

*Supplementary Material***Supplementary Tables****Supplementary Table 1** Clinicopathological characteristics of the 49 individual patients analysed for immunotherapy response.

<b>Patient ID</b>	<b>ICB Treatment (target)</b>	<b>Line of systemic treatment</b>	<b>Response status</b>	<b>Hepatitis status</b>	<b>AFP Level</b>	<b>Age (years)</b>	<b>Stage*</b>	<b>Grade<sup>#</sup></b>
HCC 001	Combination PD-1/PD-L1 + CTLA-4	1st	Progressive Disease (PD)	None	34206	70	C	3
HCC 002	PD-1/PD-L1	1st	Partial Response (PR)	HCV	2.6	60	C	2
HCC 003	Combination PD-1/PD-L1 + others	1st	Stable Disease (SD)	HBV	4.2	78	C	3
HCC 004	Combination PD-1/PD-L1 + CTLA-4	1st	Progressive Disease (PD)	None	1384	76	C	3
HCC 005	PD-1/PD-L1	1st	Progressive Disease (PD)	None	60500	73	C	2
HCC 006	PD-1/PD-L1	1st	Progressive Disease (PD)	HBV	6	52	C	2
HCC 007	Combination PD-1/PD-L1 + others	2nd	Progressive Disease (PD)	HBV	1138	62	B	3
HCC 008	PD-1/PD-L1	1st	Stable Disease (SD)	HBV	4	65	C	3
HCC 009	PD-1/PD-L1	1st	Partial Response (PR)	HCV	48392	65	C	3
HCC 010	Combination PD-1/PD-L1 + CTLA-4	1st	Partial Response (PR)	HBV	832	79	C	3
HCC 011	PD-1/PD-L1	2nd	Progressive Disease (PD)	None	20	67	B	2
HCC 012	PD-1/PD-L1	1st	Partial Response (PR)	None	11.5	81	C	2
HCC 013	PD-1/PD-L1	1st	Progressive Disease (PD)	None	16.8	71	C	2
HCC 014	PD-1/PD-L1	1st	Stable Disease (SD)	HBV	87	79	C	3

HCC 015	PD-1/PD-L1	1st	Progressive Disease (PD)	None	2.3	66	C	1
HCC 016	PD-1/PD-L1	1st	Progressive Disease (PD)	HBV	1359	72	C	N.A.
HCC 017	Combination PD-1/PD-L1 + CTLA-4	1st	Stable Disease (SD)	HBV	2104	35	C	3
HCC 018	PD-1/PD-L1	2nd	Partial Response (PR)	None	14450	68	C	1
HCC 019	PD-1/PD-L1	1st	Stable Disease (SD)	HBV	5.8	72	C	2
HCC 020	PD-1/PD-L1	2nd	Progressive Disease (PD)	None	4.1	76	C	N.A.
HCC 021	PD-1/PD-L1	1st	Progressive Disease (PD)	HBV	11.2	63	C	N.A.
HCC 022	PD-1/PD-L1	1st	Stable Disease (SD)	HBV	4373	78	C	N.A.
HCC 023	PD-1/PD-L1	2nd	Progressive Disease (PD)	None	2309	56	C	N.A.
HCC 024	Combination PD-1/PD-L1 + CTLA-4	1st	Progressive Disease (PD)	HBV	1.8	70	C	N.A.
HCC 026	PD-1/PD-L1	1st	Progressive Disease (PD)	None	31	63	C	2
HCC 027	Combination PD-1/PD-L1 + others	2nd	Stable Disease (SD)	None	858	76	B	2
HCC 028	Combination PD-1/PD-L1 + CTLA-4	3rd	Progressive Disease (PD)	None	764	55	C	N.A.
HCC 029	Combination PD-1/PD-L1 + CTLA-4	1st	Partial Response (PR)	HCV	45.4	69	C	3
HCC 030	PD-1/PD-L1	2nd	Progressive Disease (PD)	None	11	80	C	2
HCC 032	PD-1/PD-L1	2nd	Progressive Disease (PD)	HBV	164	55	C	2
HCC 033	Combination PD-1/PD-L1 + CTLA-4	1st	Progressive Disease (PD)	HBV	60500	68	C	3
HCC 034	Combination PD-1/PD-L1 + CTLA-4	2nd	Progressive Disease (PD)	HBV	27.2	60	C	3
HCC 035	Combination PD-1/PD-L1 + CTLA-4	1st	Stable Disease (SD)	HBV	1.7	45	C	2
HCC 036	PD-1/PD-L1	1st	Partial Response (PR)	None	9.9	79	C	2
HCC 037	PD-1/PD-L1	1st	Progressive Disease (PD)	HBV	121	58	C	2

HCC 038	PD-1/PD-L1	1st	Partial Response (PR)	HBV	70	54	C	3
HCC 040	Combination PD-1/PD-L1 + others	1st	Partial Response (PR)	HBV	69	72	B	2
HCC 041	Combination PD-1/PD-L1 + others	1st	Stable Disease (SD)	HBV	5.4	80	C	2
HCC 042	PD-1/PD-L1	2nd	Progressive Disease (PD)	HBV	253	62	B	
HCC 043	Combination PD-1/PD-L1 + others	2nd	progressive Disease (PD)	None	4.7	64	C	2
HCC 044	Combination PD-1/PD-L1 + CTLA-4	2nd	Partial Response (PR)	HCV	1.5	63	C	N.A.
HCC 045	Combination PD-1/PD-L1 + CTLA-4	2nd	Partial Response (PR)	HBV	301	51	C	N.A.
HCC 046	PD-1/PD-L1	4th	Progressive Disease (PD)	HBV	392	50	C	N.A.
HCC 047	PD-1/PD-L1	2nd	Stable Disease (SD)	None	16.4	76	C	N.A.
HCC 050	Combination PD-1/PD-L1 + CTLA-4	1st	Progressive Disease (PD)	None	4.1	78	C	N.A.
HCC 053	PD-1/PD-L1	1st	Progressive Disease (PD)	HBV	60500	53	C	3
HCC 055	PD-1/PD-L1	1st	Progressive Disease (PD)	HCV	60500	66	C	2 to 3
HCC 057	PD-1/PD-L1	1st	Progressive Disease (PD)	HBV	3.3	77	C	N.A.
HCC 060	PD-1/PD-L1	1st	Progressive Disease (PD)	HBV	73.8	56	C	N.A.

\* Staged according to the BCLC staging system <sup>1</sup>. ICB, immune checkpoint blockade # Graded according to the 4-scale Edmondson and Steiner grading system <sup>2</sup>. HBV, hepatitis B virus; HCV, hepatitis C virus; AFP, alpha-fetoprotein; PD, progressive disease; PR, partial response; SD, stable disease; PD-1, programmed cell death protein-1; PD-L1, programmed death-ligand 1; CTLA-4, cytotoxic T-lymphocyte-associated protein 4; TKI, Tyroxine Kinase Inhibitor; VEGF, Vascular endothelial growth factor; N.A, not applicable.

