 pathway, either by antibody blockade or by T_{reg} cell-specific genetic deletion (via $Foxp3^{Cre}Nrp1^{L/L}$), impeded intratumoral T_{reg} cell suppressive function, thereby restoring antitumor immunity, without permitting off-target peripheral autoimmunity. Compared with their $Nrp1^{t/+}$ wild-type counterparts, $Nrp1^{-/-}$ intratumoral T_{reg} cells had decreased Bcl2 and increased active caspase-3, indicating dysregulation of survival pathways. Furthermore,

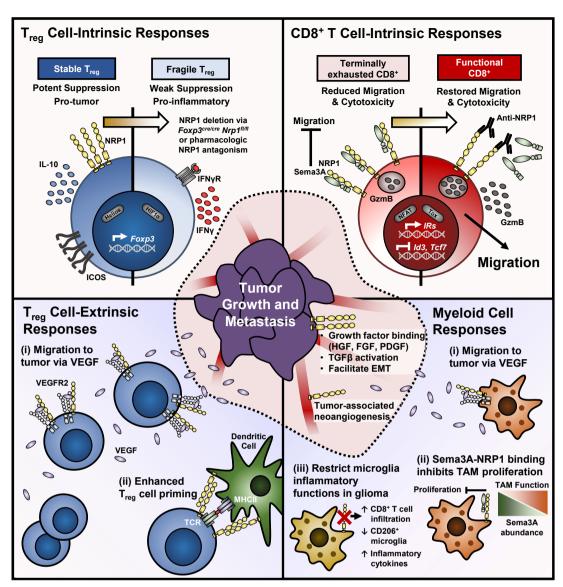


Figure 2 NRP1 contributes to central immune inhibitory mechanisms on T cells in the TME. (Top left) NRP1 marks stable intratumoral T_{reg} cells. Genetic deletion of NRP1 in mouse T_{reg} cells restore antitumor immunity by cell-intrinsically restraining T_{reg} cell function. NRP1-deficient T_{reg} cells are reprogrammed to a proinflammatory phonotype marked by interferon-γ (IFNγ) production. (Top right) Terminally exhausted CD8+ T cells express high levels of NRP1 with a module of additional inhibitory receptors. Sema3A binding to NRP1 on CD8+ T cells inhibits migration to tumors and cytotoxic function, both of which are restored with NRP1 blockade. (Bottom left) Beyond T_{reg} cell fragility (i) NRP1 enhances T_{reg} cell migration and retention in tumors secreting vascular endothelial growth factor (VEGF) by acting as a coreceptor with VEGFR2. (II) Furthermore, homotypic interactions between NRP1 on T_{reg} cells and DCs facilitate enhanced T_{reg} cell priming in response to low antigen availability and maintained peripheral tolerance. (Bottom right) NRP1 expression by tumor-associated monocytes, macrophages, and microglia (collectively TAM) tunes their inflammatory response. (i) NRP1 expression enhances chemotaxis to tumor-derived VEGF. (II) Sema3A complexes with NRP1 on TAM to inhibit their proliferation—even favoring antitumor M1 macrophage function. (III) NRP1 deletion on microglia restore antitumor immunity, marked by increased CD8+ T cell infiltration, decreased CD206 expression, and increased proinflammatory cytokine production. (Center) NRP1 has many known functions in promoting tumor growth and invasiveness, both when expressed on transformed cells or tumor-associated blood vessels. DCs, dendritic cells; NRP-1, Neuropilin-1; Sema-3A, Semaphorin-3A; TME, tumor microenvironment; T_{reg} cells, regulatory T cells; VEGF, vascular endothelial growth factor.

 $\mathit{NrpT}^{/-}$ intratumoral T_{reg} cells downregulated activation markers (including Helios, IL-10, ICOS, CD73) and adopted characteristic T helper lineage markers (such as Tbet, CXCR3, IRF-4, and RORyt). Our group showed that NRP1 localized to the immunologic synapse during T cell activation to restrain Akt (aka protein kinase B or PKB)

activity through phosphatase and tensin homolog (PTEN), thereby relieving Akt-mediated antagonism on Foxo1/3 to stabilize $\rm T_{reg}$ cell function. Follow-up work demonstrated that although intratumoral Nrp1-deficient $\rm T_{reg}$ cells retain Foxp3 expression, their loss of suppressive function is potentiated by adoption of a pro-inflammatory phenotype, marked by

increased production of IFN γ . Of further interest, IFN γ production by $Nrp\Gamma^{-}$ T_{reg} cells initiated dysfunction of neighboring NRP1 $^+$ T_{reg} cells, in a process termed infectious fragility. The clinical significance of this finding was demonstrated by the requirement for T_{reg} cell sensitivity to IFN γ in order to mediate tumor clearance on anti-PD1 immunotherapy in the MC38 tumor model. Indeed, mice harboring T_{reg} cells that lack the IFN γ receptor were insensitive to anti-PD1. These findings provide substantial clinical rationale for further investigation of NRP1 antagonism as a therapeutic agent. Sense of NRP1 antagonism as a therapeutic agent blockade mirrors the effects of T_{reg} cell-restricted NRP1 genetic deletion in murine models, both in vitro and in vivo.

