

partly due to an increase in CISH gene expression that makes the cells less responsive to cytokine stimulation.<sup>7</sup> The RNA-Seq analysis revealed that NK cell metabolism closely resembles cancer cell metabolism under hypoxic conditions, specifically an increased expression of genes related to glycolysis, amino acid synthesis and central carbon metabolism. This change in metabolism was confirmed using Seahorse assays. We also observed changes in genes related to epigenetic regulation specifically, increases in histone demethylases and decreases in DNA methyltransferases (figure 4).

**Conclusions** These results indicate that NK cells who enter the solid TME are fundamentally different than those in the bone marrow or the blood stream. Overall, the insights gained from these experiments can help overcome hypoxia induced

immune suppression in the tumor microenvironment and improve NK cell-based immunotherapy for solid tumors.

**Acknowledgements** We thank XCell biosciences for providing us with the AVATAR incubators used for these experiments

**Trial Registration** N/A

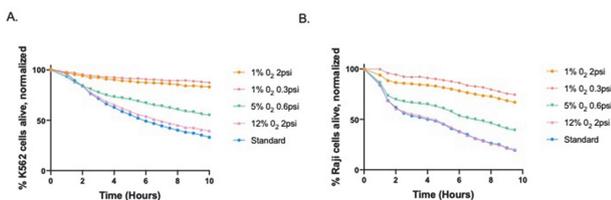
**Ethics Approval** N/A

**Consent** N/A

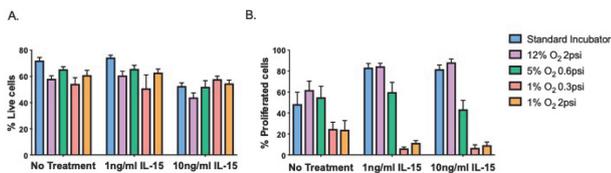
**REFERENCES**

1. Vivier E, Tomasello E, Baratin M, Walzer T, Ugolini S. Functions of natural killer cells. *Nat Immunol* 2008;**9**(5):503–10.
2. Waldhauer I, Steinle A. NK cells and cancer immunosurveillance. *Oncogene*. 2008;**27**(45):5932–43.
3. Voskoboinik I, Smyth MJ, Trapani JA. Perforin-mediated target-cell death and immune homeostasis. *Nat Rev Immunol* 2006;**6**(12):940–52.
4. Miller JS, Soignier Y, Panoskaltis-mortari A, Mcnearney SA, Yun GH, Fautsch SK, et al. Successful adoptive transfer and in vivo expansion of human haploidentical NK cells in patients with cancer. *Blood* 2005;**105**(8):3051–8.
5. Romee R, Cooley S, Berrien-Elliott MM, Westervelt P, Verneris MR, Wagner JE, et al. First-in-human phase 1 clinical study of the IL-15 superagonist complex ALT-803 to treat relapse after transplantation. *Blood* 2018;**131**(23):2515–2527.
6. Björklund AT, Carlsten M, Sohlberg E, Liu LL, Clancy T, Karimi M, et al. Complete remission with reduction of high-risk clones following haploidentical NK-Cell therapy against MDS and AML. *Clin Cancer Res* 2018;**24**(8):1834–1844.
7. Delconte RB, Kolesnik TB, Dagley LF, Rautela J, Shi W, Putz EM, et al. CIS is a potent checkpoint in NK cell-mediated tumor immunity. *Nat Immunol* 2016;**17**(7):816–24.

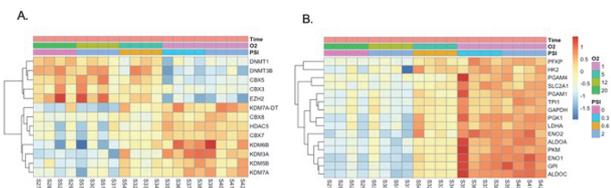
<http://dx.doi.org/10.1136/jitc-2020-SITC2020.0676>



**Abstract 675 Figure 2** Oxygen concentration effects NK cell cytotoxicity  
300,000 enriched NK cells were plated per well in 96-well round-bottom plates with 1 ng/ml IL-15. Plates were inserted in noted incubator conditions for 24 hours, 72 hours or 7 days. At the end of the incubation period NK cells were counted and plated at a 2:1 E:T ratio with fluorescently labeled K562 targets or fluorescently labeled Raji targets + Rituximab (10 ug/ml) and cells were imaged every 30 minutes. Data is shown here for K562 targets (A) and raji targets (B) at the 7 day timepoint. Representative of six separate experiments.