**Supplementary Table 2.** List of antibodies used for multiplex immunofluorescence and immunohistochemistry.

Antibody	Source	Labelling pattern
DAPI	PerkinElmer, Inc. (FP1490)	Cell nucleus
CD38	Novocastra (NCL-CD38-290)	Immune cells, cell membrane
CD68	Dako (M0876)	Immune cells, cytoplasm
CD8	Novocastra (NCL-CD8-4B11)	Immune cells, cell membrane
PD-L1	Ventana Medical Systems (SP263)	Tumour and immune cells, cell membrane

**Supplementary Table 3** List of antibodies used for flow cytometry.

#	Antigen	Fluorochrome	Clone	Manufacturer	Catalogue number
1	CD16	eFluor 450	CB16	Invitrogen; Thermo Fisher Scientific, Inc.	48-0168-42
2	CD14	BV510	M5E2	BioLegend	301842
3	CD11c	BB515	B-ly6	BD Biosciences	564490
4	CD11b	PerCP-Cy5.5	ICRF44	BioLegend	301328
5	CD38	PerCP-eFluor710	HB7	Invitrogen; Thermo Fisher Scientific, Inc.	46-0388-42
6	CD123	Alexa 647	6H6	BioLegend	306024
7	HLA-DR	APC-Fire 750	L243	BioLegend	307658
8	CD68	PE-Cy7	Y1/82A	Invitrogen; Thermo Fisher Scientific, Inc.	25-0689-42

**Supplementary Table 4.** Ten significantly enriched biological pathways associated with differentially expressed genes between high- and low-CD38 hepatocellular carcinoma, as determined by ingenuity pathway analysis.

Pathway	Genes	-Log (P-value)
Th1 and Th2 Activation Pathway	CCR1,CCR4,CCR8,CD247,CD274,CD28,CD3D,CD3E,CD3G,CD4,CD40LG,CD80,CD86,CD8A,CXCR3,CXCR4,CXCR6,HAVCR1,HAVCR2,HLA-DMA,HLA-DMB,HLA-DOA,HLA-DOB,HLA-DQA2,HLA-DRA,HLA-DRB1,HLA-DRB5,ICAM1,ICOS,IFNG,IKZF1,IL10,IL10RA,IL12A,IL12RB1,IL18,IL18R1,IL1RL1,IL24,IL2RA,IL2RB,IL2RG,IL6,ITGB2,JAK3,KLRC1,KLRD1,LTA,PIK3CD,PIK3CG,PIK3R5,PRKCQ,RUNX3,SOCS1,SPI1,STAT4,TIMD4,VAV1	2.64E+01
Granulocyte Adhesion and Diapedesis	CCL11,CCL13,CCL17,CCL18,CCL19,CCL2,CCL20,CCL21,CCL22,CCL23,CCL24,CCL3,CCL3L1,CCL3L3,CCL4,CCL4L1/CCL4L2,CLDN11,CSF3R,CXCL1,CXCL10,CXCL11,CXCL12,CXCL14,CXCL3,CXCL5,CXCL6,CXCL8,CXCL9,CXCR4,FPR1,FPR3,HRH2,ICAM1,IL18,IL18RAP,IL1B,IL1RL1,ITGA4,ITGB2,MMP12,MMP16,MMP2,MMP23B,MMP25,MMP7,MMP9,SELE,SELL,SELP,SELPLG,TNF,TNFRSF11B,VCAM1,XCL1,XCL2	2.09E+01
Th1 Pathway	CD247,CD274,CD28,CD3D,CD3E,CD3G,CD4,CD40LG,CD80,CD86,CD8A,CXCR3,HAVCR2,HLA-DMA,HLA-DMB,HLA-DOA,HLA-DOB,HLA-DQA2,HLA-DRA,HLA-DRB1,HLA-DRB5,ICAM1,ICOS,IFNG,IL10,IL10RA,IL12A,IL12RB1,IL18,IL18R1,IL6,ITGB2,JAK3,KLRC1,KLRD1,LTA,PIK3CD,PIK3CG,PIK3R5,PRKCQ,RUNX3,SOCS1,STAT4,VAV1	1.95E+01
Th2 Pathway	CCR1,CCR4,CCR8,CD247,CD28,CD3D,CD3E,CD3G,CD4,CD80,CD86,CXCR4,CXCR6,HAVCR1,HLA-DMA,HLA-DMB,HLA-DOA,HLA-DOB,HLA-DQA2,HLA-DRA,HLA-DRB1,HLA-DRB5,ICAM1,ICOS,IFNG,IKZF1,IL10,IL12A,IL12RB1,IL1RL1,IL24,IL2RA,IL2RB,IL2RG,ITGB2,JAK3,PIK3CD,PIK3CG,PIK3R5,PRKCQ,RUNX3,SPI1,STAT4,TIMD4,VAV1	1.81E+01
Agranulocyte Adhesion and Diapedesis	ACTC1,CCL11,CCL13,CCL17,CCL18,CCL19,CCL2,CCL20,CCL21,CCL22,CCL23,CCL24,CCL3,CCL3L1,CCL3L3,CCL4,CCL4L1/CCL4L2,CLDN11,CXCL1,CXCL10,CXCL11,CXCL12,CXCL14,CXCL3,CXCL5,CXCL6,CXCL8,CXCL9,CXCR4,ICAM1,IL18,IL1B,ITGA4,ITGB2,MMP12,MMP16,MMP2,MMP23B,MMP25,MMP7,MMP9,SELE,SELL,SELP,SELPLG,TNF,VCAM1,XCL1,XCL2	1.80E+01
Communication between Innate and Adaptive Immune Cells	CCL3,CCL3L3,CCL4,CCR7,CD28,CD4,CD40LG,CD79A,CD80,CD86,CD8A,CXCL10,CXCL8,FCER1G,HLA-DRA,HLA-DRB1,HLA-DRB5,IFNG,IL10,IL12A,IL18,IL1B,IL6,TLR10,TLR2,TNF,TNFRSF13B,TNFRSF13C,TNFRSF17,TNFSF13B	1.63E+01