Although T_{reg} cell expression of NRP1 differs significantly between mice and humans, numerous studies have reported increased NRP1 $^+$ T $_{\rm reg}$ cells in patients with cancer. This T $_{\rm reg}$ cell phenotype was anecdotally reported in peripheral blood of pancreatic adenocarcinoma and liver metastases from colorectal cancer.⁹¹ In chronic lymphocytic leukemia, elevated NRP1 on B cells and $T_{re\sigma}$ cells was observed in patient blood that is decreased following thalidomide treatment, linking antiangiogenic therapies to alleviation of antitumor suppression. 92 It has also been reported that there is an enrichment of NRP1 $^{+}$ T_{reg} cells in tumor-draining lymph nodes (TDLN) of patients with cervical cancer, particularly TDLN where tumor metastases are established. 93 94 Furthermore, human NRP1 $^+$ T $_{reg}$ cells are more functionally suppressive and both FOXP3 and glucocorticoidinduced TNFR-related protein (GITR) expression are increased in NRP1 $^{\!\scriptscriptstyle +}$ $T_{_{\rm reg}}$ cells. This observation matches recent findings from our group. Interestingly, NRP1 antagonism at the VEGF-binding domain reduced human intratumoral T_{reg} cell suppression by approximately 20% by inducing NRP1 protein internalization. 90 For human T_{reg} cells, NRP1 expression may also reflect exposure to ongoing inflammation as enrichment of NRP1+ T_{reg} cells has also been reported in the synovial fluid of patients with rheumatoid arthritis. 96 It is also worth noting that the frequency of NRP1⁺ T_{reg} cells decreases following therapeutic intervention, both pharmacologic and surgical. 91-93 Lastly, our group reported elevated NRP1⁺ T_{reg} cells in the peripheral blood and tumors of treatment-naive melanoma and patients with head and neck squamous cell carcinoma. 39 The prevalence of intratumoral NRP1+ T_{reg} cells negatively correlated with disease-free survival in both cohorts. Together, these findings support the notion that NRP1 $^+$ T $_{\rm reg}$ cells are functionally enhanced in cancer. Furthermore, increased NRP1 $^+$ $T_{_{\rm reg}}$ cells in peripheral blood was observed in patients with cancer. $^{39~92}$ This finding is unique among IRs, which tend to have minimal expression in blood samples and may suggest that NRP1 expression on circulating T_{reg} cells could serve as a pretreatment or on-treatment prognostic biomarker for clinical outcomes.⁹⁷

NRP1 ON CD8⁺ T CELLS: THE LINK BETWEEN T CELL DYSFUNCTION AND ABERRANT TUMOR-SPECIFIC CD8⁺ T CELL MEMORY

In contrast to the constitutive expression of NRP1 on thymically derived murine T_{reg} cells, its expression on naive CD8⁺ T cells is undetectable (both mouse and human) and is only induced on T cell activation. Transcriptional upregulation of NRP1 by CD8+ T cells was first documented in an early molecular and functional profiling of CD8⁺ T cell differentiation using LCMV-specific (P14) TCR transgenic T cells, 98 where Nrp1 transcription peaked at the effector CD8⁺ T cells and the effector-tomemory transition phases. Nrp1 upregulation coincided with a group of genes encoding proteins involved in T cell migration and adhesion, such as CCR5, CD44, and p-selectin glycoprotein ligand 1 (PSGL-1). This raises the question of whether NRP1 also modulates CD8⁺ T cell migration, as it does in neuronal or endothelial cells. Consistent with this finding, our group observed upregulation of NRP1 expression (both gene transcription and protein level) on polyclonal intratumoral effector CD8⁺ T cells as well as activated tumor-antigen specific CD8⁺ T cells. Therefore, TCR engagement seems to be necessary to drive NRP1 expression in CD8⁺ T cells, a feature shared by most known T cell coreceptors. However, despite the observed upregulation, the functional role for NRP1 during the early priming of CD8⁺ T cells is unknown.

Some early observations have suggested NRP1 may be an IR-like molecule in CD8⁺ T cells. It was first found highly induced on a subset of immunosuppressive intestinal CD8⁺ T cells (the Foxp3⁺ CD8⁺ T_{reg} cells), along with molecules known to be associated with $\tilde{C}D4^{+}T_{re\sigma}$ cells such as PD1 and CD103. These CD8 $^+$ T $_{\rm reg}$ cells may contribute to maintaining intestinal homeostasis in vivo by downmodulating effector functions of T cells. 99 Consistently, in a later report using Gag-specific (TCRGag) CD8+T cells to understand cell intrinsic mechanisms regulating CD8+ T cell tolerance versus immunity, 100 it was determined that NRP1 was preferentially expressed on tolerant, selfreactive CD8⁺ T cells, mirroring PD1, LAG3 and CTLA4, although NRP1 was dispensable for tolerance. Additional evidence suggested that NRP1 may have a role in T cell dysfunction, a term used to describe T cells that are anergized or exhausted as a result of lacking costimulation or persistent antigen exposure. T cell dysfunction is phenotypically characterized by high IR coexpression and reduced effector marker expression, 101 and it was found that NRP1 belongs to a core transcriptional signature of 174 genes shared by all aforementioned T cell dysfunctional states. 102

Indeed, a recent report indicated that CD8⁺ T cell NRP1 expression in mice and humans is exclusive to a subset of intratumoral CD8⁺ T cells marked by high expression of PD1, whereas NRP1 is minimally detected on the PD1^{neg} intratumoral CD8⁺ T cells.⁴⁰ Compared with the NRP1⁻PD1⁻ and NRP1⁻PD1⁺ counterparts, the NRP1⁺PD1^{hi} cells exhibited higher expression of classical IRs (eg, LAG3, TIM3, TIGIT, 2B4), as well as markers

related to cell proliferation (eg. Ki67) and cytotoxicity (eg, Granzyme B). They also express higher levels of exhaustion-associated transcription factors, such as NFATc1, TOX, Blimp1 and IRF4, but decreased levels of genes associated with cell survival (Bcl2) and memory/ exhaustion precursor cells (TCF1). This is highly reminiscent of 'terminally exhausted' CD8⁺ T cells that have been defined in both chronic viral infection and tumor models. 103 Importantly, NRP1 was functionally involved in the terminal exhaustion of intratumoral CD8+ T cells, rather than a mere consequence of this dysfunctional state. Specifically, the CD8⁺ T cells recovered from B16F10 tumors treated with a neutralizing anti-NRP1 antibody, expressed higher levels of Perforin and Granzyme B, the key molecules mediating the cytotoxic activity of CD8⁺ T cells, and exhibited enhanced cell killing towards autologous tumor cells ex vivo. 40 In the same study, in vivo NRP1 blockade led to reduced tumor growth, which further synergized with PD1 blockade, although such synergy was not observed in terms of enhanced cytotoxic activity ex vivo. The differences between in vivo and ex vivo settings may be due to enhanced tumor recruitment of recently activated CD8+ T cells with anti-NRP1 treatment through blockade of Sema3A-NRP1 axis. Importantly, there might also be the contribution from blocking NRP1 on cell types (eg, T_{reg} cells) other than CD8⁺ T cells under an in vivo anti-NRP1 regimen. Indeed, the latter seems to be supported by our study utilizing a CD8+ T cell-specific Nrp1-deficient mouse strain (E81^{Cre}Nrp1^{L/L}) in which primary tumor growth was similar to wild-type counterparts. Nevertheless, with the E81^{Cre}Nrp1^{L/L} mice we also observed clear synergy between genetic ablation of Nrp1 and PD1 blockade in the MC38 colon adenocarcinoma model, further supporting the notion that NRP1 contributes to, although indirectly, the defective CD8⁺ T cell-mediated primary tumor control. 104

