Copy number loss in granzyme genes confers resistance to immune checkpoint inhibitor in nasopharyngeal carcinoma

Yuxiang Ma,¹ Xi Chen,² Ao Wang,³ Hongyun Zhao,¹ Qingguang Lin,⁴ Hua Bao,³ Yang Zhang,¹ Shaocong Hong ,² Wanxiangfu Tang ,³ Yan Huang,² Yunpeng Yang,² Xue Wu,³ Yang Shao,^{3,5} Wenfeng Fang,² Li Zhang²

To cite: Ma Y, Chen X, Wang A, *et al.* Copy number loss in granzyme genes confers resistance to immune checkpoint inhibitor in nasopharyngeal carcinoma. *Journal for ImmunoTherapy of Cancer* 2021;**9**:e002014. doi:10.1136/jitc-2020-002014

► Additional material is published online only. To view please visit the journal online (<http://dx.doi.org/10.1136/jitc-2020-002014>).

YM, XC, AW and HZ contributed equally.

Accepted 14 February 2021

© Author(s) (or their employer(s)) 2021. Re-use permitted under CC BY-NC. No commercial re-use. See rights and permissions. Published by BMJ.

For numbered affiliations see end of article.

Correspondence to

Dr Li Zhang, Department of Medical Oncology, Sun Yat-sen University Cancer Center, Guangzhou, China; zhangli@sysucc.org.cn

Dr Wenfeng Fang, Department of Medical Oncology, Sun Yat-sen University Cancer Center, Guangzhou, China; fangwf@sysucc.org.cn

Dr Yang Shao, Medical Department, Nanjing Geneseeq Technology Inc., Nanjing, China; yang.shao@geneseeq.com

ABSTRACT

Background Anti-programmed death (PD)-1 therapy has recently been used in recurrent or metastatic (R/M) nasopharyngeal carcinoma (NPC). The long-term survival and its biomarkers responding to anti-PD-1 treatment in patients with R/M NPC remain unclear.

Methods Patients with R/M NPC were enrolled between March 2016 and January 2018 from two phase I clinical trials. The median follow-up period was 24.7 months. Eligible patients progressed on standard chemotherapy had measurable disease by Response Evaluation Criteria in Solid Tumor V.1.1. Non-obligatory contemporaneous tumor samples were collected for whole-exome sequencing. The primary outcome was objective response rate (ORR). Duration of response (DOR), progression-free survival (PFS), and overall survival (OS) were secondary outcomes assessed in all patients.

Results Among 124 evaluable patients, anti-PD-1 therapy achieved an ORR of 29.8% and a durable clinical benefit rate of 60.5%. The median OS (mOS) was 17.1 months (95% CI 14.2 to 24.7), median PFS (mPFS) was 3.8 months (95% CI 3.4 to 6.0), and median DOR was 9.5 months. Significant OS benefit from treatment was observed in patients without liver metastasis (23.8 vs 13.3 months, $p=0.006$). Copy number deletion in genes encoding granzyme B or granzyme H (*GZMB/H*) was associated with poor treatment outcome (mPFS altered vs wildtype: 1.7 vs 3.6 months, $p=0.03$; mOS altered vs wildtype: 10.1 vs 18 months, $p=0.012$).

Conclusions Anti-PD-1 treatment provided promising clinical benefit in pretreated patients with R/M NPC. Copy number loss in either *GZMB* or *GZMH* genes was associated with reduced survival.

BACKGROUND

Nasopharyngeal carcinoma (NPC) is a head and neck cancer arising from the epithelial lining of the nasopharynx,^{1 2} with a high prevalence in Southeast Asian populations.³ Unlike other head and neck cancer, NPC is characterized by extensive Epstein-Barr virus (EBV) infection,⁴ which is implicated in driving tumorigenesis via its evasion of host

innate immunity.⁵ Currently, gemcitabine plus cisplatin is used as standard first-line treatment for recurrent or metastatic (R/M) NPC.^{6 7} Patients who progressed on first-line treatment have poor prognosis and few available follow-up treatment options. Active agents used in second line include paclitaxel, docetaxel, gemcitabine, capecitabine and so on, either as single agent or in combination with platinum. Nonetheless, second-line treatments often yield unsatisfactory results, with estimated median progression-free survival (mPFS) at 5.2 months and estimated median overall survival (mOS) at 12.5 months.⁸