Altered T Cell and B Cell Signaling in Rheumatoid Arthritis	CCL21,CD28,CD40LG,CD79A,CD80,CD86,FASLG,FCER1G,HLA-DMA,HLA-DMB,HLA-DOA,HLA-DOB,HLA-DRA,HLA-DRB1,HLA-DRB5,IFNG,IL10,IL12A,IL18,IL1B,IL6,LTA,LTB,SLAMF1,TLR10,TLR2,TNF,TNFRSF13B,TNFRSF13C,TNFRSF17,TNFSF13B	1.61E+01
T Helper Cell Differentiation	CD28,CD40LG,CD80,CD86,FCER1G,HLA-DMA,HLA-DMB,HLA-DOA,HLA-DOB,HLA-DRA,HLA-DRB1,HLA-DRB5,ICOS,IFNG,IL10,IL10RA,IL12A,IL12RB1,IL18,IL18R1,IL21R,IL2RA,IL2RG,IL6,STAT4,TNF,TNFRSF11B	1.42E+01
iCOS-iCOSL Signaling in T Helper Cells	CAMK4,CD247,CD28,CD3D,CD3E,CD3G,CD4,CD40LG,CD80,CD86,FCER1G,GRAP2,HLA-DMA,HLA-DMB,HLA-DOA,HLA-DOB,HLA-DRA,HLA-DRB1,HLA-DRB5,ICOS,IKBKE,IL2RA,IL2RB,IL2RG,INPP5D,ITK,LCK,LCP2,PIK3CD,PIK3CG,PIK3R5,PRKCQ,PTPRC,VAV1,ZAP70	1.42E+01
Allograft Rejection Signaling	CD28,CD40LG,CD80,CD86,FASLG,FCER1G,GZMB,HLA-DMA,HLA-DMB,HLA-DOA,HLA-DOB,HLA-DQA2,HLA-DRA,HLA-DRB1,HLA-DRB5,IFNG,IL10,PRF1,TNF	1.35E+01

**Supplementary Table 5.** Analysis of mPFS in patients with HCC treated with anti PD-1/PD-L1 single agent (n=30).

Factor	mPFS (months)	OR (95% CI)	P-value
<b>Intratumoural total CD38<sup>+</sup> cell proportion</b>			
Low	1.45	Reference	
High	11.56	0.397 (0.177, 0.892)	<b>0.0253*</b>
<b>Intratumoural CD38<sup>+</sup>CD68<sup>+</sup> macrophage density</b>			
Low	1.58		
High	19.09	0.381 (0.157, 0.923)	<b>0.0325*</b>
<b>Intratumoural CD38<sup>-</sup>CD68<sup>+</sup> macrophage density</b>			
Low	1.58	Reference	
High	2.20	1.000 (0.467, 2.170)	0.9998
<b>Intratumoural CD38<sup>+</sup>CD68<sup>-</sup> cells density</b>			
Low	2.66	Reference	
High	1.61	1.025 (0.476, 2.206)	0.9503
<b>Intratumoural CD8<sup>+</sup> T cell density</b>			
Low	2.68	Reference	
High	1.61	0.710 (0.295, 1.707)	0.4435
<b>PD-L1 tumour proportion score (TPS)</b>			
< 1	1.74	Reference	
≥ 1%	1.68	0.952 (0.426, 2.129)	0.9056

\*P<0.05 indicated a statistically significant difference. #PD-L1, programmed death-ligand 1; mPFS, median progression free survival.



**Supplementary Table 6.** Analysis of mOS in patients with HCC treated with anti PD-1/PD-L1 single agent (n=30).

Factor	mOS (months)	OR (95% CI)	P-value
<b>Intratumoural total CD38<sup>+</sup> cell proportion</b>			
Low	6.87	Reference	
High	13.96	0.418 (0.175, 0.993)	<b>0.0483*</b>
<b>Intratumoural CD38<sup>+</sup>CD68<sup>+</sup> macrophage density</b>			
Low	6.87		
High	34.43	0.374 (0.145, 0.966)	<b>0.0422*</b>
<b>Intratumoural CD38<sup>-</sup>CD68<sup>+</sup> macrophage density</b>			
Low	12.65	Reference	
High	8.41	1.336 (0.582, 3.066)	0.4946
<b>Intratumoural CD38<sup>+</sup>CD68<sup>-</sup> cells density</b>			
Low	6.87	Reference	
High	10.53	0.895 (0.391, 2.049)	0.7926
<b>Intratumoural CD8<sup>+</sup> T cell density</b>			
Low	7.03	Reference	
High	12.65	0.769 (0.304, 1.946)	0.5792
<b>PD-L1 tumour proportion score (TPS)</b>			
< 1	13.96	Reference	
≥ 1%	7.03	1.076 (0.454, 2.553)	0.8672

\*P<0.05 indicated a statistically significant difference. # PD-L1, programmed death-ligand 1; mOS, median overall survival.

**Supplementary Table 7.** Analysis of mPFS and mOS in patients with viral-related HCC (n=31) treated with anti PD-1/PD-L1 single agent.

<b>Progression free Survival (PFS)</b>			
Factor	mPFS (months)	OR (95% CI)	P-value
Stage			
B (n=3)	1.317	Reference	
C (n=28)	2.701	0.558 (0.163, 1.907)	0.3518
Age			
<65 (n= 16)	1.318	Reference	
≥65 (n=15)	3.887	0.776 (0.358, 1.678)	0.5185
AFP marker			
<400 (n=22)	2.701	Reference	
≥400 (n=9)	1.515	1.231 (0.530, 2.863)	0.6289
ECOG			
0 (n=22)	2.701	Reference	
≥ 1 (n=9)	1.746	1.090 (0.454, 2.614)	0.8475
Child-Pugh score			
A5 (n = 17)	5.271	Reference	
A6 (n=12)	1.515	1.233 (0.530, 2.870)	0.6273
A6, B7, and B8 (n=2)	1.515	1.460 (0.658, 3.243)	0.3521
Macrovascular invasion			
Yes (n=9)	2.701	Reference	
No (n=22)	1.746	1.689 (0.670, 4.256)	0.2667
Extra-hepatic spread			
Yes (n=23)	5.271	Reference	
No (n=7)	1.285	4.019 (1.546, 10.444)	0.0043
<b>Intratumoural total CD38<sup>+</sup> cell proportion</b>			
Low (n=20)	1.680	Reference	
High (n=11)	19.141	0.254 (0.097, 0.663)	<b>0.0051*</b>
<b>Intratumoural CD38<sup>+</sup>CD68<sup>+</sup> macrophage density</b>			
Low (n=8)	1.680	Reference	
High (n=23)	5.535	0.338 (0.137, 0.854)	<b>0.0217*</b>
<b>Overall Survival (OS)</b>			