In fact, the most striking phenotype of the E81^{Cre}Nr $pI^{L/L}$ mice is their improved protection against secondary tumors in a mouse model of postsurgical tumor immunity. 104 This observation suggested a primary role for NRP1 in CD8⁺ T cells may be it contributes to the defective CD8⁺ T cell-mediated immunological memory against tumors. Of clinical significance, this model resembles control of disease relapse in patients with cancer. Further investigation revealed that $Nrp\Gamma^{-}$ intratumoral CD8⁺ T cells were more capable of sustaining a stem celllike, memory/exhaustion T cell progenitor phenotype, as opposed to the irreversible differentiation of a terminally exhausted T cell phenotype. Consequently, a larger pool of tumor-specific memory CD8⁺ T cells are formed from these Nrp1^{-/-} memory precursors, a scenario which is not reported when blocking any of the other IRs, either by genetic deficiency or by antibody blockade.

In contrast to effector CD8⁺ T cells, the reported expression of NRP1 on memory CD8⁺ T cells varies substantially between studies. During acute viral infection, Nrp1 gene transcription was downregulated (although higher than naive cells) on long-lived memory cells compared with

T effectors. 98 Such downregulation of NRP1 expression was even more striking on tumor-specific memory CD8⁺ T cells, wherein Nrp1 transcription decreased to baseline level in naïve cells (unpublished data). Contrary to these observations, one report described NRP1 as a surface marker for liver-primed memory CD8⁺ T cells generated through liver sinusoidal endothelial cells crosspresenting antigen under non-inflammatory conditions, while it was functionally dispensable for these memory CD8⁺ T cells. ¹⁰⁵ The discrepancies between these studies may be explained in part by different in vivo conditions (eg, inflammatory milieu and the type of antigen) under which functional memory is generated. Thus, the expression of NRP1 on memory CD8⁺ T cells (likely the same for other IRs) is dictated by the environmental milieu during memory formation.

In summary, the evolving biology of NRP1 in CD8⁺ T cells contributes to a hypothesis that independent sets of IRs, or immune 'checkpoints', seem to exist, which respectively control the effector versus memory functions of intratumoral CD8⁺ T cells. Further mechanistic elucidation of this hypothesis is critical, in the hope of improving the durability of T cell-targeted immunotherapeutics, including immune checkpoint blockade and adoptive T cell transfers.

NRP1-TARGETING IN CANCER THERAPY: A NEXT GENERATION **CHECKPOINT MOLECULE**

Clinical investigation of NRP1 has predominantly focused on its contributions to tumor angiogenesis by acting as a coreceptor with VEGFR2, rather than its immune regulatory function through binding semaphorins or other ligands. Initial pharmacokinetic studies of an anti-NRP1 monoclonal antibody (MNRP1685A) demonstrated potent inhibition of the human VEGF pathway with additive efficacy in combination with anti-VEGFA, bevacizumab. 106 However, subsequent safety analysis in Phase I studies revealed high levels of adverse events, most notably grade 2 or 3 proteinuria (protein accumulation in the urine) in over 50% of subjects in one study, that ultimately terminated therapeutic investigation of this agent. 107-109 A recent investigation of an anti-VEGFR2 agent, ramucirumab, found marked impacts on the T_{reg} cell compartment, leaving room for speculation about whether NRP1 function and expression contributed to the observations. 110

To date, only one anti-NRP1 monoclonal antibody is being clinically assessed for its inhibitory effect of $T_{_{\rm reg}}$ cell function. ASP1948 (human IgG4, Astellas Pharma Inc) in combination with Nivolumab is under Phase Ib evaluation (NCT03565445) for patients with advanced solid tumors. Results from this trial are expected in 2022 and will be critical to further assess the clinical significance and potential of targeting NRP1. Immune monitoring of T_{reg} cell and CD8⁺ T cell alterations, even in the patient periphery, could be informative for therapeutic response and mechanism of action.

		Benchmark IRs		NDD4
		PD1	CTLA4	NRP1
CD8+ T cells	Expressed by CD8+ T cells at basal state	×	×	8
	Driven by T cell activation (CD8+ T cells)	>	>	>
	Signaling antagonizes proximal TCR signaling or costimulation	>	>	?
	Inhibits CD8+ T cell effector function (proliferation/cytokine production)	>	>	×
	Inhibits CD8+ T cell memory response	8	×	>
	Blockade <i>increases</i> CD8+ T cell functions	>	>	>
T _{reg} cells	Expressed by T _{reg} cells at basal state	✓ /⊗	\	2
	Driven by T cell activation $(T_regcells)$	>	>	2
	Boosts T _{reg} cell contact-dependent suppression	8	>	>
	Boosts T _{reg} cell cytokine production	8	8	>
	Blockade decreases T _{reg} cell functions	√ /⊗	>	>
TME	Expressed by myeloid cells (basal and TME) and/or tumor cells	√ /⊗	×	\
	Functions in tumor-associated angiogenesis	×	×	<
	Functions in cancer cell epithelial- mesenchymal transition	8	8	<
	Functions in neuronal development	×	×	>
	Clinical Development Status	Standard of care (Many cancers)	Standard of care (Melanoma)	Phase 1b NCT03565445 (Astellas Pharma)

Figure 3 NRP1 is a unique checkpoint target with diverse impacts on tumor immunity. Comparison between NRP1 and benchmark IRs, PD1 and CTLA4, as a checkpoint target in the setting of cancer immunotherapy. ¹ In murine studies, NRP1 ligation was found to recruit PTEN to the immunological synapse to antagonize Akt activation. This has not yet been confirmed in CD8⁺ T cells. ² Whereas mouse tT_{reg} cells constitutively express NRP1, resting human T_{reg} cells do not express NRP1, though it is upregulated with T cell stimulation. CTLA4, cytotoxic T-lymphocyte associated protein 4; IRs, inhibitory receptors; NRP-1, Neuropilin-1; PD1, programmed cell death protein 1; tT_{rec}cells, thymically-derived regulatory T cells.

