Tumor-associated carbohydrates and immunomodulatory lectins as targets for cancer immunotherapy

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To cite: Rodrigues Mantuano N, Natoli M, Zippelius A, *et al.* Tumor-associated carbohydrates and immunomodulatory lectins as targets for cancer immunotherapy. *Journal for ImmunoTherapy of Cancer* 2020;8:e001222. doi:10.1136/jitc-2020-001222

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Accepted 28 August 2020

ABSTRACT

During oncogenesis, tumor cells present specific carbohydrate chains that are new targets for cancer immunotherapy. Whereas these tumor-associated carbohydrates (TACA) can be targeted with antibodies and vaccination approaches, TACA including sialic acid-containing glycans are able to inhibit anticancer immune responses by engagement of immune receptors on leukocytes. A family of immune-modulating receptors are sialic acid-binding Siglec receptors that have been recently described to inhibit antitumor activity mediated by myeloid cells, natural killer cells and T cells. Other TACA-binding receptors including selectins have been linked to cancer progression. Recent studies have shown that glycan-lectin interactions can be targeted to improve cancer immunotherapy. For example, interactions between the immune checkpoint T cell immunoglobulin and mucin-domain containing-3 and the lectin galectin-9 are targeted in clinical trials. In addition, an antibody against the lectin Siglec-15 is being tested in an early clinical trial. In this review, we summarize the previous and current efforts to target TACA and to inhibit inhibitory immune receptors binding to TACA including the Siglec-sialoglycan axis.

INTRODUCTION

Recent advancements in the stimulation of the immune microenvironment and anticancer immune responses with immune checkpoint inhibitors (ICI) has improved the outcome of treatments for patients and has led to impressive long-term remissions in some patients with advanced disease.^{1–5} However, primary and acquired resistance significantly diminish the success of ICI and only a minority of patients benefit from currently available cancer immunotherapies.^{6,7} Thus, new strategies are urgently needed in order to induce long-term remissions with cancer immunotherapy in many more of our patients.

Carbohydrates belong to the major biomolecules of living organisms. Carbohydrates can be attached to proteins (glycoproteins), lipids and exist as chains of carbohydrates (glycosaminoglycans). Glycans—carbohydrate-containing macromolecules—are ubiquitous in biological systems and are essential for

numerous biological functions.^{8–10} Cell surfaces and extracellular proteins are significantly glycosylated. In addition, glycosaminoglycans can be found in the extracellular matrix. Glycans are used as storage for energy (glycogen), are structurally important (see later for the stability of programmed cell death protein 1 (PD-1)) and can mediate signals. Whereas proteins undergo substantial post-translational modifications, in particular N-glycosylation and O-glycosylation,^{8–10} intracellular modification of tyrosine with O-GlcNAc serves for intracellular signaling.¹¹ Changes of glycosylation have a significant impact on cancer biology and cancer progression.^{12–15} Of note, altered glycan structures represent antigenic targets for cancer immunotherapy. In this review, we summarize how cancer-associated changes in glycosylation can be used to improve cancer immunotherapy.

CANCER-SPECIFIC CHANGES IN GLYCOSYLATION

Altered glycosylation is a common feature of tumor cells and leads to the formation of tumor-associated carbohydrates (TACA) (figure 1). Three common changes are often associated with cancer: a) increased expression of truncated or incomplete glycans, b) increased branching of N-glycans and c) augmented or changed presence of sialic acid-containing glycans.¹⁵ Tumor-cell-surface glycans are known to promote cancer progression by affecting tumor growth, cell invasiveness and negatively regulate immune responses.^{15–17} Changes of glycosylation observed in cancer depend on the expression and changes of enzymes involved in glycan biosynthesis and glycan-modifying enzymes including transferases and glycosidases as well as transporter for saccharides and precursors.^{12,14,18} Expression of these glycan-modifying proteins are altered in cancer due

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Figure 1 Overview on cancer-associated glycosylation. Three main changes can be found in cancer that are regulated by genetic or epigenetic alterations in genes of glycan-modifying enzymes or enzymes involved in carbohydrate biosynthesis. N-glycans show often an increased branching due to increased MGAT5 expression. Another often observed change is the truncation of O-glycans and the exposure of new tumor-associated carbohydrates (TACA) including the T antigen, Tn antigen and the sialyl-Tn antigen (STn). In addition, changes of sialylation of both glycoproteins and glycolipids can be observed. Increased sialylation (hypersialylation) is often observed. The introduction of the non-human sialic acid Neu5Gc can also be observed. Fuc, fucose; GlcNAc, N-acetyl-glucosamine; Gal, galactose; GalNAc, N-acetyl-galactosamine; Glc, glucose; Man, mannose; N-acetyl-neuraminic acid; Neu5Gc, N-glycosyl-neuraminic acid.

to genetic and epigenetic alterations and differ between cancer types.