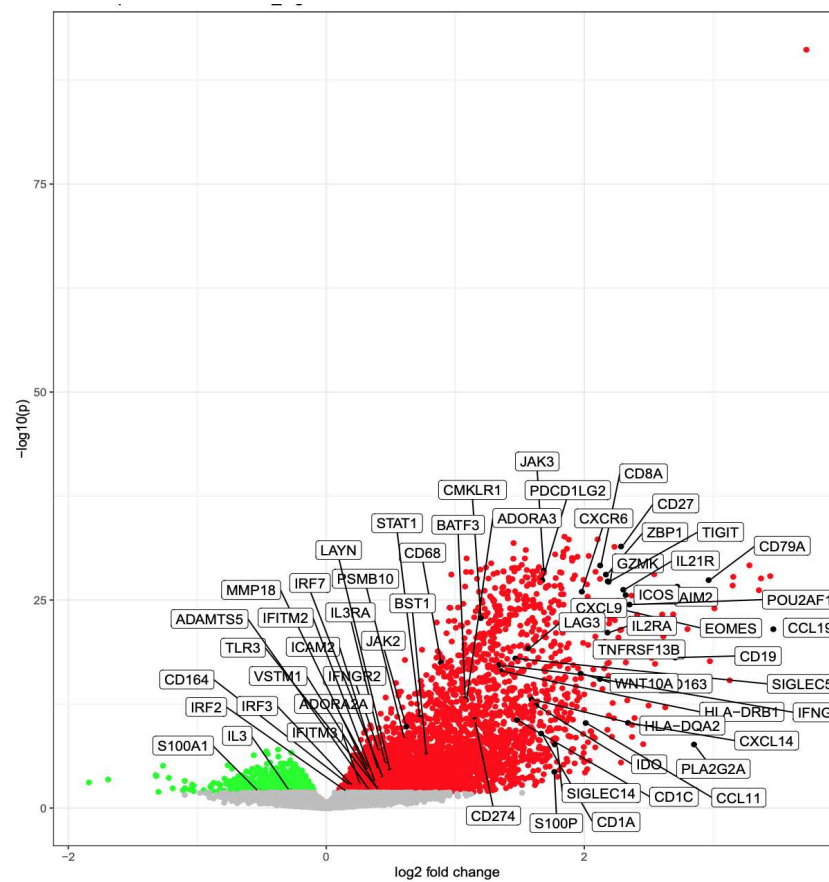
Factor	mOS (months)	OR (95% CI)	P-value
<b>Stage</b>			
B (n=3)	7.874	Reference	
C (n=28)	9.656	0.576 (0.167, 1.985)	0.3823
<b>Age</b>			
<65 (n= 16)	7.050	Reference	
≥65 (n=15)	14.232	0.949 (0.417, 2.162)	0.9017
<b>AFP marker</b>			
<400 (n=22)	8.335	Reference	
≥400 (n=9)	18.581	0.920 (0.375, 2.259)	0.8563
<b>ECOG</b>			
0 (n=22)	9.686	Reference	
≥ 1 (n=9)	6.885	1.403 (0.567, 3.471)	0.4643
<b>Child-Pugh score</b>			
A5 (n = 17)	15.879	Reference	
A6 (n=12)	5.106	1.448 (0.576, 3.639)	0.4313
A6, B7, and B8 (n=14)	5.139	1.715 (0.724, 4.065)	0.2204
<b>Macrovascular invasion</b>			
Yes (n=9)	9.686	Reference	
No (n=22)	8.434	1.147 (0.448, 2.935)	0.7747
<b>Extra-hepatic spread</b>			
Yes (n=23)	15.879	Reference	
No (n=7)	5.139	3.799 (1.425, 10.126)	0.0076
<b>Intratumoural total CD38<sup>+</sup> cell proportion</b>			
Low (n=20)	7.050	Reference	
High (n=11)	34.525	0.334 (0.121, 0.919)	<b>0.0337*</b>
<b>Intratumoural CD38<sup>+</sup>CD68<sup>+</sup> macrophage density</b>			
Low (n=8)	8.433	Reference	
High (n=23)	15.879	0.492 (0.198, 1.222)	0.1264

**Supplementary Table 8.** Analysis of mPFS and mOS in patients with non-viral related HCC (n=18) treated with anti PD-1/PD-L1 single agent.

<b>Progression free Survival (PFS)</b>			
Factor	mPFS (months)	OR (95% CI)	P-value
<b>Stage</b>			
B (n=2)	1.581	Reference	
C (n=16)	1.680	1.082 (0.239, 4.893)	0.9186
<b>Age</b>			
<65 (n= 4)	1.614	Reference	
≥65 (n=14)	1.680	0.602 (0.180, 2.008)	0.4086
<b>AFP marker</b>			
<400 (n=11)	1.614	Reference	
≥400 (n=7)	3.624	0.579 (0.207, 1.616)	0.2963
<b>ECOG</b>			
0 (n=13)	1.680	Reference	
≥ 1 (n=5)	1.581	1.282 (0.436, 3.774)	0.6517
<b>Child-Pugh score</b>			
A5 (n = 10)	1.614	Reference	
A6 (n=7)	2.998	0.864 (0.306, 2.440)	0.7821
A6, B7, and B8 (n=8)	2.998	0.974 (0.361, 2.627)	0.9587
<b>Macrovascular invasion</b>			
Yes (n=5)	4.052	Reference	
No (n=13)	1.614	2.302 (0.728, 7.274)	0.1556
<b>Extra-hepatic spread</b>			
Yes (n=13)	1.680	Reference	
No (n=5)	1.680	0.637 (0.200, 2.031)	0.4457
<b>Intratumoural total CD38<sup>+</sup> cell proportion</b>			
Low (n=8)	2.998	Reference	
High (n=10)	1.680	0.727 (0.254, 2.080)	0.5528
<b>Intratumoural CD38<sup>+</sup>CD68<sup>+</sup> macrophage density</b>			
Low (n=4)	1.548	Reference	
High (n=14)	3.624	0.341 (0.099, 1.182)	0.0898