In addition to this, insight into the potential outcomes of NRP1-directed trials may be gleaned from examining how the semaphorin family, which is the primary group of NRP1 ligands implicated in its immune regulation, is being therapeutically targeted. To this end, although the primary ligand for NRP1 on T cells, Sema4A, has yet to

be targeted in the clinic, preclinical evaluation of agents against Sema3A (a known NRP1 ligand) have yielded promising results. Sema3A is considered a vasculature normalizing factor through its interactions with NRPs complexed with plexins. While multiple studies have shown that Sema3A function inhibits tumor cell growth

and migration, ¹¹¹ ¹¹² one recent study in glioblastoma suggests that neutralization of Sema3A impedes tumor growth in patient-derived xenograft (PDX) models. ¹¹³ The discrepancies between these findings remains to be reconciled; however, it is possible that differences in tumor vasculature requirements and immune infiltration across tumor types could determine whether Sema3A has pro-tumor or antitumor function. In fact, Sema3A intrinsically regulates T cell activation ¹¹⁴ ¹¹⁵ and thus therapeutic blockade of this interaction may mediate enhanced antitumor immunity.

Whereas prior small molecule or peptide antagonists of NRP1 primarily targeted the b1 domain interaction with VEGF-A to reduce tumor cell migration and angiogenesis,⁷⁷ 116 117 novel candidates also appear to intrinsically regulate T_{reg} cell function in vitro. ¹¹⁸ This combinatorial action may permit lower dosing regimens to mitigate potential side effects; however, the therapeutic efficacy and safety profile of the lead candidate (EG01377) has yet to be evaluated with preclinical in vivo models. Although human T_{reg} cells selectively express NRP1 in the context of cancer, making it an attractive tumor-specific immune target, NRP1 is constitutively expressed by subsets of both hematopoietic and non-hematopoietic cells including pDCs and endothelial cells. The impact of EG01377 on the function of these cell types in steady state and disease will further shape its clinical applicability.

CONCLUSIONS AND FUTURE DIRECTIONS

A growing body of literature demonstrates that NRP1 is a unique immune modulator in cancer immunotherapy (figure 3). In the TME, NRP1 intrinsically regulates both T_{reg} cell and CD8⁺ T cell function to collectively impede antitumor immunity and is expressed concurrently with multiple IRs. NRP1 benefits the function of intratumoral T_{reg} cells by both facilitating their recruitment to the tumor bed and enforcing their functional stability amidst ongoing inflammation. Patients with cancer have a higher abundance of NRP1⁺ T_{reg} cells across malignancies and therapeutic intervention is associated with decreased NRP1 expression in peripheral T_{reg} cells. Though of relatively minor consequence to $CD8^+T$ cell effector function, the impact of NRP1 on memory formation and durable response is distinct compared with the function of other IRs. These novel characteristics may translate into nonoverlapping clinical efficacy with existing standard of care checkpoint inhibitors, thereby providing informed rationale for therapeutic combinations.

As the opportunity rises to improve NRP1-targeted cancer therapy by focusing on its immunoregulatory roles, key questions remain regarding fundamental NRP1 biology and translational application.

 Can we improve our understanding of the molecular basis underlying how NRP1 signals in subsets of T cells (ie, T_{reg} cells, CD4⁺ and CD8⁺ T cells), particularly in humans? The delineation of these mechanisms is crucial for developing specific intervention strategies, facilitating

- optimal design for combinatorial immunotherapies, as well as informing ways to reduce the therapy-induced immune-related adverse events.
- 2. Among the numerous ligands for NRP1, which are the most relevant within tumors, particularly for intratumoral $T_{\rm reg}$ cells and CD8⁺ T cells? The answer to this question will help inform blocking strategies that specifically target the relevant tumor-associated NRP1-ligand interactions while sparing physiological NRP1 functions.
- 3. Will NRP1-targeted approaches provide new strategies to improve the durability of adoptive T cell therapy, such as the chimeric antigen receptor (CAR) T cell therapy? NRP1 may be a viable genetic target in this setting to overcome adopted CD8⁺ T cell dysfunction and enhance in vivo duration of response.
- 4. Does NRP1 antagonism intrinsically impact the function of other cells in the TME including APCs and stromal cells? If so, is the effect to potentiate antitumor immunity?
- 5. Will combining NRP1-targeted therapy with standard of care checkpoint inhibitors, such as anti-PD1/PD-L1, achieve better clinical efficacy? What will be the optimal dosing regimen?
- 6. Can peripheral T cell NRP1 expression (either T_{reg} cells or CD8⁺ T cells) be a diagnostic or prognostic biomarker for human patients with cancer? Can it be used to identify optimal candidates for immunotherapy or evaluate on-treatment response?

In conclusion, the translation of NRP1 biology into viable therapeutic interventions for patients with cancer holds substantial future promise, particularly in combination with immunotherapies that do not directly target T_{reg} cell function or CD8 $^+$ T cell memory formation. However, a more holistic view of NRP1 in the complete TME, namely tumor, stroma and immune cells, is necessary to design the most optimal clinical strategies surrounding this promising next generation immune modulator.

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Competing interests DAV has submitted patents covering NRP1 that are licensed or pending and is entitled to a share in net income generated from licensing of these patent rights for commercial development.