Changes in sialylation

Cancer-associated glycans often exhibit an increased amount of sialic acid. Augmented sialylation of tumor cells has been correlated with a metastatic phenotype and poor prognosis in patients with cancer.^{19–20} Sialic acids are predominantly found at the non-reducing end of N-linked and O-linked glycans attached to proteins or glycolipids.²¹ Hypersialylation facilitates interactions with sialic acid binding receptors such as selectins and Siglecs with consequences for cancer progression. Moreover, the incorporation of the non-human sialic acid N-glycolyl-neuraminic acid (Neu5Gc) into glycans and the interaction with circulating anti-Neu5Gc antibodies influences cancer progression.^{22–26} Neu5Gc is biosynthesized from N-acetyl-neuraminic acid (Neu5Ac) via the enzyme CMP-Neu5Ac hydroxylase (CMAH), which is not present in humans.²⁷ However, various studies have found an increased presence of Neu5Gc-containing glycans in cancer, which could be associated with uptake of meat from Neu5Gc producing mammals.^{28–29}

Truncation of O-glycoproteins

O-glycans are ubiquitously found on cells and are particularly secreted into the extracellular matrix or into the

lumen of internal organs. For example, mucins, highly O-glycosylated proteins such as CA19-9,³⁰ can serve as a biomarker for some cancer types. One of the most common cancer-associated changes in glycosylation is the truncation of O-linked carbohydrate chains (figure 1).³¹ Usually, a GalNAc sugar residue is attached to a serine or threonine of the glycoprotein (GalNAc α 1-O-Ser/Thr, Tn antigen) and usually elongated by the T-synthase (core 1 β 3-galactosyltransferase) in the Golgi apparatus that attaches a galactose residue to Tn antigen. The resulting glycan is called T antigen (sometimes also called the Thomsen-Friedenreich (TF) antigen). The T-synthase requires a chaperone for the correct folding and enzymatic activity.³¹ The chaperone was termed Core 1 β 3-galactosyltransferase Specific Molecular Chaperone (COSMC).³² The X linked *COSMC* gene is mutated in various cancer types leading to the presence of Tn antigen or its sialylated form, the sialyl-Tn (STn) antigen.³¹ Interestingly, truncated O-glycosylation is shown to have an immunomodulatory effect. Tn antigen binds to macrophage galactose-type lectin on dendritic cells and macrophages that inhibits the migration of immature antigen-presenting cells (APCs) and increases M2-like tumor associated macrophages.^{33–35}

Truncated O-glycans represent epitopes which may selectively target cancer cells. Various cancer tissues have been analyzed for the expression for the T, Tn and STn antigen.^{36–38} The human-mucin 1 (MUC1) is over-expressed in many adenocarcinomas, presenting high levels of truncated glycans as STn-MUC1, Tn-MUC1 and T-MUC-1. Yet, these antigens are rarely expressed in normal tissue compared with cancer tissue.^{39–44} Further studies are needed to determine the specificity of the expression patterns if we consider to target these epitopes with antibodies and chimeric antigen receptors (CARs).

Altered branching of N-glycoproteins

Increased branching of N-glycans, mediated by β 1,6-N-acetylglucosaminyltransferase V (MGAT5, figure 1),^{12–15} can influence cell adhesion, migration and metastasis of tumor cells.^{45–46} Upregulation of MGAT5 has also been shown to directly influence cytokine signaling and tumor progression⁴⁷ while the knockdown of *Mgat5* led to activation of CD4⁺ T cells and macrophages in breast cancer.⁴⁸ Altered N-glycosylation of immune cells could also affect the antitumor immune response. Increased branching of N-glycans can directly inhibit T cell activation by increasing T cell receptor (TCR) clustering.^{49–50} This effect was attributed to interaction with galectin-3.⁴⁹ On the other side, TCR signaling also directly influences enzymes modulating N-glycosylation.⁵¹

Alterations in glycolipids

Gangliosides are sialylated glycan-containing lipids of the cell membrane show also often changes on cancer cells. The gangliosides GM3, GM2, CD3 and GD2 are present in normal tissue but are often overexpressed in different cancers including lung cancer, melanoma and neurogenic

tumors such as neuroblastoma.^{52–55} Glycolipids can significantly influence cell signaling by mediating the formation of lipid rafts.⁵⁶ Tumor-associated gangliosides have been investigated for their immunosuppressive properties and role in cancer progression. Furthermore, the plasma concentration of gangliosides is often elevated,⁵⁷ making them potential therapeutic targets and diagnostic tools. GM3 contains sialic acid-residues and several studies have shown that GM3 containing the non-human sialic acid Neu5Gc (Neu5Gc-GM3) is relatively specific for different types of cancer.²⁹

Cancer-associated changes in glycosylation and immune phenotypes

The association of specific glycan changes with the immune state of a cancer is currently being studied. Immune phenotypes such as T cell excluded tumors were associated recently with galectin-1 expression and interactions with glycan-ligands.⁵⁸ The Cancer Genome Atlas has been used to study the expression of different glycan-modifying enzymes, for example, for sialic acid-modifying enzymes.⁵⁹ However, further systematic studies including also lectin stainings are needed on tissue sections to correlate immune phenotypes with.