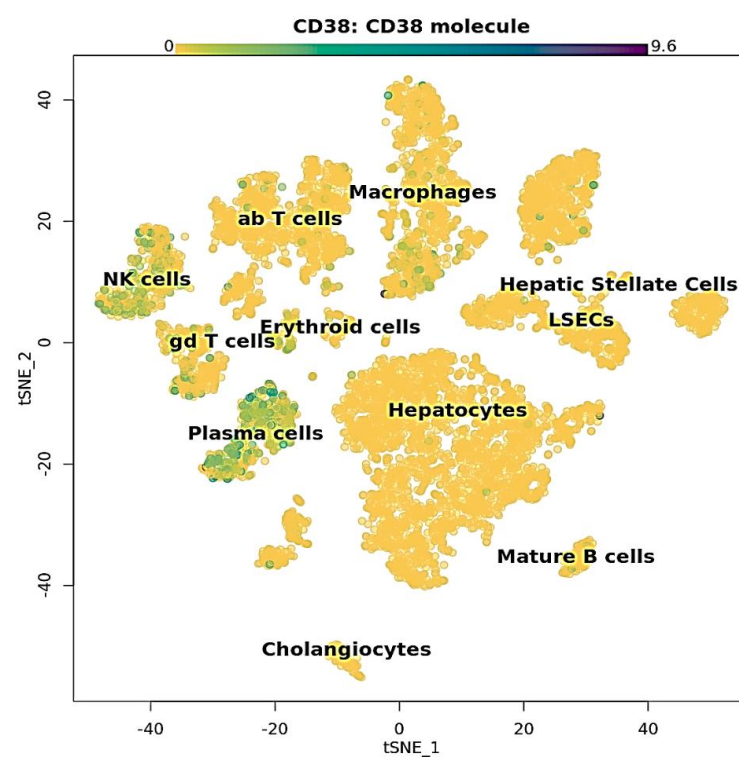
Overall Survival (OS)			
Factor	mOS (months)	OR (95% CI)	P-value
<b>Stage</b>			
B (n=2)	22.633	Reference	
C (n=16)	12.684	3.154 (0.397, 25.080)	0.2776
<b>Age</b>			
<65 (n= 4)	9.620	Reference	
≥65 (n=14)	14.001	0.621 (0.164, 2.362)	0.4849
<b>AFP marker</b>			
<400 (n=11)	14.001	Reference	
≥400 (n=7)	18.976	0.865 (0.268, 2.788)	0.8077
<b>ECOG</b>			
0 (n=13)	14.001	Reference	
≥ 1 (n=5)	Not reached	0.563 (0.121, 2.606)	0.4620
<b>Child-Pugh score</b>			
A5 (n = 10)	14.001	Reference	
A6 (n=7)	12.684	1.134 (0.329, 3.910)	0.8425
A6, B7, and B8 (n=8)	12.684	1.366 (0.429, 4.353)	0.5977
<b>Macrovascular invasion</b>			
Yes (n=5)	22.633	Reference	
No (n=13)	14.001	1.688 (0.443, 6.429)	0.4432
<b>Extra-hepatic spread</b>			
Yes (n=13)	9.620	Reference	
No (n=5)	22.633	0.489 (0.127, 1.894)	0.3010
<b>Intratumoural total CD38<sup>+</sup> cell proportion</b>			
Low (n=8)	14.528	Reference	
High (n=10)	12.684	0.676 (0.204, 2.246)	0.5229
<b>Intratumoural CD38<sup>+</sup>CD68<sup>+</sup> macrophage density</b>			
Low (n=4)	2.504	Reference	
High (n=14)	14.528	0.342 (0.085, 1.385)	0.1327

**Supplementary Figure 1** High resolution volcano plot of the genes that are differentially expressed between high and low CD38 HCC (as seen in Fig. 1C)



**Supplementary Figure 2** Single cell CD38 gene expression levels in the human liver.

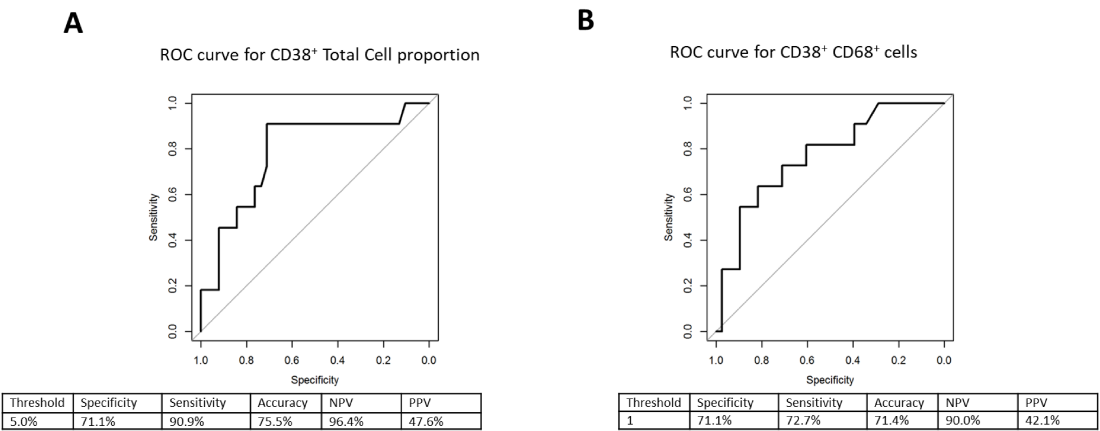
tSNE dimension reduction plot showing the expression of CD38 in the various identified cell populations. Low expression is shown in yellow, while high expression is shown in purple.



**Supplementary Figure 3** Receiver operating characteristic curves used to predict responders.

(A) Receiver operating characteristic curve for CD38<sup>+</sup> immune cells (AUC=0.785). (B) Receiver operating characteristic curve for CD38<sup>+</sup> CD68<sup>+</sup> macrophages (AUC=0.768).

*Sensitivity refers to the proportion of true positive subjects with the disease among subjects with disease. Specificity refers to the proportion of true negative subjects without the disease among subjects without disease. PPV refers to the proportion of patients with positive results among subjects with positive results. NPV refers to the proportion of subjects without disease with a negative result among subjects with negative results. Accuracy refers to the proportion of subjects correctly classified among all subjects. AUC, area under the curve; PPV, positive predictive value, NPV, negative predictive value. PFS, progression free survival.*