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REFERENCES

- 1 Bashey A, Medina B, Corringham S, et al. Ctla4 blockade with ipilimumab to treat relapse of malignancy after allogeneic hematopoietic cell transplantation. Blood 2009;113:1581–8.
- 2 Hodi FS, Butler M, Oble DA, et al. Immunologic and clinical effects of antibody blockade of cytotoxic T lymphocyte-associated antigen 4 in previously vaccinated cancer patients. Proc Natl Acad Sci U S A 2008:105:3005–10.
- 3 Hodi FS, O'Day SJ, McDermott DF, et al. Improved survival with ipilimumab in patients with metastatic melanoma. N Engl J Med 2010:363:711–23.
- 4 O'Day SJ, Maio M, Chiarion-Sileni V, et al. Efficacy and safety of ipilimumab monotherapy in patients with pretreated advanced melanoma: a multicenter single-arm phase II study. Ann Oncol 2010:21:1712–7.
- 5 Wolchok JD, Neyns B, Linette G, et al. Ipilimumab monotherapy in patients with pretreated advanced melanoma: a randomised, double-blind, multicentre, phase 2, dose-ranging study. Lancet Oncol 2010:11:155–64.
- 6 Brahmer JR, Drake CG, Wollner I, et al. Phase I study of single-agent anti-programmed death-1 (MDX-1106) in refractory solid tumors: safety, clinical activity, pharmacodynamics, and immunologic correlates. J Clin Oncol 2010;28:3167–75.
- 7 Brahmer JR, Tykodi SS, Chow LQM, et al. Safety and activity of anti-PD-L1 antibody in patients with advanced cancer. N Engl J Med 2012;366:2455–65.
- 8 Topalian SL, Hodi FS, Brahmer JR, et al. Safety, activity, and immune correlates of anti-PD-1 antibody in cancer. N Engl J Med 2012;366:2443–54.
- 9 Robert C, Long GV, Brady B, et al. Nivolumab in previously untreated melanoma without BRAF mutation. N Engl J Med 2015;372:320–30.
- 10 Ferris RL, Blumenschein G, Fayette J, et al. Nivolumab for recurrent squamous-cell carcinoma of the head and neck. N Engl J Med Overseas Ed 2016;375:1856–67.
- 11 Herbst RS, Soria J-C, Kowanetz M, *et al.* Predictive correlates of response to the anti-PD-L1 antibody MPDL3280A in cancer patients. *Nature* 2014;515:563–7.
- 12 Alexandrov LB, Nik-Zainal S, Wedge DC, et al. Signatures of mutational processes in human cancer. Nature 2013;500:415–21.
- 13 Snyder A, Makarov V, Merghoub T, et al. Genetic basis for clinical response to CTLA-4 blockade in melanoma. N Engl J Med Overseas Ed 2014;371:2189–99.
- 14 Łuksza M, Riaz N, Makarov V, et al. A neoantigen fitness model predicts tumour response to checkpoint blockade immunotherapy. Nature 2017;551:517–20.
- 15 Huang AC, Postow MA, Orlowski RJ, et al. T-Cell invigoration to tumour burden ratio associated with anti-PD-1 response. Nature 2017;545:60–5.
- 16 Mariathasan S, Turley SJ, Nickles D, et al. Tgfβ attenuates tumour response to PD-L1 blockade by contributing to exclusion of T cells. Nature 2018;554:544–8.
- 17 Wolchok JD, Chiarion-Sileni V, Gonzalez R, et al. Overall survival with combined nivolumab and ipilimumab in advanced melanoma. N Engl J Med 2017;377:1345–56.
- 18 Hodi FS, Chiarion-Sileni V, Gonzalez R, et al. Nivolumab plus ipilimumab or nivolumab alone versus ipilimumab alone in advanced melanoma (CheckMate 067): 4-year outcomes of a multicentre, randomised, phase 3 trial. *Lancet Oncol* 2018;19:1480–92.
- 19 Ascierto PA, Melero I, Bhatia S, et al. Initial efficacy of anti-lymphocyte activation gene-3 (anti-LAG-3; BMS-986016) in combination with nivolumab (nivo) in PTS with melanoma (MEL) previously treated with anti-PD-1/PD-L1 therapy. JCO 2017;35:9520.
- 20 Andrews LP, Marciscano AE, Drake CG, et al. LAG3 (CD223) as a cancer immunotherapy target. Immunol Rev 2017;276:80–96.
- 21 Fourcade J, Sun Z, Chauvin J-M, et al. Cd226 opposes TIGIT to disrupt Tregs in melanoma. JCI Insight 2018;3:e121157.
- 22 Solomon BL, Garrido-Laguna I. Tigit: a novel immunotherapy target moving from bench to bedside. *Cancer Immunol Immunother* 2018;67:1659–67.
- 23 Ngiow SF, Teng MWL, Smyth MJ. Prospects for TIM3-Targeted antitumor immunotherapy. *Cancer Res* 2011;71:6567–71.
- 24 Anderson AC, Joller N, Kuchroo VK. Lag-3, Tim-3, and TIGIT: Coinhibitory receptors with specialized functions in immune regulation. Immunity 2016;44:989–1004.
- 25 Pardoll DM. The blockade of immune checkpoints in cancer immunotherapy. Nat Rev Cancer 2012;12:252–64.
- 26 Wherry EJ, Kurachi M. Molecular and cellular insights into T cell exhaustion. *Nat Rev Immunol* 2015;15:486–99.