MODALITIES TO TARGET TUMOR-ASSOCIATED GLYCANS

ADCC and CDC

Antibody-dependent cellular cytotoxicity (ADCC) or complement-dependent cytotoxicity (CDC) is triggered by the interaction between antibody-bound target cells—for example, infected or tumor cells—and effector immune cells or complement factors, respectively. Several glycan-targeted monoclonal antibodies (mAbs), which are in clinical use or development, are known to elicit ADCC and/or CDC (figure 2). Among these, dinutuximab, which targets ganglioside GD2 on melanoma, and neuroblastoma (table 1),^{60–62} is approved for the treatment of high-risk neuroblastoma pediatric patients.⁶³ Similarly, KW871, a chimeric mAb which targets the ganglioside GD3, exhibited antitumor activity in combination with IFN α 2b in vitro.⁶⁴ In patients with metastatic melanoma, the combination of KW871 with interferon was shown to be well tolerated, although not highly efficacious.⁶⁵ An anti-idiotypic antibody was generated to elicit an immune response against Neu5Gc-GM3.⁶⁶ Early trials have shown interesting activity in patients with non-small cell lung cancer (NSCLC).^{66–67} Trials testing the efficacy of this antibody called racotumomab in a larger population are currently recruiting (eg, NCT01460472).

Since the 1990s, several other mAbs have been designed that target Lewis antigens (Le), expressed by a broad range of tumor cells, and that are able to elicit ADCC and/or CDC in preclinical and clinical studies. Examples of such mAbs are BR96,⁶⁸ targeting Le^y; hu3S193,⁶⁹ a humanized anti-Le^y mAb which showed low toxicity in an early clinical trial,⁷⁰ but insufficient efficacy in a subsequent phase II trial.⁷¹

Figure 2 Overview on targeting approaches for cancer-associated glycosylation. (A) Tumor-associated carbohydrates (TACA) can serve as tumor-specific antigen and be approached with antibody-dependent cellular cytotoxicity (ADCC)-inducing or complement-dependent cytotoxicity (CDC)-inducing antibodies or with antibodies carrying a payload (chemotherapy or even enzyme). (B) Chimeric antigen receptor (CAR) expressing immune cells could be redirected towards TACA-presenting tumors. Similarly, bispecific antibodies could be used to direct immune cells to TACA-expressing tumors (not shown). (C) TACA such as sialoglycans can engage immune receptors including inhibitory Siglec receptors on T cells and myeloid cells and improve anticancer immunity directly. ADC, antibody-drug conjugates.

As for targeting of tumor-associated MUC1 (TA-MUC1) expressed on several malignancies including ovarian, breast and cervical cancers,⁷² the mAb PankoMab-GEX (gatipotuzumab) has shown therapeutic efficacy, particularly in heavily pretreated patients with ovarian cancer.^{73–74} Yet, the subsequent phase II trial did not show any outcome advantage.⁷⁵ Another high-affinity mAb named 5E5 targets the aberrant Tn glycoform of mucin MUC1 and has been shown to lyse breast cancer cells via both ADCC and CDC.⁷⁶

Glycosylation is also under investigation to be used as tool for the enhancement of ADCC and CDC immune response mechanisms. Indeed, manipulation of specific residues of the Fc N-glycan has been shown to modulate antibody-dependent effector functions via modification of the Fc binding affinity to Fc receptors expressed on different immune cells. Particularly, core fucosylation and sialylation of Fc regions results in decreased ADCC, while N-glycans with low or no sialic acid are better suited to trigger ADCC. These approaches are reviewed by Wang and Ravetch⁷⁷ and Mastrangeli and colleagues.⁷⁸

Antibody-drug conjugates

Antibody-drug conjugates (ADCs) unify the properties of cytotoxic chemotherapy and mAbs, in an effort to selectively target and lyse cancer cells. Similarly, antiglycan-directed antibodies can be exploited to deliver selected anticancer payloads specifically to tumor cells (figure 2). First attempts used the previously mentioned anti-Le^y BR96 mAb, conjugated to doxorubicin and docetaxel. This ADC was tested in phase II trials for advanced NSCLC

Table 1 Previous and ongoing clinical trials targeting tumor-associated glycans or lectins

Target	Drug candidate	Modality	Phase	Cancer type	Citation/Trial ID
NeuGcGM3	Racotumomab	Vaccine	II/III	NSCLC	(^{66 67}), NCT01460472
Lewis-Y antigen	hu3S193	mAb	I/II	Advanced epithelial cancer; platinum resistant/ refractory ovarian, Fallopian tube and primary peritoneal carcinoma	(^{70 71})
MUC1	PankoMab-GEX (Gatipotuzumab)	mAb	I/II	Advanced carcinoma; recurrent ovarian carcinoma	(^{74 75})
Lewis-Y antigen	SGN-15	ADC	II	NSCLC	(⁷⁹)
Lewis-Y antigen	BMS-182248-1	ADC	II	Metastatic breast cancer	(⁸⁰)
CD22/CD19	DT2219	BsAb	I	Refractory B-cell malignancies	(⁸⁹)
TAG72	CART72	CAR T	I	Advanced colorectal cancer	(⁹⁶)
Lewis-Y antigen	Le ^y -CAR T	CAR T	I	AML	(¹⁰²)
Lewis-Y antigen	Le ^y -CAR T	CAR T	I	Advanced solid tumors	NCT03851146
GD2	GD2-iCAR-PBT	CAR T	I	Metastatic melanoma	(¹⁰⁵)
GD2	GD2-CAR3 combined with lymphodepletion and PD-1 blockade	CAR T	I	Relapsed or refractory neuroblastoma	(¹⁰⁶)
IL15-GD2	GINAKIT	CAR-NKT	I	Pediatric neuroblastoma	NCT03294954
STn	Theratope	ADC	III	Metastatic breast adenocarcinoma	(^{115 116})
Siglec-15	NC318	mAb	I/II	Advanced or metastatic solid tumors	NCT03665285

ADC, antibody-drug conjugates; AML, acute myeloid leukemia; mAb, monoclonal antibody; MUC1, mucin 1; NSCLC, non-small cell lung cancer; PD-1, programmed cell death protein 1.