**Abstract 675 Figure 3** Oxygen concentration impacts NK cell proliferation  
300,000 PMBCs were CellTrace labeled and plated per well in 96-well round-bottom plates with noted treatments. The NK cells were incubated under noted incubator conditions for 7 days. At the end of 7 days, LiveDead dye was used to assess viability (A), while proliferation was assessed by evaluating CellTrace dye dilution on gated (CD56 +CD3-) NK cells (B). (N=6)



**Abstract 675 Figure 4** RNA-Seq reveals changes in gene expression  
An RNA-Seq analysis was performed on enriched NK cells incubated in noted oxygen and pressure concentrations for 24 hours, 72 hours or 7 days. A heat map of epigenetic regulation genes (A) and glycolysis genes (B) are shown for the day 7 timepoint. (N=4)

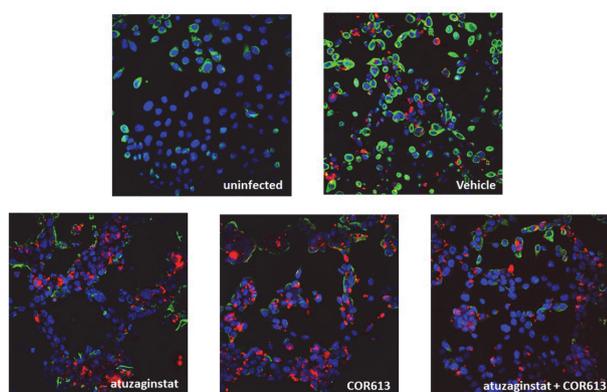
**676 PD-L1 IS INDUCED BY THE PERIODONTAL PATHOGEN PORPHYROMONAS GINGIVALIS AND CAN BE BLOCKED BY SMALL MOLECULE GINGIPAIN INHIBITORS, INCLUDING ATUZAGINSTAT**

Shirin Arastu-Kapur\*, Mai Nguyen, Sean Broce, Joseph Vacca, Kirk Ehmsen, Florian Ermini, Ursula Haditsch, Aurora Martinez-Horta, Debasish Raha, Leonardo Rodriguez, Casey Lynch, Stephen Dominy, Leslie Holsinger. *Cortexyme, Inc., South San Francisco, CA, USA*

**Background** The periodontal pathogen *Porphyromonas gingivalis* (Pg) has been linked to esophageal and other cancers through epidemiology studies. Pg’s protease virulence factors known as gingipains have been identified in esophageal cancer tissue and correlate with worse disease prognosis. Anti-PD-1 antibodies have shown some success in esophageal cancer treatment, but further understanding of the induction of PD-L1 in esophageal cells is needed to identify potential treatment modalities. Pg has been shown to induce PD-L1 on the surface of infected cells, suggesting that the presence of Pg in esophageal cancer cells may contribute to PD-L1 expression and immune escape. One of the pathways known to induce PD-L1 is wnt pathway activation resulting in b-catenin translocation to the nucleus. Prior studies have demonstrated that Pg activates the wnt pathway by a non-canonical mechanism, leading to b-catenin nuclear localization.

**Methods** An immortalized non-transformed esophageal cell line, Het-1A, was used to investigate the level of PD-L1 induction by Pg infection using quantitative immunofluorescence. PD-L1 expression was measured using irreversible gingipain inhibitors against lysine-gingipain (Kgp) and arginine-gingipain (Rgp). Pg-induced PD-L1 expression pathways were investigated by Western blot and qPCR. PD-L1 induction by Pg was characterized in cancer cell lines that have an endogenous level of PD-L1 expression, including tongue squamous cell carcinoma (SCC25) and neuroblastoma (SH-SY5Y). PD-L1 induction by Pg was assessed in a murine derived RAW macrophage cell line that is critical for anti-PD-1 responses.

**Results** Pg infection increased PD-L1 expression on Het-1A cells within 24 hours of infection and increased PD-L1 mRNA within 4 hours of infection. PD-L1 expression level correlated with cellular bacterial burden on the cells in a dose-dependent manner. PD-L1 expression was decreased by the Kgp inhibitor, atuzaginstat, or an Rgp inhibitor, COR613, and PD-L1 expression was completely blocked when both gingipain inhibitors were used together (figure 1). Pg also induced expression of PD-L1 on the surface of infected SCC-25, SH-SY5Y, and RAW cell lines. Western blot analysis and qPCR revealed that Kgp inhibition, but not Rgp inhibition, was able to inhibit the non-canonical activation of b-catenin and down regulation of classical wnt pathway effectors at both the mRNA and protein level.



**Abstract 676 Figure 1** Gingipain inhibitors block PD-L1 induced by Pg

Pg grown with and labeled by red fluorescent membrane-incorporated dye was pre-treated with vehicle or the compounds listed for 30 min. Het-1A cells were infected (MOI = 20) for 24 hours, washed, fixed and stained for visualization of the nuclei (DAPI, blue), PD-L1 protein (anti-PDL1 primary and secondary antibodies, green), and Pg infection (red). Images were captured with immunofluorescent confocal microscopy.

**Conclusions** In host cells infected with Pg, gingipains mediate the induction of PD-L1 as a mechanism of immune evasion through the non-canonical activation of the wnt pathway. Further studies to elucidate induction mechanisms are in progress. In esophageal cancer and other cancers infected with Pg, combining gingipain inhibitors with anti-PD-1 therapy may improve treatment outcomes.