- 27 Yano H, Andrews LP, Workman CJ, et al. Intratumoral regulatory T cells: markers, subsets and their impact on anti-tumor immunity. <u>Immunology</u> 2019;157:232–47.
- Wing K, Onishi Y, Prieto-Martin P, et al. Ctla-4 control over Foxp3+ regulatory T cell function. Science 2008;322:271–5.
- 29 Kurtulus S, Sakuishi K, Ngiow S-F, et al. Tigit predominantly regulates the immune response via regulatory T cells. J Clin Invest 2015:125:4053–62.
- 30 Joller N, Lozano E, Burkett PR, et al. Treg cells expressing the Coinhibitory molecule TIGIT selectively inhibit proinflammatory Th1 and Th17 cell responses. *Immunity* 2014;40:569–81.
- 31 Liu Z, McMichael EL, Shayan G, et al. Novel Effector Phenotype of Tim-3⁺ Regulatory T Cells Leads to Enhanced Suppressive Function in Head and Neck Cancer Patients. Clin Cancer Res 2018;24:4529–38.
- 32 Gautron A-S, Dominguez-Villar M, de Marcken M, et al. Enhanced suppressor function of TIM-3+ Foxp3+ regulatory T cells. Eur J Immunol 2014;44:2703–11.
- 33 Lowther DE, Goods BA, Lucca LE, et al. Pd-1 marks dysfunctional regulatory T cells in malignant gliomas. JCI Insight 2016;1:e85935.
- 34 Kamada T, Togashi Y, Tay C, et al. PD-1 ⁺ regulatory T cells amplified by PD-1 blockade promote hyperprogression of cancer. Proc Natl Acad Sci U S A 2019;116:9999–10008.
- 35 Zhang Q, Chikina M, Szymczak-Workman AL, et al. LAG3 limits regulatory T cell proliferation and function in autoimmune diabetes. Sci Immunol 2017;2:eaah4569.
- 36 Thaker YR, Andrews LP, Workman CJ, et al. Treg-specific LAG3 deletion reveals a key role for LAG3 in regulatory T cells to inhibit CNS autoimmunity. J Immunol 2018;200:101.7.
- 37 Bruder D, Probst-Kepper M, Westendorf AM, et al. Frontline: neuropilin-1: a surface marker of regulatory T cells. Eur J Immunol 2004;34:623–30.
- 38 Delgoffe GM, Woo S-R, Turnis ME, et al. Stability and function of regulatory T cells is maintained by a neuropilin-1–semaphorin-4a axis. *Nature* 2013;501:252–6.
- 39 Overacre-Delgoffe AE, Chikina M, Dadey RE, et al. Interferon-γ Drives T_{reg} Fragility to Promote Anti-tumor Immunity. Cell 2017;169:1130–41.
- 40 Leclerc M, Voilin E, Gros G, et al. Regulation of antitumour CD8 T-cell immunity and checkpoint blockade immunotherapy by neuropilin-1. Nat Commun 2019;10:3345.
- 41 Rossignol M, Gagnon ML, Klagsbrun M. Genomic organization of human neuropilin-1 and neuropilin-2 genes: identification and distribution of splice variants and soluble isoforms. *Genomics* 2000:70:211–22
- 42 Kitsukawa T, Shimono A, Kawakami A, et al. Overexpression of a membrane protein, neuropilin, in chimeric mice causes anomalies in the cardiovascular system, nervous system and limbs. *Development* 1995;121:4309–18.
- 43 Kawasaki T, Kitsukawa T, Bekku Y, et al. A requirement for neuropilin-1 in embryonic vessel formation. *Development* 1999:126:4895–902.
- 44 Yuan L, Moyon D, Pardanaud L, et al. Abnormal lymphatic vessel development in neuropilin 2 mutant mice. *Development* 2002;129:4797–806.
- 45 Rizzolio S, Tamagnone L. Multifaceted role of neuropilins in cancer. *Curr Med Chem* 2011;18:3563–75.
- 46 Roy S, Bag AK, Singh RK, et al. Multifaceted role of neuropilins in the immune system: potential targets for immunotherapy. Front Immunol 2017;8:1228.
- 47 Gu C, Limberg BJ, Whitaker GB, et al. Characterization of neuropilin-1 structural features that confer binding to semaphorin 3A and vascular endothelial growth factor 165. J Biol Chem 2002:277:18069–76.
- 48 Geretti E, Shimizu A, Klagsbrun M. Neuropilin structure governs VEGF and semaphorin binding and regulates angiogenesis. *Angiogenesis* 2008;11:31–9.
- 49 Valdembri D, Caswell PT, Anderson KI, et al. Neuropilin-1/GIPC1 signaling regulates α5β1 integrin traffic and function in endothelial cells. PLoS Biol 2009;7:e25.
- 50 Prahst C, Héroult M, Lanahan AA, et al. Neuropilin-1-VEGFR-2 complexing requires the PDZ-binding domain of neuropilin-1. J Biol Chem 2008;283:25110–4.
- 51 Kolodkin AL, Levengood DV, Rowe EG, et al. Neuropilin is a semaphorin III receptor. Cell 1997;90:753–62.
- 52 Glinka Y, Prud'homme GJ. Neuropilin-1 is a receptor for transforming growth factor beta-1, activates its latent form, and promotes regulatory T cell activity. J Leukoc Biol 2008;84:302–10.
- 53 Glinka Y, Stoilova S, Mohammed N, et al. Neuropilin-1 exerts co-receptor function for TGF-beta-1 on the membrane of cancer