The last decade has seen significant development in immune-checkpoint inhibitors (ICIs), which have shown promising antitumor activity in a range of tumors including NPC.^{9–11} Additionally, previous studies reported the short-term efficacy and safety of ICIs in NPC.^{12–14} Among these, we have published preliminary results from two phase I clinical trials composing the largest cohort of R/M NPC treated with anti-PD-1 to date^{12 14} that demonstrated anti-PD-1 therapy have comparable immediate benefit to chemotherapies. Tumor mutation burden (TMB) was promising biomarkers of immune-oncology (IO) therapy in other cancer types.^{15 16} However, others reported that the prognostic value of TMB was minimal since patients with NPC generally had lower mutation burden compared with non-small cell lung cancer (NSCLC) or melanoma. Therefore, NPC-specific molecular markers that can predict patients' response to IO therapy are urgently needed.

Granzyme B (*GZMB*) is a serine protease that plays an important role in T cell and NK cell mediated tumor killing.¹⁷ It has been reported that granzyme H (*GZMH*)

has a similar capability.¹⁸ Recently, *GZMB* has emerged as a predictor of IO therapy response in different cancer types. One study found that the expression of *GZMB* in tumor samples from patients with melanoma was significantly correlated with response, and the level of *GZMB* expression increased during treatment in responders.¹⁹ Another study in patients with stage IV NSCLC found that high serum *GZMB* level was associated with better survival and patients with germline variants in *GZMB* had reduced survival compared with wildtype.²⁰ Given the significance of granzyme function in lymphocyte cytotoxicity and implication in IO therapy outcome, we hypothesize that the granzyme family is predictive of response to anti-PD-1 therapy in NPCs and loss of function in this pathway is associated with reduced survival.

METHODS

Study design and patients

Patients with R/M NPC were consecutively enrolled in two phase I clinical trials for advanced solid tumors (camrelizumab and nivolumab, ClinicalTrials.gov identifiers: NCT02721589 and NCT02593786) between March 2016 and January 2018.^{12,14} The details of studies design of dose escalation and expansion phases were already reported. Camrelizumab trial enrolled 33 patients with NPC in dose escalation phase, and 60 patients with NPC in dose expansion phase (200 mg fixed dose every 2 weeks). Nivolumab trial enrolled 33 patients with NPC in dose escalation phase. Sample sizes in dose escalation phases were based on safety (dose-limiting toxicity), and in dose expansion phase were decided for overall response rate (ORR) consideration compared with historical control. The distribution of patients' treatment in the current study was shown in online supplemental figure S1. Baseline tumor samples and matched peripheral blood were provided non-obligatory right before the initiation of anti-PD-1 treatment. Written informed consent were provided by all patients.

Treatment and evaluation

Eligible patients received intravenous infusion of camrelizumab at dosages range from 1 mg/kg, 3 mg/kg, 200 mg and 10 mg/kg every 2 weeks; or nivolumab at dosage of 3 mg/kg, 240 mg every 2 weeks and 360 mg every 3 weeks. Radiographic tumor assessments were taken at baseline and approximately every 6 weeks. Treatment response was assessed by investigators per Response Evaluation Criteria in Solid Tumors (RECIST) version 1.1. ORR was the sum of complete response (CR) and partial response (PR). Durable clinical benefit (DCB) was defined as the percentage of patients who achieved CR or PR or stable disease (SD) lasted >6 months; non-durable clinical benefit (NDB) was defined as PD or SD that lasted ≤6 months. Duration of response (DOR) was define as the duration from objective response to progression disease (PD) in responders. Treatment continued until confirmed PD, intolerable toxicities, death or withdrawal

of consent. PFS was defined as the time from first dose to PD, or prior death. Censored data documented last radiographic assessment before cut-off, loss of follow-up or change of treatment. Treatment beyond initial RECIST disease progression was permitted as long as patients satisfied the criteria in protocols. Survival follow-up was approximately every 3 months by clinic visits or telephone calls. Overall survival (OS) was duration from first dose to death, patients who remained alive were censored at the date of their last follow-up.