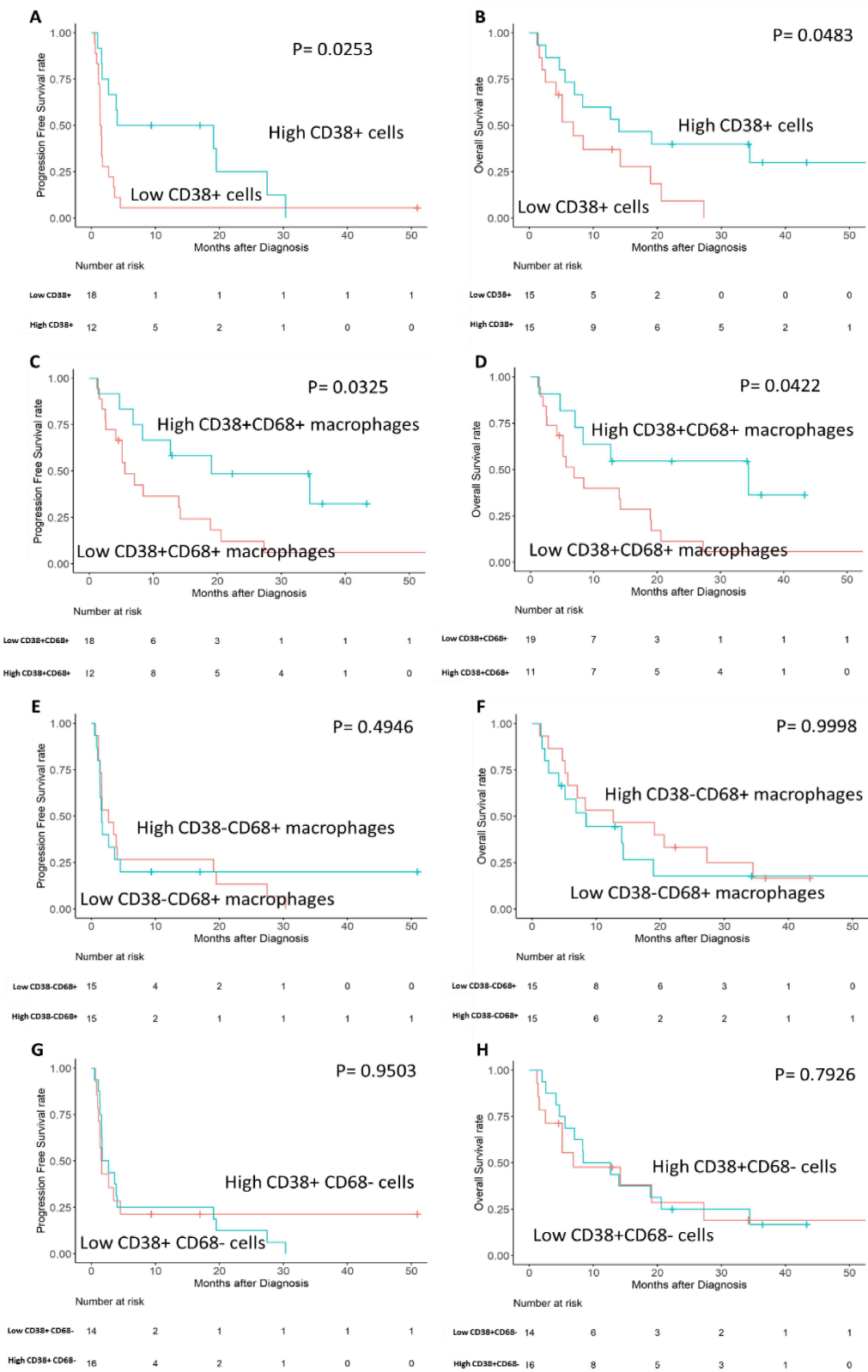




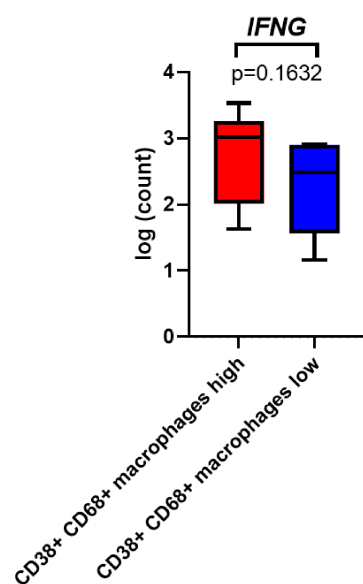
**Supplementary Figure 4** Response to anti-PD-1/PD-L1 single agent therapy in patients with hepatocellular carcinoma, in relation to intratumoural total CD38<sup>+</sup> CD68<sup>+</sup> macrophage density and CD38<sup>+</sup> CD68<sup>-</sup> cells density.

(A) The total CD38<sup>+</sup> cell proportion within the tumours of responders and non-responders treated with anti-PD-1/PD-L1 single agent. (B) Kaplan Meier curve showing the association between a high total CD38<sup>+</sup> cell proportion and improved PFS after treatment with anti-PD-1/PD-L1 single agent. (C) Kaplan Meier curve showing the association between a high total CD38<sup>+</sup> cell proportion and improved OS after treatment with anti-PD-1/PD-L1 single agent. (D) The CD38<sup>+</sup> CD68<sup>+</sup> macrophage density of responders and non-responders treated with anti-PD-1/PD-L1 single agent. (E) Kaplan Meier curve showing the association between high CD38<sup>+</sup> CD68<sup>+</sup> macrophage density and improved PFS after treatment with anti-PD-1/PD-L1 single agent. (F) Kaplan Meier curve showing the association between high CD38<sup>+</sup> CD68<sup>+</sup> macrophage density and improved OS after treatment with anti-PD-1/PD-L1 single agent. (G) Kaplan Meier curve showing the association between high CD38<sup>+</sup> CD68<sup>-</sup> cells density and improved PFS after treatment with anti-PD-1/PD-L1 single agent. (H) Kaplan Meier curve showing the association between high CD38<sup>+</sup> CD68<sup>-</sup> cells density and improved OS after treatment with anti-PD-1/PD-L1 single agent.

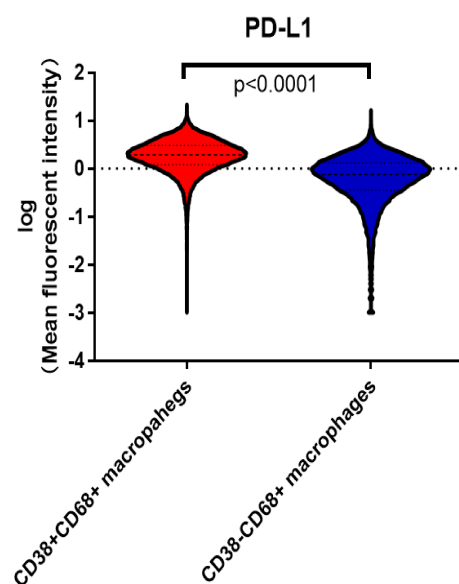
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**Supplementary Figure 5.** IFNG gene level showed a trend higher in HCC patients that harboured higher CD38<sup>+</sup> CD68<sup>+</sup> macrophages