(table 1)⁷⁹ and for metastatic breast cancer.⁸⁰ Although initial results were encouraging, the mAb was later discontinued, likely due to low efficacy. More recent attempts at exploiting tumor glycan expression for drug delivery include work by Sedlik *et al*, who used a Tn-directed mAb (Chi-Tn) to deliver cytotoxic drugs; they showed promising antitumor activity of the chiTn ADC in vitro and in vitro, when this was conjugated to saporin or auristatin F.⁸¹ These effects were particularly dependent on high Tn expression in tumor cells.⁸¹ Similarly, Prendergast *et al* developed murine mAbs able to target tumor cells expressing STn with high avidity and further exploited a subset of these mAbs as ADCs by conjugating them to monomethyl auristatin E (MMAE).⁸² These ADCs showed high efficacy in vitro, in the presence of STn-expressing cancer cell lines, as well as tumor inhibition in multiple *in vivo* model, including breast and colorectal cancer.⁸² Furthermore, humanized anti-STn antibodies conjugated to MMAE have been shown to determine in vitro cytotoxicity specific to STn-expressing ovarian cancer cell lines as well as efficacy in tumor control in *in vivo* models for ovarian cancers, including in patient-derived xenografts.⁸³ These humanized aSTn-ADCs were further shown to present a low toxicity profile as they did not cross-react to any tissue of human origin. Most recently, the murine mAb FG129 and corresponding chimeric human variant CH129, were developed to target sialyl-di-Le^a-containing glycoproteins and shown to bind to a range of cancer tissues including colorectal, pancreatic, gastric and ovarian tumors.⁸⁴ Conjugation of CH129 to

either MMAE or maytansinoid (DM1 and DM4) resulted in cell death in vitro, as well as in vivo tumor control.⁸⁴

Bispecific antibodies

Bispecific antibodies (bsAbs) are molecules designed to recognize two distinct antigens or epitopes and have been recently emerging as potential key actors in cancer immunotherapy. In terms of tumor-associated glycans, research on bsAbs has so far focused on targeting GD2 ganglioside and MUC1 combined (table 1).^{85–88} Recent developments involve also targeting of glycan-binding lectins on tumor cells. bsAbs designed to target lectins including Siglecs are currently under investigation, for example, bsAb against CD22/CD19 on B cells—currently in a phase I–II trial, targeting B cell malignancies⁸⁹ or bsAb bridging CD33 on AML cells to CD3 on T cells^{90 91} or to CD16 on natural killer (NK) cells.⁹²

Redirection of immune cells

CARs can redirect immune cells to tumor cells or the tumor microenvironment by targeting tumor-specific antigens.^{93 94} However, design of CARs directed at solid cancers presents several challenges, due to the immunosuppressive mechanisms within the TME and the difficulty in finding antigens that are solely cancer-specific.⁹⁴

Targeting CARs to glycans may be advantageous due to the specificity of aberrant glycosylation on tumor cells. Several attempts at designing glycan-targeted CARs have been made and are currently being tested in preclinical and clinical studies (table 1). Tumor-associated glycoprotein

72 (TAG72) antigen is the truncated sialyl-Tn found on many O-glycoproteins and overexpressed by various types of cancer cells, including lung, colorectal and ovarian cancer cells. TAG72-directed CAR T cells were shown to be effective *in vitro* on gastrointestinal cancer cell lines.⁹⁵ A much more recent clinical trial using anti-TAG72 CAR T cells in advanced colorectal cancer patients failed to show effective clinical responses; this was potentially due to the murine origin of the scFv or to lack of co-stimulation⁹⁶; the second-generation TAG72-targeted CAR was shown to effectively target ovarian cancer cell lines and patient-derived primary ovarian cancer cells *in vitro*, as well as to reduce tumor growth and improve survival in mice, with sequential intraperitoneal administrations.⁹⁷ Attempts at designing MUC1-targeted CARs relied on two main antibodies, SM3 and HMFG2.⁹⁸ More recent Tn-MUC1-targeting CARs have been developed from the 5E5 mAb and shown to present cancer-specificity and weak reactivity against healthy tissues, as well as to effectively target and kill cancer cells in pancreatic and leukemia xenograft models.⁹⁹ A successful target for CAR T cell therapy is the TACA Le^y. A second generation fully humanized CAR construct targeting Le^y was shown to be effectively transduced in PBMC-derived T cells and to lyse Le^y positive tumor cell lines *in vitro*.¹⁰⁰ Similarly, *in vivo*, adoptive transfer of Le^y-targeted CAR T cells resulted in tumor homing and subsequently inhibiting growth of myeloma and ovarian cancer xenografts.^{100 101} The same CAR T cells were tested in four patients with relapsed AML, presenting Le^y positive blasts, in a phase I clinical trial.¹⁰² An ongoing Phase I clinical trial is now testing the safety and tolerability of using these CAR T cells in patients with advanced solid tumors presenting Le^y surface expression (NCT03851146).