WES, mutation calling and copy number analysis

Available tumor tissues from 60 patients in this cohort underwent whole-exome sequencing (WES). Genomic DNAs from Formalin-fixed, paraffin-embedded (FFPE) or biopsy tumor samples and blood samples were extracted with QIAamp DNA FFPE Tissue Kit and DNeasy Blood and tissue kit (Qiagen, USA), respectively, and quantified by Qubit V.3.0 using the dsDNA HS Assay Kit (ThermoFisher Scientific, USA). WES was performed on Illumina HiSeq4000 platform using PE150 sequencing chemistry (Illumina, USA). Trimmomatic was used for FASTQ file quality control.²¹ Leading/trailing low quality (quality reading below 20) or N bases were removed. Paired-end reads were then aligned to the reference human genome (build hg19), using the Burrows-Wheeler Aligner.²² PCR deduplication was performed using Picard.²³ GATK3 was used for local realignment around indels and base quality score recalibration.²⁴ Cross-sample contamination was estimated using ContEst²⁵ (Broad Institute, contamination rate <0.02). Somatic Single Nucleotide Variant calling was generated by Mutect²⁶ and insertion/deletions were called with Scalpel.²⁷ Copy number alteration (CNA) analysis from sequencing data was performed using CNVKit.²⁸ Focal level gain and loss were defined as normalized log2 depth ratio ≥1 or ≤−0.6,²⁹ respectively.

Statistical analysis

Comparisons of proportion between groups and association between categorical variables were examined using the χ^2 test. In biomarker subgroup analysis, quantitative variables such as TMB were stratified at multiple thresholds to identify association with survival. For survival analyses, Kaplan-Meier curves were compared using the log-rank test, and HRs were calculated by Cox proportional hazards model. Multivariate analysis was performed to assess potential confounding effects among clinical variables. The focal CNA analysis was performed using a previously described pipeline.⁹ CNA p value was corrected for multiple hypothesis testing using the Benjamini-Hochberg method. All statistical analyses were performed in R (V.3.3.2).

RESULTS

Survival outcomes of the entire cohort

In the current study, 124 patients with R/M NPC were retrospectively examined. OS outcome and treatment

response to anti-PD-1 antibody were analyzed. Tumors from 60 patients were profiled by WES to uncover mutational profiles related to treatment outcome. The median follow-up duration was 24.7 months (95% CI 23.3 to 26.6). The majority of the patients (66.1%) failed at least two prior lines of therapy. All baseline characteristics were shown in [table 1](#). Additionally, the baseline demographic from the 60 patients with WES data were not showing any significant differences compared with the other patients (online supplemental table S1).

Treatment by anti-PD-1 in all enrolled patients achieved a median OS of 17.1 months (95% CI 14.2 to 24.7) and a median PFS of 3.8 months (95% CI 3.4 to 6.0). One-year OS rate was 62.6% (95% CI 54.5% to 71.9%), and the 2-year OS rate was 39.7% (95% CI 31.5% to 50%; [figure 1A](#)). The ORR for all patients was 29.8% (2 CR and 35 PR), and the DCB rate was 60.5% (online supplemental table S2). The median DOR was 9.5 months (95% CI 6.0 to 13.0).

The patient's best response strongly predicted OS (mOS CR: not reached (NR); PR: NR; SD: 19.8 months; PD: 5.8 months; [figure 1B](#)). In univariate analysis ([figure 2](#)), prior radiation was associated with better survival, whereas the number of distant metastatic sites and liver metastasis were associated with reduced survival (online supplemental figure S2). Other baseline clinical variables were not associated with survival in univariate and multivariate models ([figure 2](#), online supplemental table S3).

Molecular markers associated with response

Among the 60 patients with WES data, the median TMB was 52. The distribution of TMB was not significantly different between patients with PR, SD, or PD (online supplemental figure S3A). Patients were grouped binarily as TMB high versus low based on four different cut-off values (20%, 40%, 60%, and 80% of TMB ranking), neither PFS nor OS differed between groups (online supplemental figure S3B,C). We divided patients into TMB high and low group by median and showed neither PFS nor OS were different between groups (online supplemental figure S3D,E), which was probably due to the overall low TMB levels within this cohort (data not shown). Additionally, the proportions of TMB high group among patients who received camrelizumab and nivolumab are not statistically different (28.6% vs 27.8%, Fisher's exact test, $p=1$, detail data not shown). Likewise, the tumor neoantigen burden showed no impact on neither PFS nor OS (online supplemental figure S4A,B).