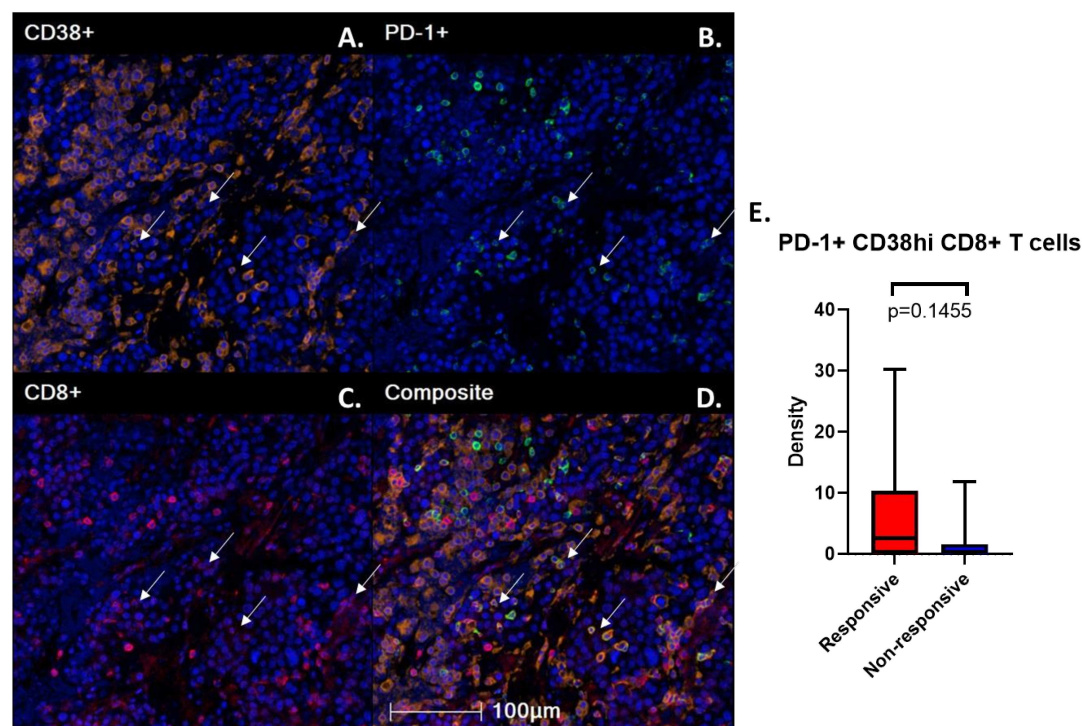


**Supplementary Figure 6.** The mean fluorescent intensity of PD-L1 on CD38<sup>+</sup>CD68<sup>+</sup> macrophages are significantly higher than the CD38<sup>-</sup>CD68<sup>+</sup> macrophages in HCC

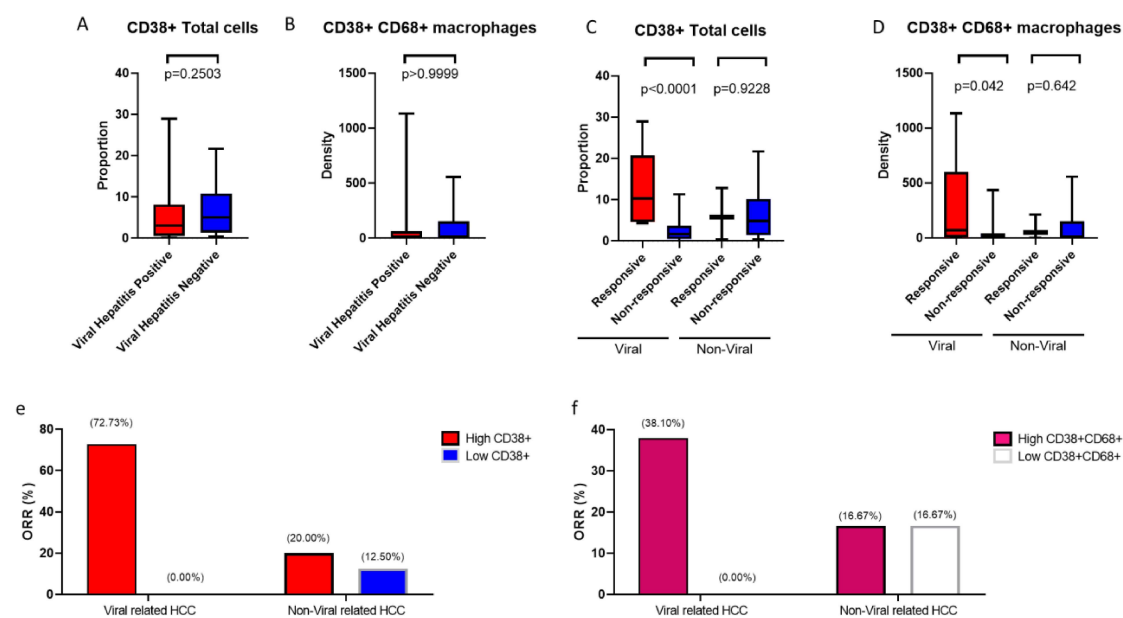


**Supplementary Figure 7.** (A-D) mIHC/IF revealed that CD38 (orange), PD-1 (green) and the T- cell lymphocyte marker CD8 (red) were expressed in the HCC TME. CD8 (red) is co-localised with PD-1 (green) and CD38 (orange) in the HCC TME. Cell nuclei are counterstained with DAPI for mIHC/IF (blue). Images are shown at a magnification of 400X for A, B, C, and D. (E) There is no statistical significance ( $p=0.1455$ ) of the density of CD38<sup>+</sup>PD-1<sup>+</sup>CD8<sup>+</sup> cells between responsive and non-responsive patients.

*HCC, hepatocellular carcinoma; TME, tumour microenvironment, IHC, immunohistochemistry; mIHC/IF, multiplex immunohistochemistry/immunofluorescence.*