In high-risk neuroblastoma patients with Epstein-Barr virus (EBV)-associated malignancies, infusion of EBV-specific and GD2 CAR-transduced T cells showed persistence of these cells and tumor necrosis in some patients,¹⁰³ with three patients showing complete responses and two long-term remission (up to 48 months) in a long-term follow-up study.¹⁰⁴ Combination of GD2-targeted CARs with PD-1 blockade is also under investigation, with studies showing that, *in vitro*, anti-PD-1 can rescue GD2 CARs from activation-induced cell death,¹⁰⁵ a phenomenon also observed *in vivo* in patients with metastatic melanoma treated with GD2-specific CAR T cells in a phase I clinical trial.¹⁰⁵ In another phase I clinical trial, combination of a third-generation GD2-targeted CAR with lymphodepletion and PD-1 blockade resulted in improved *in vivo* expansion of the CAR T cells, which were safely tolerated, but modest clinical responses.¹⁰⁶ A recent proof-of-concept study showed that incorporating IL-12 or IL-18 within a GD2-targeting CAR resulted in enhanced effector functions and increased monocyte recruitment *in vitro*.¹⁰⁷ Another phase I clinical trial testing IL15-GD2 CAR NKT cells in pediatric patients with neuroblastoma is currently ongoing (NCT03294954). More recent attempts at using immune cells other than T

cells for GD-2 targeted CARs include work by Mitwasi *et al*, who used a 'universal' CAR platform (UniCAR) to target NK cell line NK-92 to GD2 expressing cells.¹⁰⁸ UniCARs present an 'on/off system', which allows for controlling the activity of CAR expressing cells, depending on the presence of a target module, that is, an antibody-based cell-binding domain specific to GD2.¹⁰⁸

Vaccination

Several vaccines have been developed based on different glycans Tn-MUC1,¹⁰⁹ MUC16,¹¹⁰ GD2,^{111 112} GM2¹¹² and Neu5Gc-GM3,^{113 114} but most vaccines had major issues relating to the poor immunogenicity of glycans and glycopeptides. Glycan-based vaccine research is therefore currently focused on addressing these problems and enhancing immunogenicity of these immunotherapies. A strategy to increase immunogenicity is conjugation to carrier proteins; one such example is Theratope, an STn-based vaccine conjugated to carrier protein keyhole limpet hemocyanin. Though initial evaluations of Theratope in patients with metastatic adenocarcinomas produced encouraging results,¹¹⁵ including increased antibody titres and prolonged survival, this vaccine was later shelved after failing to meet its primary endpoints in a phase III clinical trial.¹¹⁶ Other strategies to enhance glycan immunogenicity include designing polyvalent vaccines which contain multiple glycan structures, such as Globo-H, STn, Tn, TF and Le^y¹¹⁷ or using peptide mimetics of TACAs, such as for SLe^x, SLe^a, SLe^y or Tn, to prevent enzymatic degradation and improve antitumor efficacy.^{118 119}

Glycan modifications can also be used to target shared or neo-antigens to APCs including dendritic cells.¹²⁰ Dendritic cells express lectins binding to glycan epitopes. For example, dendritic cells express the lectins Dendritic Cell-Specific Intercellular adhesion molecule-3-Grabbing Non-integrin (DC-SIGN) and Langerin that can be targeted with fucosylated glycans of Lewis-type oligosaccharides.¹²¹ In another work, Siglec-1 on macrophages was targeted to induce an immune response to a lipid antigen and robustly activate NK T cells.¹²²

TARGETING GLYCAN-RECEPTOR INTERACTIONS TO IMPROVE CANCER IMMUNOTHERAPY

Siglec-sialoglycan interactions

Recent evidence also suggests that cancer-associated glycosylation can directly influence anticancer immunity by binding to glycan binding receptors, which are also called lectins.^{12-14 123 124} Hypersialylated glycan structures have been identified in various types of cancer.^{14 22 24} The high density of sialoglycans on some tumor cells can engage inhibitory sialoglycan-binding receptors called Siglecs.^{25 125 126} Siglecs belong to the family of immunoglobulin (I)-type lectins.^{125 127 128} Siglecs are single-pass transmembrane proteins and bind sialoglycans through their carbohydrate-binding domain (table 2).^{125 127 128} In humans, there are 14 functionally active Siglec receptors

Table 2 Summary of human Siglec receptors, their expression on the surface of different cells, their intracellular signaling and their binding to sialic acid-containing glycan ligands