Next, we sought to characterize key immune-related pathways in the tumor samples, including 17 previously reported immune-related pathways from the Immunology Database and Analysis Portal (ImmPort, <https://www.immport.org/>), and the gasdermin, granzyme pathways. Because mutations were rare in these immune genes in this cohort, the distribution of CNA in key immune-related pathways was examined. Within these 19 immune-related pathways, only interferon (IFN), gasdermin and granzyme gene deletions occurred in more than 4 patients

Table 1 Patients' demographics and baseline clinical characteristics

No. (%)	All patients (124)
Median age, years (range)	46 (23–73)
Sex (male/female)	
Male	95 (76.6)
Female	29 (23.4)
ECOG performance status score	
0	35 (28.2)
1	89 (71.8)
Smoking	
Never	85 (68.6)
Former/current	35 (28.2)
Unknown	4 (3.2%)
Stage	
Primary metastasis	13 (10.5)
Recurrent with distant metastasis	111 (89.5)
WHO histological classification	
Undifferentiated non-keratinised	96 (77.4)
Differentiated non-keratinised	14 (11.3)
Keratinised squamous carcinoma	14 (11.3)
Distant metastasis sites	
Lung	69 (55.6)
Liver	69 (55.6)
Bone	50 (40.3)
Distant lymph	46 (37.1)
Others	17 (13.7)
None	8 (6.5)
Prior lines for advanced disease	
1	42 (33.9)
2	35 (28.2)
3	25 (20.2)
4 or more	22 (17.7)
Prior chemotherapy for advanced disease	
Paclitaxel	74 (59.7)
Gemcitabine	69 (55.6)
5-Fluorouracil	52 (41.9)
Docetaxel	37 (29.8)
Tegafur	25 (20.2)
Cetuximab	23 (18.5)
Capecitabine	19 (15.3)
Ipilimumab	8 (6.5)
Comorbidity	
Yes	83 (66.9)
No	41 (33.1)
Prior radiation	
Yes	111 (89.5)
No	13 (10.5)
WES detection	
Yes	60 (48.4)
No	64 (51.6)

ECOG, Eastern Cooperative Oncology Group; WES, whole-exome sequencing; WHO, world health organization.

Figure 1 Overall survival (OS) outcome and association with clinical variables. (A) Kaplan-Meier curve of OS in all patients with 95% CI band represented by the shaded area. CI, confidence interval. (B) OS in all treated patients by best overall response. CR, complete remission; PR, partial remission; SD, stable disease; PD, progression disease; NR, not reached. (C) Swimmer plot that shows time to first response and duration of response. The length of the bars represents OS month. The first timepoint of response or progression, treatment discontinuation, and time of death were labeled for patients with events. Patients that continue treatment are labeled with an arrow.

(online supplemental table S4), and these 3 pathways were therefore selected for downstream analysis.

Frequent deletion in granzyme genes was observed in this cohort, among which copy number loss in *GZMB* and *GZMH* occurred at the highest frequency (figure 3, online supplemental table S5). Deletion was also observed in gasdermins and type I IFN but at lower frequencies compared with granzymes. Mutations and deletions on other genes were also analyzed as shown in online supplemental tables S6 and S7). We examined the effect of *GZMB* and *GZMH* deletion on survival and found patients with copy number loss in either gene had worse PFS (1.7 vs 3.6 months, $p=0.03$) and OS (10.1 vs 18 months, $p=0.012$) compared with the wildtype, as shown in figure 4. Fisher's exact test confirmed that the proportions of *GZMB/H* loss were not statistically different among the camrelizumab and nivolumab group (47.6% vs 61.1%, $p=0.405$,

detail data not shown). Furthermore, multivariate analysis using treatment agents and *GZMB/H* status confirms that *GZMB/H* loss have significant attribution towards patient PFS ($p=0.027$) and OS ($p=0.013$), while different treatment reagents had no significant attribution, as shown in online supplemental figure S5A,B. Similarly, these patients also had lower PR ratio ($p=0.039$) and DCB ratio ($p=0.066$) compared with the wildtype, as shown in figure 4A,B.