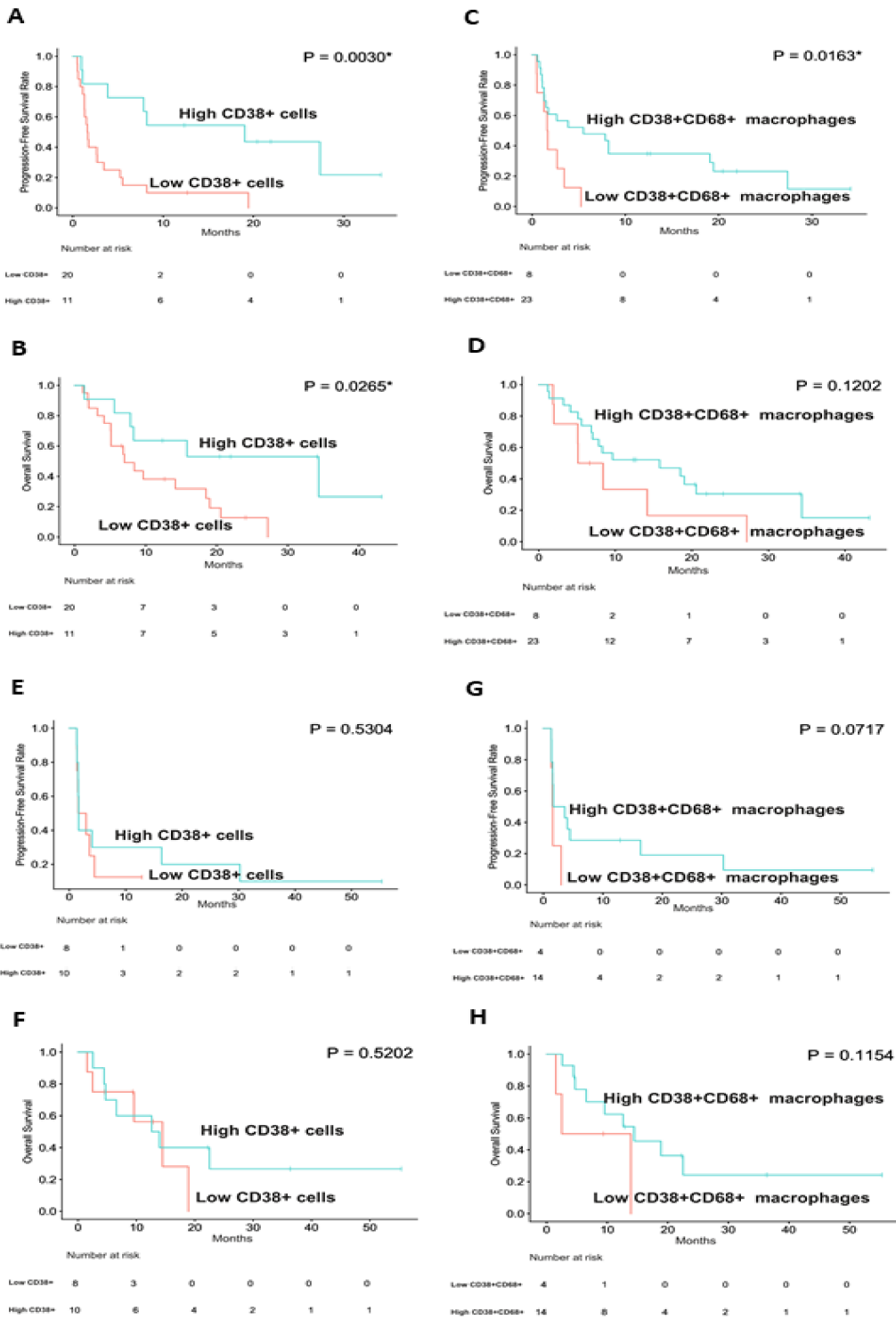


**Supplementary Figure 8.** Abundance of (A) CD38<sup>+</sup> cells and (B) CD38<sup>+</sup>CD68<sup>+</sup> macrophages between patients with or without viral hepatitis in our cohort (31 vs 18). The proportion of (C) CD38<sup>+</sup> cells is associated with responsiveness to ICB in the viral-related HCC but not in the non-viral related HCC. (D) The density of CD38<sup>+</sup>CD68<sup>+</sup> macrophages is associated with responsiveness to ICB in the viral-related HCC but not in the non-viral related HCC. (E-F) Overall response rates of each biomarker in the viral-related HCC and the non-viral related HCC, respectively.



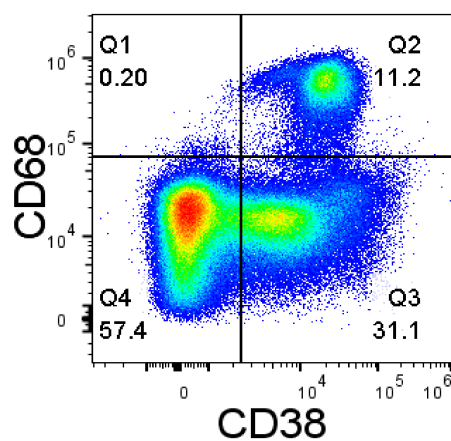
**Supplementary Figure 9.** (A) Kaplan Meier curve showing the association between a high total CD38<sup>+</sup> cell proportion and improved PFS after treatment with ICB in viral-related HCC. (B) Kaplan Meier curve showing the association between a high total CD38<sup>+</sup> cell proportion and improved OS after treatment with ICB in viral-related HCC. (C) Kaplan Meier curve showing the association between high CD38<sup>+</sup> CD68<sup>+</sup> macrophage density and improved PFS after treatment with ICB in viral-related HCC. (D) Kaplan Meier curve showing the association between high CD38<sup>+</sup> CD68<sup>+</sup> macrophage density and improved OS after treatment with ICB in viral-related HCC. (E) Kaplan Meier curve showing the association between a high total CD38<sup>+</sup> cell proportion and improved PFS after treatment with ICB in non-viral-related HCC. (F) Kaplan Meier curve showing the association between a high total CD38<sup>+</sup> cell proportion and improved OS after treatment with ICB in non-viral-related HCC. (G) Kaplan Meier curve showing the association between high CD38<sup>+</sup> CD68<sup>+</sup> macrophage density and improved PFS after treatment with ICB in non-viral-related HCC. (H) Kaplan Meier curve showing the association between high CD38<sup>+</sup> CD68<sup>+</sup> macrophage density and improved OS after treatment with ICB in non-viral-related HCC. ICB, immune checkpoint blockade.

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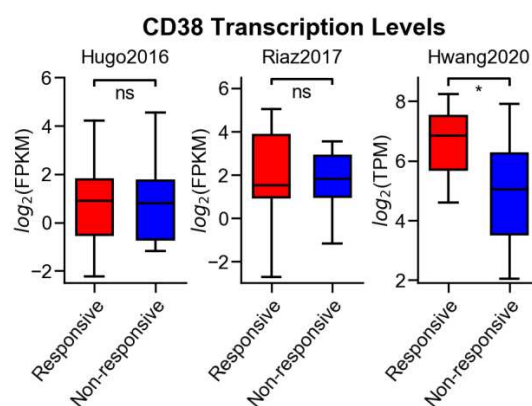




**Supplementary Figure 10.** Co-expression of CD38 and CD68 observed in HCC PBMC. The representative pseudocolour plot was gated from the total single live cell population. HCC, hepatocellular carcinoma.



**Supplementary Figure 11.** CD38 transcription levels in pre-anti-PD-1 therapy melanoma samples as measured by RNA-sequencing. In two melanoma cohorts (Hugo2016 and Riaz2017<sup>3,4</sup>) and one NSCLS cohort (Hwang2020<sup>5</sup>), patients were classified as being responsive (R) or non-responsive (NR) to anti-PD-1 therapy (Hugo2016: R n=15, NR n=13, p-value=0.925; Riaz2017: R n=26, NR n=23, p-value=0.226; Hwang2020: R n=9, NR n=12, p-value=0.032). Transcription levels of CD38 were retrieved based on publicly available RNA-seq data from these studies. Two RNA-seq datasets from Hugo2016 were from different sites in the same patient. One RNA-seq dataset from Riaz2017 was dropped as the CD38 transcription level was not recorded. p-value was calculated by a two-tailed t-test.



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