Siglec	Expression	Function	Binding specificity
Siglec-1, sialoadhesin	Macrophages, monocytes	No intracellular ITIM or ITIM-like motifs, supports phagocytosis	$\alpha 2,3 > \alpha 2,6$
Siglec-2, CD22	B cells	Inhibitory Siglec, inhibitory B cell receptor	$\alpha 2,6$
Siglec-3, CD33	Myeloid cells (including neutrophils, monocytes and progenitors)	Inhibitory Siglec (ITIM, ITIM-like)	$\alpha 2,6 > \alpha 2,3$
Siglec-4, myelin-associated glycoprotein	Myelin producing cells	No intracellular ITIM or ITIM-like motifs, important for	$\alpha 2,3 > \alpha 2,6$
Siglec-5	Monocytes, macrophages, neutrophils, activated T cells	Inhibitory Siglec (ITIM, ITIM-like), paired receptor with Siglec-14	$\alpha 2,3$
Siglec-6	Trophoblast, chronic lymphocytic leukemia cells	Inhibitory Siglec (ITIM, ITIM-like)	$\alpha 2,6$
Siglec-7	NK cells, intratumoral T cells	Inhibitory Siglec (ITIM, ITIM-like)	$\alpha 2,8 > \alpha 2,6 > \alpha 2,3$
Siglec-8	Eosinophils	Inhibitory Siglec (ITIM, ITIM-like)	$\alpha 2,3 > \alpha 2,6$
Siglec-9	Myeloid cells, NK cells, intratumoral T cells, dendritic cells	Inhibitory Siglec (ITIM, ITIM-like)	$\alpha 2,3 = \alpha 2,6$, broadly binding, also binding to some protein ligand
Siglec-10	B cells, T cells	Inhibitory Siglec (ITIM, ITIM-like)	$\alpha 2,3 = \alpha 2,6$
Siglec-11	Macrophages, microglia	Inhibitory Siglec (ITIM, ITIM-like), paired receptor with	$\alpha 2,8$
Siglec-XII	Epithelial cells, cancer cells	No binding to sialic acid-containing ligands (mutation of essential arginine in carbohydrate-recognition)	No binding to sialic acid ligands
Siglec-14	Monocytes, macrophages, neutrophils	Activating Siglec, positively charged amino acid mediates binding to DAP12, paired receptor with Siglec-5	$\alpha 2,3$ similar binding as its paired receptor Siglec-5
Siglec-15	Macrophages, dendritic cells, osteoclasts	Activating Siglec, positively charged amino acid mediates binding to DAP12	$\alpha 2,6$
Siglec-16	Macrophages, microglia	Activating Siglec, positively charged amino acid mediates binding to DAP12, paired receptor with Siglec-11	$\alpha 2,8$ similar binding as its paired receptor Siglec-11

ITIM, immunoreceptor tyrosine-based inhibitory motif.

that can be divided into two groups. On one side, there are evolutionary conserved Siglec receptors including Siglec-1, Siglec-2 (CD22), Siglec-4 and Siglec-15. On the other side, CD33-related Siglecs have rapidly evolved also within mammalian species.^{22 129} Siglec-3 (CD33), Siglec-5, Siglec-6, Siglec-7, Siglec-8, Siglec-9, Siglec-10, Siglec-11 and Siglec-14 belong to the CD33-related Siglecs and there are for most of them no direct orthologs, for example, in mice and humans.¹²⁸ Most Siglec receptors have intracellular domains containing immunoreceptor tyrosine-based inhibitory motifs (ITIM) or ITIM-like motifs. Engagement of these inhibitory receptors lead to a phosphorylation of Src homology region 2 domain-containing phosphatase (SHP)-1 and SHP-2, which inhibit cell activation.^{22 128} Most Siglec receptors are expressed on immune cells and engagement inhibits immune cell activation. Diversification of CD33-related Siglecs is probably due to interactions with pathogens exploiting inhibitory Siglec receptors by covering themselves with sialoglycans.¹³⁰ Some pathogens

as the group B streptococci have even developed proteins to engage inhibitory Siglecs on myeloid cells and thereby evade immune-mediated killing.¹³¹

The role of Siglec receptors as new targetable immune checkpoints has been recently reviewed in depth.^{23 25 125 126 132} Expression of inhibitory Siglec receptors on innate immune cells have been linked to inhibition of anticancer immunity.^{12 13 125} Siglec-7 is expressed on the majority of NK cells and Siglec-9 can be found on some subpopulations of NK cells.^{128 133} Upregulation of sialoglycans on cancer cells can inhibit NK cell-mediated tumor cell killing by engaging Siglec-7 and in some instances also Siglec-9.^{133 134} Siglec-9 expression has been shown to skew macrophage polarization to a protumorigenic phenotype and increased programmed death ligand 1 (PD-L1) expression on macrophages.¹³⁵ Siglec-9 engagement was shown to be dependent in this set of experiments on STn-modified mucins.¹³⁵ Other experiments in mice showed that myeloid polarization is affected by inhibitory Siglec

receptors including Siglec-E, the functional paralog of Siglec-9 in mice.¹³⁶ Recent evidence demonstrated that Siglec-10 on macrophages can act as a “don’t eat me” signal to inhibit phagocytosis.¹³⁷

Of note, the expression of Siglecs on adaptive immune cells including CD8⁺ T cells are inhibiting effective anticancer immunity as shown by us and others. Siglec-9 is upregulated on tumor-infiltrating T cells in various cancers including NSCLC, colorectal cancer, epithelial ovarian cancer and melanoma.^{138–139} Tumor growth was significantly enhanced in a transgenic mouse model of overexpression of Siglec-9 on T cells.¹³⁸ Siglec-9 was expressed mainly on tumor-specific T cells shown by a reduced TCR repertoire in tumor infiltrating CD8⁺ T cells.¹³⁹ Interestingly, Siglec-9 positive CD8⁺ T cells represent a less dysfunctional intratumoral T cell subtype and Siglec-9 blockade could reactivate these T cells.¹³⁸ The conserved Siglec-15 was identified as inhibitor of T cell activation on APCs.¹⁴⁰