<http://dx.doi.org/10.1136/jitc-2020-SITC2020.0676>

677

#### EVALUATION OF ANTI-PD1 EFFICACY IN GERM-FREE AND ANTIBIOTIC-TREATED SPF MICE BEARING MC38 TUMORS

<sup>1</sup>Ying Jin, <sup>2</sup>Xue Liang, <sup>3</sup>Frieda Zhang, <sup>4</sup>Guangmao Mu, <sup>4</sup>Wei Zhou, <sup>4</sup>Annie An, <sup>5</sup>Marvin Ouyang, <sup>2</sup>Cai Li, <sup>2</sup>Andy Liaw, <sup>4</sup>Henry Li, <sup>4</sup>Davy Ouyang\*. <sup>1</sup>Crown Bioscience Inc, San Diego, CA, USA; <sup>2</sup>Merck Research Laboratories, Kenilworth, NJ, USA; <sup>3</sup>Cyagen Biosciences Inc, Taicang, China; <sup>4</sup>Crown Bioscience Inc., San Diego, CA, USA; <sup>5</sup>Cyagen Biosciences Inc., Taicang, China

**Background** Increasing evidence has indicated the important role of gut microbes in mediating normal and pathologic immune responses to cancer in both patients and animal models. There is growing effort in modulating microbiota composition to improve the outcome of cancer immunotherapy. To

investigate the immunomodulatory roles of microbiota-based therapeutics preclinically, germ-free (GF) mice are often used because they are free of microorganisms. However, logistic challenges and inherited physiological deficits in GF mice are also generally acknowledged. Alternative approach of depleting gut microbiota in using specific pathogen-free (SPF) mice with broad-spectrum antibiotics has also been adopted. Potential challenges with this approach are possible acquisition of antibiotic-resistant bacteria and potential expansion of fungi. Here we report on the efficacy assessment of anti-PD-1 mAb on MC38 syngeneic tumors in both GF mice and antibiotic-treated SPF mice.

**Methods** C57BL/6 mice were inoculated subcutaneously with MC38 tumor cells. In the GF study, GF mice (Taconic, provided by Cyagen) were housed in germ-free isolators at a Cyagen facility, and a cohort of SPF mice (Taconic) were used as controls. Both GF and SPF mice were randomized for isotype or anti-PD-1 mAb (mDX400) treatment when the tumors were established (80–120 mm<sup>3</sup>) and were continuously monitored for tumor growth over time. In the antibiotic treatment study, different antibiotic regimens were administered to SPF mice (Lingchang) in drinking water starting 2 weeks prior to MC38 tumor inoculation and continued throughout the study. Mice were treated with vehicle control or anti-PD-1 mAb (RMP1-14; CrownVivo™).

**Results** Tumor growth is significantly faster in GF than SPF mice, and mDX400 slowed the tumor growth rate in both GF and SPF mice. The tumors achieved complete regressions on 4 out of 10 GF mice as compared to 6 out of 10 SPF mice, yet the difference of mDX400 efficacy in GF vs SPF mice did not reach statistical significance. In antibiotic-treated SPF mice, none of the antibiotic regimens showed significant impact on MC38 tumor growth nor anti-PD-1 efficacy in SPF mice, which was contrary to most reported data. Immune profiling on tumor infiltrating lymphocytes in these mice and microbiota analysis by 16S rRNA gene amplicon sequencing are ongoing and the data will be presented at the meeting.

**Conclusions** We have observed faster tumor growth in GF mice, however, the efficacy of anti-PD-1 antibody is not impacted by GF condition or treatment with broad-spectrum antibiotics. These results are different from previously published work, indicating high variability in preclinical models. Ongoing analysis with antibiotic-treated mice will provide further insight.

**Ethics Approval** Animal experiments were conducted in accordance with animal welfare law, approved by local authorities, and in accordance with the ethical guidelines of CrownBio (Taicang).

<http://dx.doi.org/10.1136/jitc-2020-SITC2020.0677>

678

#### ADDITION OF A SINGLE BACTERIA FACILITATES ANTI-TUMOR IMMUNITY AND LONG-TERM SURVIVAL IN COLORECTAL CANCER

<sup>1</sup>Abigail Overacre-Delgoffe\*, <sup>1</sup>Anthony Cillo, <sup>1</sup>Hannah Bumgarner, <sup>1</sup>Ansen Burr, <sup>2</sup>Justin Tometich, <sup>2</sup>Amrita Bhattacharjee, <sup>3</sup>Tullia Bruno, <sup>3</sup>Dario Vignali, <sup>1</sup>Timothy Hand. <sup>1</sup>University of Pittsburgh, Pittsburgh, USA; <sup>2</sup>Children's Hospital of Pittsburgh, Pittsburgh, PA, USA; <sup>3</sup>UPMC Cancer Center, Pittsburgh, PA, USA

**Background** Colorectal cancer remains one of the most common and deadliest cancers worldwide and effective therapies are lacking. While immunotherapy has revolutionized treatment for many cancers, the overwhelming majority of