In addition, we looked at the overall effect of deletion in granzymes, gasdermins, and IFNs on survival. As shown in online supplemental figure S6A–C), the median PFS among patients with alternation in these pathways was worse compared with the wildtypes, although it was not significant for gasdermin alterations (mPFS granzyme 1.7 vs 3.6 months, $p=0.012$; gasdermin 1.8 vs 3.5 months, $p=0.39$; IFN 1.7 vs 3.5 months, $p=0.023$). Similarly,

Figure 2 Associations of baseline demographic and clinical characteristics with (A) overall survival (OS) and (B) progression-free survival (PFS) in univariate analysis. All variables are categorical. HR greater than 1 indicates association with reduced survival. P values are labeled on the right side of each individual plot. ECOG, Eastern Cooperative Oncology Group; NR, not reached.

patients with alteration in their tumor had inferior OS compared with these without alteration (mOS granzyme 7.4 vs 18 months, $p=0.0046$; gasdermin 4.3 vs 15.9 months, $p=0.042$; IFN 8 vs 18 months, $p=0.00079$; online supplemental figure S7A–C). Furthermore, patients who had alteration in any of the pathways had significantly reduced PFS and OS compared with these with normal pathway functions (mPFS 1.7 vs 4.4 months, $p=0.0047$; mOS 7.4 vs 21 months, $p<0.0001$; online supplemental figure S7D).

Interestingly, in a cohort of patients with NPC treated with radiotherapy instead of checkpoint blockade and had tumor sequenced by WES, *GZMB* and *GZMH* deletion was not associated with OS nor PFS (online supplemental figure S8A,B).³⁰ Furthermore, to test if *GZMB* and *GZMH* deletion were associated with response to checkpoint inhibitor in non-microbial-associated cancers, we examined a pan-cancer cohort including NSCLC, melanoma, bladder, and several other cancer types.¹⁰ Patients with deletion in either gene had reduced PFS and OS compared with wildtype although the difference was not significant (mPFS 2.6 vs 4.6 months, $p=0.084$; mOS 10.0 vs 21.7 months, $p=0.11$; online supplemental figure S8C,D).

DISCUSSION

This study reported the first and largest cohort of long-term survival outcome under anti-PD-1 therapy in R/M NPC. Overall, our results demonstrated that anti-PD-1 treatment achieved durable survival benefit in patients with R/M NPC, and might yielded notable improved OS

compared with historical control of second-line chemotherapies (median 12.5 months).^{8 31 32} Furthermore, anti-PD-1 achieved durable survival benefit in responders and SD in a subset of patients. For those 2-year survivors, mDOR was as long as 19.4 months. Furthermore, several clinical biomarkers showed strong correlation with anti-PD-1 treatment outcome in R/M NPC. Patients with liver metastasis at baseline achieved reduced PFS and OS, which is also consistent with other findings in melanoma and NSCLC treated with anti-PD-1.^{33 34} These findings suggest the long-term survival benefit of anti-PD-1 therapy for R/M NPC and could be realized through individualized treatment selection by biomarkers.

Several important immune-related pathways were frequently altered in this cohort and associated with resistance to anti-PD-1 therapy. Specifically, we found that alteration in granzyme, gasdermin, and type I IFN were enriched in refractory tumors. Both *GZMB* and *GZMH* are known to induce rapid apoptosis of target cells by cytotoxic T cell or nature killer cells.¹⁸ In addition, *GZMH* also plays an important role in controlling viral infection. One study has shown that *GZMH* is important in clearing of hepatitis B virus by cytotoxic lymphocytes by direct targeting of viral proteins.³⁵ *GZMB* and *GZMH* deletion were recurrent in this cohort and strongly correlated with poor outcome. Moreover, in patients with NPC treated with non-IO regimen, *GZMB*/*GZMH* deletion was not associated with OS. Additionally, patients with *GZMB* deletion also had inferior survival in IO cohort of other cancer types as reported by several studies.^{19 20} *GZMB* and

Figure 3 Distribution of copy number loss of pyroptosis pathways in 60 patients with nasopharyngeal carcinoma (NPC). Progression-free survival (PFS) and clinical characteristics of each patient were shown at the top. Loss of function (copy number loss and frame shift deletion) in interferon (IFN), gasdermins and granzyme genes were shown at the bottom. PR, partial remission; PD, progression disease; SD, stable disease.