Approaches to target sialoglycan-Siglec interactions could involve both blocking the Siglec receptor, for example, with an antibody. Siglec-15 was targeted with an antibody in preclinical mouse models¹⁴⁰ and is already in an early clinical trial for advanced solid tumors (NCT03665285). On the other side, the ligand could be targeted enzymatically with sialidases or with small molecules interfering with sialoglycan biosynthesis.^{141–142} The use of sialic acid analogs *in vivo* by intratumoral injection could block the sialoglycan biosynthesis and led to a strong activation intratumoral T cells,¹⁴¹ however the systemic application of these analogs would be quite toxic. Bacterial sialidase was tested in a system linked to the HER2-targeting antibody trastuzumab and NK cell-mediated tumor cell killing was tested *in vitro*.¹⁴² Recently, HER2-targeted bacterial sialidase was tested *in vivo* in syngeneic mice and showed efficacy.¹⁴³ Currently, a humanized version of this tumor-targeted sialidase is in clinical development and the toxicity profile needs to be further tested.

Targeting selectins

Selectins belong to the class of C-type lectins and bind mainly to sialoglycans.¹⁴⁴ The three selectins, E-selectin, P-selectin and L-selectin (CD62E, CD62P and CD62L) bind to different sialylated ligands, often containing sialyl-Le^x in a relatively selective manner and mediate interactions between platelets expressing P-selectin, leukocytes expressing L-selectin and endothelial cells expressing P-selectin and E-selectin.^{145–146} Expression of L-selectin on T cells can enhance cancer immunotherapy in mouse models.¹⁴⁷ The P-selectin ligand carrying protein P-selectin glycoprotein ligand (PSGL)-1 was identified to be involved in T cell exhaustion,¹⁴⁸ and was also shown to inhibit in anticancer immunity through its protein-protein interaction with the immune checkpoint VISTA, an inhibitory receptor expressed on T cells.¹⁴⁹ However, these studies have not directly implicated interactions of the carbohydrate-modification of PSGL-1 to their effect

on anticancer immunity and these effects are probably P-selectin independent. As sialoglycan-selectin interactions play an important role in leukocyte trafficking and mediation of immune responses, it is likely that sialoglycan-selectin interactions could influence anticancer immunity. Indeed, recent experiments demonstrate that modification of CAR T cells to bind to selectins can improve anticancer efficacy.^{150–151} However, further studies are needed to determine the role of selectins in anticancer immunity and how these interactions can be exploited for cancer immunotherapy.

Galectin-mediated interactions

Galectins are a class of carbohydrates-binding proteins capable of recognizing β -galactose via their carbohydrate-binding domain.^{152–154} Aberrant expression of galectins is frequent in cancer cells as well as in stromal cells and is associated with tumor progression.¹⁵³ Importantly, galectins are involved in mediating interactions between tumor cells and innate and adaptive immune cells; upregulation of galectins by tumor cells is regarded as a mechanism of tumor immune escape,^{152–154} with some galectins being ligands for immune checkpoint receptors. For example, galectin-3 binds to cytotoxic T lymphocyte antigen 4 (CTLA-4) and lymphocyte activation gene 3 (LAG-3),^{155–156} whereas galectin-9 binds to T cell immunoglobulin and mucin-domain containing-3 (TIM-3).¹⁵⁷

Galectin-1 is found upregulated in many different tumors and has been shown to antagonistically bind to the TCR, thus disrupting TCR signaling, and to determine T cell apoptosis, via redistribution of CD3 and CD45 clusters as well as CD7 and CD43 clusters.^{158–160} Early studies suggested that silencing expression of galectin-1 in tumor cells may be a strategy to enhance T-cell-mediated anti-tumor responses.¹⁶¹ Recently, a novel Gal-1-targeting DNA aptamer (AP-74 M-545) was developed and shown to suppress lung carcinoma growth in immunocompetent models. This was accompanied by an increase in CD4⁺ and CD8⁺ T cells, possibly by blocking the binding of galectin-1 to CD45.¹⁶² Nambial and colleagues showed that galectin-1 also prevents T cell migration into the tumor by upregulating PD-L1 and galectin-9 expression on endothelial cells. Blockade of galectin-1 resulted in increased T cell infiltration in multiple head and neck cancer mouse models as well as enhanced response to PD-1 blockade and radiotherapy combinations.⁵⁸ Similarly, knockdown of galectin-1 in a mouse model of pancreatic ductal adenocarcinoma increased survival and enhanced T cell infiltration.¹⁶³ In NSCLC, high expression of galectin-3 in the tumor microenvironment is associated with poor outcome¹⁶⁴ and targeting of this galectin with an antagonist has been shown to inhibit lung adenocarcinoma growth and enhance response to PD-L1 blockade *in vivo*.¹⁶⁵ Moreover, LAG-3 and interactions with galectin-3 are also under investigation as a target for cancer immunotherapy in preclinical and clinical studies.^{156–166–168}

Similarly, galectin-9/TIM-3 interactions are a recent target of immunotherapeutic intervention¹⁶⁹ due to its suppressive properties of the antitumor immune response.^{168–170}