- cells and enhances responses to both latent and active TGF-beta. *Carcinogenesis* 2011;32:613–21.
- Muhl L, Folestad EB, Gladh H, et al. Neuropilin 1 binds PDGF-D and is a co-receptor in PDGF-D-PDGFRβ signaling. J Cell Sci 2017:130:1365–78.
- 55 Ruffini F, Levati L, Graziani G, et al. Platelet-Derived growth factor-C promotes human melanoma aggressiveness through activation of neuropilin-1. Oncotarget 2017;8:66833–48.
- 56 Niland S, Ditkowski B, Parrandier D, et al. Rhodocetin-αβ-induced neuropilin-1-cMet association triggers restructuring of matrix contacts in endothelial cells. Arterioscler Thromb Vasc Biol 2013;33:544–54.
- 57 Prud'homme GJ, Glinka Y, Lichner Z, et al. Neuropilin-1 is a receptor for extracellular miRNA and AGO2/miRNA complexes and mediates the internalization of miRNAs that modulate cell function. Oncotarget 2016;7:68057–71.
- Kitsukawa T, Shimizu M, Sanbo M, et al. Neuropilin–Semaphorin III/ D-Mediated chemorepulsive signals play a crucial role in peripheral nerve projection in mice. Neuron 1997;19:995–1005.
- 59 Gu C, Rodriguez ER, Reimert DV, et al. Neuropilin-1 conveys semaphorin and VEGF signaling during neural and cardiovascular development. *Dev Cell* 2003;5:45–57.
- 60 Dzionek A, Fuchs A, Schmidt P, et al. BDCA-2, BDCA-3, and BDCA-4: three markers for distinct subsets of dendritic cells in human peripheral blood. *J Immunol* 2000;165:6037–46.
- 61 Grage-Griebenow E, Löseke S, Kauth M, et al. Anti-BDCA-4 (neuropilin-1) antibody can suppress virus-induced IFN-alpha production of plasmacytoid dendritic cells. *Immunol Cell Biol* 2007;85:383–90.
- 62 Miyauchi JT, Caponegro MD, Chen D, et al. Deletion of neuropilin 1 from microglia or bone Marrow–Derived macrophages slows glioma progression. Cancer Res 2018;78:685–94.
- 63 Wilson AM, Shao Z, Grenier V, et al. Neuropilin-1 expression in adipose tissue macrophages protects against obesity and metabolic syndrome. Sci Immunol 2018;3:eaan4626.
- 64 Casazza A, Laoui D, Wenes M, et al. Impeding macrophage entry into hypoxic tumor areas by Sema3A/Nrp1 signaling blockade inhibits angiogenesis and restores antitumor immunity. Cancer Cell 2013;24:695–709.
- Wallerius M, Wallmann T, Bartish M, et al. Guidance molecule Sema3A restricts tumor growth by differentially regulating the proliferation of tumor-associated macrophages. Cancer Res 2016;76:3166–78.
- 66 DeFalco T, Bhattacharya I, Williams AV, et al. Yolk-sacderived macrophages regulate fetal testis vascularization and morphogenesis. Proc Natl Acad Sci U S A 2014;111:E2384–93.
- 67 Solomon BD, Mueller C, Chae W-J, et al. Neuropilin-1 attenuates autoreactivity in experimental autoimmune encephalomyelitis. Proc Natl Acad Sci U S A 2011;108:2040–5.
- 68 Renand A, Milpied P, Rossignol J, et al. Neuropilin-1 expression characterizes T follicular helper (Tfh) cells activated during B cell differentiation in human secondary lymphoid organs. PLoS One 2013:8:085580
- 69 Milpied P, Massot B, Renand A, et al. IL-17–producing invariant NKT cells in lymphoid organs are recent thymic emigrants identified by neuropilin-1 expression. *Blood* 2011;118:2993–3002.
- 70 Lepelletier Y, Smaniotto S, Hadj-Slimane R, et al. Control of human thymocyte migration by Neuropilin-1/Semaphorin-3A-mediated interactions. Proc Natl Acad Sci U S A 2007;104:5545–50.
- 71 Tordjman R, Lepelletier Y, Lemarchandel V, et al. A neuronal receptor, neuropilin-1, is essential for the initiation of the primary immune response. Nat Immunol 2002;3:477–82.
- 72 Sarris M, Andersen KG, Randow F, et al. Neuropilin-1 expression on regulatory T cells enhances their interactions with dendritic cells during antigen recognition. *Immunity* 2008;28:402–13.
- 73 Bourbié-Vaudaine S, Blanchard N, Hivroz C, et al. Dendritic cells can turn CD4⁺ T lymphocytes into vascular endothelial growth factor-carrying cells by intercellular neuropilin-1 transfer. J Immunol 2006;177:1460-9.
- 74 Oussa NAE, Dahmani A, Gomis M, et al. Vegf requires the receptor NRP-1 to inhibit lipopolysaccharide-dependent dendritic cell maturation. J Immunol 2016;197:3927–35.
- 75 Yaqoob U, Cao S, Shergill U, et al. Neuropilin-1 Stimulates Tumor Growth by Increasing Fibronectin Fibril Assembly in the Tumor Microenvironment. Cancer Res 2012;72:4047–59.
- 76 Niland S, Eble JA. Neuropilins in the context of tumor vasculature. Int J Mol Sci 2019;20:639.
- 77 Jia H, Cheng L, Tickner M, et al. Neuropilin-1 antagonism in human carcinoma cells inhibits migration and enhances chemosensitivity. Br J Cancer 2010;102:541–52.