GZMH are, in theory, predominantly expressed in the immune cells. Recent studies of single-cell transcriptomic analysis in patients with NPC have shown that *GZMB* and *GZMH* genes are highly expressed in the immune cells, but not expressed in the NPC cancer cells.^{36 37} It is likely that such signature loss in the immune cells are detected by our sequencing process, due to tumor-infiltrating immunes. Similarly, Gao *et al* has reported that the loss of IFN genes in the tumor cells, despite mostly expressed in immune cells, can be used as prognostic markers in patients receiving immune checkpoint therapy.³⁸ Moreover, we noticed a difference of *GZMB/H* predictive power between NPC and other cancer types, which can be explained by the various levels of tumor-infiltrating lymphocytes among these tumor types. However, more experiments are needed to validate this hypothesis. Taken together, *GZMB* and *GZMH* are critical in augmenting the anti-PD-1 antibody mediated immune function and tumor killing.

Traditionally, apoptosis was considered the dominant form of programmed cell death induced by cytotoxic lymphocytes. However, recently studies have shown that non-apoptotic killing including pyroptosis is also used by these immune cells to clear infection or cancer cells.^{39 40} Pyroptosis, a proinflammatory form of cell death,⁴¹ is induced when gasdermin B are cleaved by

granzyme A and this effect is upregulated by IFN.³⁹ Similarly, *GZMB* can also activate pyroptosis in target cells by gasdermine E.⁴² We found alteration in granzyme, gasdermin, and IFN genes resulted in reduced IO therapy response likely due to dysfunctional pyroptosis pathway and therefore weakened cytotoxic lymphocyte function. Evidence had shown that, during EBV latency and lytic cycles, type I and II IFN Jak-STAT signaling pathways were manipulated; expression and activity of IFN regulating factors were altered and host apoptosis signaling pathways were repressed.⁵ Therefore, compared with other tumors, the immune system of NPC may largely depend on non-apoptotic killing such as pyroptosis to eliminate viral transformed cells. Our findings suggest that even when the immune-checkpoint brakes are released by anti-PD-1 antibodies, intact downstream effector functions of cytotoxic lymphocytes are necessary for cancer clearance. Future studies are needed to validate the prognostic value of these markers in more cancer types and to further examine their association with pyroptosis.

The present study was limited by its retrospective nature. Whether clinical and genomic variables we found were direct causes for treatment outcome is not clear and should be studied in prospective trials with larger sample size. Since only a subset of patients with both available tumor and blood samples was included in the WES

Figure 4 The effect of granzyme B or H (*GZMB* or *GZMH*) deletion on patients' responses and survival. (A) Bar graph of Response Evaluation Criteria in Solid Tumors (RECIST) response in patients with alternation in either *GZMB* or *GZMH* gene versus wildtype. Fisher's exact test. (B) Bar graph of durable clinical response in patients with alternation in either *GZMB* or *GZMH* gene versus wildtype. Fisher's exact test. (C) Kaplan-Meier curve of progression-free survival (PFS) in patients with altered *GZMB* or *GZMH* genes versus wild type. (D) Kaplan-Meier curve of overall survival (OS) in patients with altered *GZMB* or *GZMH* genes versus wild type. DCB, durable clinical benefit; NDB, non-durable clinical benefit; PD, progression disease; PR, partial remission; SD, stable disease.

profiling, results derived from these analyses should be interpreted with caution. Furthermore, the WES profiling failed to distinguish between the immune cells and cancer cells. Single-cell level transcriptomic analysis are needed to better understand the potential of *GZMB/H* as biomarkers. Nonetheless, our results offered new insights

on potential biomarkers of anti-PD-1 therapy in patients with R/M NPC.

In summary, we reported the long-term survival of patients with R/M NPC treated with anti-PD-1 and identified baseline clinical factors that were associated with survival outcome. Our results suggested alteration

in pyroptosis pathways including IFN, gasdermin, and granzyme coding genes were associated with poor survival. These findings should be validated in prospective trials.

Author affiliations

¹Department of Clinical Research, Sun Yat-sen University Cancer Center, State Key Laboratory of Oncology in South China, Collaborative Innovation Center for Cancer Medicine, Guangzhou, China

²Department of Medical Oncology, Sun Yat-sen University Cancer Center, State Key Laboratory of Oncology in South China, Collaborative Innovation Center for Cancer Medicine, Guangzhou, China

³Medical Department, Nanjing Geneseeq Technology Inc, Nanjing, Jiangsu, China

⁴Department of Ultrasonography, Sun Yat-sen University Cancer Center, State Key Laboratory of Oncology in South China, Collaborative Innovation Center for Cancer Medicine, Guangzhou, China

⁵School of Public Health, Nanjing Medical University, Nanjing, Jiangsu, China

Contributors YM, XC, AW and HZ contributed equally to the work.