Interactions with other lectins

C-type lectins are known to be involved in immunity, cell proliferation, tumor invasion and metastasis making them potential targets for cancer research such as selectins, DC-SIGN, Mincle, Dectin 1 and NKG2D.^{171–172} The C-type lectins are a superfamily of proteins that recognize a broad repertoire of ligands which the main feature is the C-type lectin-like domain. Cells of the adaptive and innate immune system commonly express c-type lectins including all myeloid cells, lymphocytes and dendritic cells.^{172–174} The lectin DC-SIGN can be recognized by glycosylated intercellular adhesion molecule 2 (ICAM-2) forming a DC-SIGN-ICAM-2 complex. This complex allows the maturation of dendritic cells that induces a specific cytotoxic T lymphocyte-modulated immune response promoting anti-tumor activity.¹⁷¹ Consistent with this, the DC-SIGN-Mac2 complex also inhibited the maturation of dendritic cells in colorectal carcinoma.¹⁷⁵ DC-SIGN also has other ligands such as carcinoembryonic antigen and Le.¹⁷⁶ A recent work has found that sialylation of antibodies could dampen autoimmune disease with potential consequences.¹⁷⁷ Dectin-1 expressed on dendritic cells and macrophages is critical to the NK-mediated killing of tumor cells that expressed N-glycans in high levels.¹⁷⁸ Furthermore, NK62DG is expressed on the surface of dendritic cells and its soluble ligands are very high in cancer leading to immunosuppression and poor prognosis in patients with cancer.^{179–180} In addition, the engagement of macrophage-inducible C-type lectin (Mincle) has been associated with immunosuppression and tumor progression.¹⁸¹

Impact of glycosylation on immune checkpoints

Glycosylation can mediate stability of different receptors and can influence cancer progression.¹² Immune checkpoints are also glycosylated and targeting glycosylation of PD-1 can improve anticancer immunity.¹⁸² PD-1 in T cells is N-glycosylated and its N-glycosylation is critical for the stability on the cell surface.¹⁸² In addition, a glycosylation site on PD-1 was critical for interaction with PD-L1.¹⁸² On the other side, glycosylation of PD-L1 is also critical for its stability and targeting PD-L1 glycosylation could be used to improve anticancer immunotherapy.^{183–184} Moreover, the detection of PD-L1 in human cancers is dependent on glycosylation and could influence the predictive power of PD-L1 staining and the use of PD-(L)1 blocking antibodies in patients.¹⁸⁵ Also, CTLA-4-mediated interactions are glycan-dependent and binding as well as stability could be influenced.^{186–187}

Glycosylation and signaling pathways involved in resistance to immune checkpoint inhibitors

Several intracellular signaling pathways have been associated with resistance to ICI therapy including the

activation of WNT/ β -catenin signaling, MYC signaling and loss of phosphatase and tensin homolog (PTEN).¹⁸⁸ Activation of the WNT/ β -catenin signaling pathway has been shown to increase the production of immunosuppressive cytokines,¹⁸⁹ prevention of the recruitment of BATF3⁺ dendritic cells^{190–191} and a shift towards an increase in regulatory T cells in the tumor.¹⁹² WNT/ β -catenin signaling is also regulating glycosylation by promoting the expression of the enzyme DPAGT1 and thereby enhancing increased N-glycosylation.¹⁹³ Increased N-glycosylation could further enhance immune evasion by providing ligands for immunomodulatory receptors such as Siglecs. MYC can influence the expression of PD-L1 and thereby influence the immune microenvironment in tumors.¹⁹⁴ In addition, MYC signaling can directly mediate immune cell exhaustion in NSCLC.¹⁹⁵ MYC is directly regulated by O-GlcNAcylation.¹⁹⁶ O-GlcNAcylation was needed for proliferation and MYC stability.¹⁹⁶

CONCLUSION AND OUTLOOK

Recent discoveries and progress have revealed that tumor-associated carbohydrates are an interesting new target for cancer immunotherapy interventions. Many therapeutic opportunities are ready to be explored and targeting of TACA has just started as a new field within cancer immunotherapy. Further studies are needed to determine patient populations that could benefit from such interventions. Whereas for proteins, the epidemiology and expression patterns are often well studied, the epidemiology of glycan expression will be important in order to determine the frequency of specific expression in cancer types and also during therapeutic interventions. In addition, glycan-mediated interactions with immunomodulatory lectins on leukocytes including Siglecs can directly affect anticancer immunity. Siglec-sialoglycan interactions have been described as potential new immune checkpoint. A first successful application in an early clinical trial with the Siglec-15 blocking agent (NC318) is promising. Besides the role as antigen and immunomodulatory factor, glycans can directly influence the stability and turnover of immune receptors including immune checkpoints.

We are confident that the field of glyco-immunology will enable us to improve cancer immunotherapy and help many of our patients by further studying mechanisms involved in glycan-mediated immune suppression and developing new approaches to target cancer-associated glycans.

Contributors NRM, MN, AZ and HL have written the manuscript. All authors have approved the final version for publication.

Funding This work was supported by funding from the Goldschmidt-Jacobson Foundation, the Swiss National Science Foundation (SNSF grant #3 10 030–1 84 720), the Schoenemakers-Müller Foundation and the Cancer League of Basel (Klbb).

Competing interests HL received travel grants and consultant fees from Bristol Myers Squibb (BMS) and Merck, Sharp and Dohme (MSD). HL received research support from BMS and Pallean Pharmaceuticals.

Patient consent for publication Not required.

Provenance and peer review Not commissioned; externally peer reviewed.

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