- 78 Weiss JM, Bilate AM, Gobert M, et al. Neuropilin 1 is expressed on thymus-derived natural regulatory T cells, but not mucosa-generated induced Foxp3+ T reg cells. J Exp Med 2012;209:1723–42.
- 79 Singh K, Hjort M, Thorvaldson L, et al. Concomitant analysis of Helios and neuropilin-1 as a marker to detect thymic derived regulatory T cells in naïve mice. Sci Rep 2015;5:7767.
- 80 Yadav M, Louvet C, Davini D, et al. Neuropilin-1 distinguishes natural and inducible regulatory T cells among regulatory T cell subsets in vivo. J Exp Med 2012;209:1713–22.
- 81 Hill JA, Feuerer M, Tash K, et al. Foxp3 Transcription-Factor-Dependent and -independent regulation of the regulatory T cell transcriptional signature. Immunity 2007;27:786–800.
- 82 Zheng Y, Josefowicz SZ, Kas A, et al. Genome-Wide analysis of FOXP3 target genes in developing and mature regulatory T cells. Nature 2007;445:936–40.
- 83 Marson A, Kretschmer K, Frampton GM, et al. Foxp3 occupancy and regulation of key target genes during T-cell stimulation. *Nature* 2007:445:931–5.
- 84 Milpied P, Renand A, Bruneau J, et al. Neuropilin-1 is not a marker of human Foxp3+ Treg. Eur J Immunol 2009;39:1466–71.
- 85 Hansen W, Hutzler M, Abel S, et al. Neuropilin 1 deficiency on CD4+Foxp3+ regulatory T cells impairs mouse melanoma growth. J Exp Med 2012;209:2001–16.
- 86 Soker S, Takashima S, Miao HQ, et al. Neuropilin-1 is expressed by endothelial and tumor cells as an isoform-specific receptor for vascular endothelial growth factor. Cell 1998;92:735–45.
- 87 Soker S, Miao H-Q, Nomi M, et al. Vegf165 mediates formation of complexes containing VEGFR-2 and neuropilin-1 that enhance VEGF165-receptor binding. J Cell Biochem 2002;85:357–68.
- 88 Overacre-Delgoffe AE, Vignali DAA. Treg fragility: a prerequisite for effective antitumor immunity? Cancer Immunol Res 2018;6:882–7.
- 89 Overacre AE, Vignali DAA. Treg stability: to be or not to be. Curr Opin Immunol 2016;39:39–43.
- 90 Jung K, Kim J-A, Kim Y-J, et al. A neuropilin-1 antagonist exerts antitumor immunity by inhibiting the suppressive function of intratumoral regulatory T cells. Cancer Immunol Res 2020;8:46–56.
- 91 Chaudhary B, Khaled YS, Ammori BJ, et al. Neuropilin 1: function and therapeutic potential in cancer. Cancer Immunol Immunother 2014;63:81–99.
- 92 Piechnik A, Dmoszynska A, Omiotek M, et al. The VEGF receptor, neuropilin-1, represents a promising novel target for chronic lymphocytic leukemia patients. Int J Cancer 2013;133:1489–96.
- 93 Battaglia A, Buzzonetti A, Monego G, et al. Neuropilin-1 expression identifies a subset of regulatory T cells in human lymph nodes that is modulated by preoperative chemoradiation therapy in cervical cancer. Immunology 2008;123:129–38.
- 94 Battaglia A, Buzzonetti A, Baranello C, et al. Metastatic tumour cells favour the generation of a tolerogenic milieu in tumour draining lymph node in patients with early cervical cancer. cancer immunology. *Immunotherapy* 2009;58:1363–73.
- 95 Battaglia A, Buzzonetti A, Martinelli E, et al. Selective changes in the immune profile of Tumor-Draining lymph nodes after different neoadjuvant chemoradiation regimens for locally advanced cervical cancer. Int J Radiat Oncol Biol Phys 2010;76:1546–53.
- 96 E XQ, Meng HX, Cao Y, et al. Distribution of regulatory T cells and interaction with dendritic cells in the synovium of rheumatoid arthritis. Scand J Rheumatol 2012;41:413–20.
- 97 Naik A, Al-Zeheimi N, Bakheit CS, et al. Neuropilin-1 associated molecules in the blood distinguish poor prognosis breast cancer: a cross-sectional study. Sci Rep 2017;7:3301.
- 98 Kaech SM, Hemby S, Kersh E, et al. Molecular and functional profiling of memory CD8 T cell differentiation. Cell 2002;111:837–51.
- PFleissner D, Hansen W, Geffers R, et al. Local induction of immunosuppressive CD8+ T cells in the gut-associated lymphoid tissues. PLoS One 2010;5:e15373.
- 100 Jackson SR, Berrien-Elliott M, Yuan J, et al. Neuropilin-1 expression is induced on tolerant self-reactive CD8+ T cells but is dispensable for the tolerant phenotype. PLoS One 2014;9:e110707.
- 101 Martinez GJ, Pereira RM, Äijö T, et al. The transcription factor NFAT promotes exhaustion of activated CD8⁺ T cells. Immunity 2015;42:265–78.
- 102 Liu X, Wang Y, Lu H, et al. Genome-Wide analysis identifies NR4A1 as a key mediator of T cell dysfunction. Nature 2019;567:525–9.
- 103 Miller BC, Sen DR, Al Abosy R, et al. Subsets of exhausted CD8⁺ T cells differentially mediate tumor control and respond to checkpoint blockade. Nat Immunol 2019;20:326–36.
- 104 Liu C, Somasundaram A, Manne S, et al. Neuropilin-1 is a T cell memory checkpoint limiting long-term anti-tumor immunity. Nat Immunol 2020.



- 105 Böttcher JP, Schanz O, Wohlleber D, et al. Liver-primed memory T cells generated under noninflammatory conditions provide antiinfectious immunity. Cell Rep 2013;3:779–95.
- 106 Xin Y, Li J, Wu J, et al. Pharmacokinetic and pharmacodynamic analysis of circulating biomarkers of anti-NRP1, a novel antiangiogenesis agent, in two phase I trials in patients with advanced solid tumors. Clin Cancer Res 2012;18:6040–8.
- 107 Weekes CD, Beeram M, Tolcher AW, et al. A phase I study of the human monoclonal anti-NRP1 antibody MNRP1685A in patients with advanced solid tumors. *Invest New Drugs* 2014;32:653–60.
- 108 Weekes CD, LoRusso P, Ramakrishnan V, et al. A phase Ib study for MNRP1685A (anti-NRP1) administered intravenously with bevacizumab with or without paclitaxel to patients with advanced solid tumors. Journal of Clinical Oncology 2011;29:3050.
- 109 Patnaik A, LoRusso PM, Messersmith WA, et al. A phase lb study evaluating MNRP1685A, a fully human anti-NRP1 monoclonal antibody, in combination with bevacizumab and paclitaxel in patients with advanced solid tumors. Cancer Chemother Pharmacol 2014;73:951–60.
- 110 Tada Y, Togashi Y, Kotani D, et al. Targeting VEGFR2 with Ramucirumab strongly impacts effector/ activated regulatory T cells and CD8⁺ T cells in the tumor microenvironment. J Immunother Cancer 2018;6:106.
- 111 Chakraborty G, Kumar S, Mishra R, et al. Semaphorin 3A suppresses tumor growth and metastasis in mice melanoma model. PLoS One 2012;7:e33633.

- 112 Gioelli N, Maione F, Camillo C, et al. A rationally designed NRP1-independent superagonist Sema3A mutant is an effective anticancer agent. Sci Transl Med 2018;10:eaah4807.
- 113 Lee J, Shin YJ, Lee K, et al. Anti-SEMA3A antibody: a novel therapeutic agent to suppress glioblastoma tumor growth. Cancer Res Treat 2018;50:1009–22.
- 114 Lepelletier Y, Moura IC, Hadj-Slimane R, et al. Immunosuppressive role of semaphorin-3A on T cell proliferation is mediated by inhibition of actin cytoskeleton reorganization. Eur J Immunol 2006;36:1782–93.
- 115 Catalano A, Caprari P, Moretti S, et al. Semaphorin-3A is expressed by tumor cells and alters T-cell signal transduction and function. Blood 2006;107:3321–9.
- 116 Starzec A, Vassy R, Martin A, et al. Antiangiogenic and antitumor activities of peptide inhibiting the vascular endothelial growth factor binding to neuropilin-1. Life Sci 2006;79:2370–81.
- 117 Jarvis A, Allerston CK, Jia H, et al. Small molecule inhibitors of the neuropilin-1 vascular endothelial growth factor A (VEGF-A) interaction. J Med Chem 2010;53:2215–26.
- 118 Powell J, Mota F, Steadman D, *et al*. Small molecule neuropilin-1 antagonists combine antiangiogenic and antitumor activity with immune modulation through reduction of transforming growth factor beta (TGFβ) production in regulatory T-cells. *J Med Chem* 2018;61:4135–54.