Funding This work was supported by Chinese National Natural Science Foundation project (No. 81572659 and 81972556 to WF) and Physician scientist training program of Sun Yat-Sen University Cancer Center (No. 14zxqk03 to WF).

Competing interests LZ declares research grant from Lilly. AW, HB, WT, XW and YS are the employees of Nanjing Geneseeq Technology.

Patient consent for publication Not required.

Ethics approval All study protocols were approved by the ethics committee of Sun Yat-sen University Cancer Center and in accordance with international standards of good clinical practice.

Provenance and peer review Not commissioned; externally peer reviewed.

Data availability statement The data of this study are available upon reasonable request from the corresponding author, LZ.

Supplemental material This content has been supplied by the author(s). It has not been vetted by BMJ Publishing Group Limited (BMJ) and may not have been peer-reviewed. Any opinions or recommendations discussed are solely those of the author(s) and are not endorsed by BMJ. BMJ disclaims all liability and responsibility arising from any reliance placed on the content. Where the content includes any translated material, BMJ does not warrant the accuracy and reliability of the translations (including but not limited to local regulations, clinical guidelines, terminology, drug names and drug dosages), and is not responsible for any error and/or omissions arising from translation and adaptation or otherwise.

Open access This is an open access article distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited, appropriate credit is given, any changes made indicated, and the use is non-commercial. See <http://creativecommons.org/licenses/by-nc/4.0/>.

ORCID iDs

Shaodong Hong <http://orcid.org/0000-0002-5632-6857>

Wanxiangfu Tang <http://orcid.org/0000-0002-9045-2478>

REFERENCES

- Carioli G, Negri E, Kawakita D, *et al.* Global trends in nasopharyngeal cancer mortality since 1970 and predictions for 2020: focus on low-risk areas. *Int J Cancer* 2017;140:2256–64.
- Wei WI, Sham JST. Nasopharyngeal carcinoma. *Lancet* 2005;365:2041–54.
- Yu MC, Yuan J-M. Epidemiology of nasopharyngeal carcinoma. *Semin Cancer Biol* 2002;12:421–9.
- Shanmugaratnam K, Sobin LH. The world Health organization histological classification of tumours of the upper respiratory tract and ear. A commentary on the second edition. *Cancer* 1993;71:2689–97.
- Ning S. Innate immune modulation in EBV infection. *Herpesviridae* 2011;2:1.
- Zhang L, Huang Y, Hong S, *et al.* Gemcitabine plus cisplatin versus fluorouracil plus cisplatin in recurrent or metastatic nasopharyngeal carcinoma: a multicentre, randomised, open-label, phase 3 trial. *The Lancet* 2016;388:1883–92.
- Jin Y, Shi Y-X, Cai X-Y, *et al.* Comparison of five cisplatin-based regimens frequently used as the first-line protocols in metastatic nasopharyngeal carcinoma. *J Cancer Res Clin Oncol* 2012;138:1717–25.
- Chua DTT, Kwong DLW, Sham JST, *et al.* A phase II study of ifosfamide, 5-fluorouracil and leucovorin in patients with recurrent nasopharyngeal carcinoma previously treated with platinum chemotherapy. *Eur J Cancer* 2000;36:736–41.
- Fang W, Ma Y, Yin JC, *et al.* Comprehensive Genomic Profiling Identifies Novel Genetic Predictors of Response to Anti-PD-(L)1 Therapies in Non-Small Cell Lung Cancer. *Clin Cancer Res* 2019;25:5015–26.
- Miao D, Margolis CA, Vokes NI, *et al.* Genomic correlates of response to immune checkpoint blockade in microsatellite-stable solid tumors. *Nat Genet* 2018;50:1271–81.
- Ma BBY, Lim W-T, Goh B-C, *et al.* Antitumor activity of nivolumab in recurrent and metastatic nasopharyngeal carcinoma: an international, multicenter study of the Mayo clinic phase 2 Consortium (NCI-9742). *J Clin Oncol* 2018;36:1412–8.
- Fang W, Yang Y, Ma Y, *et al.* Camrelizumab (SHR-1210) alone or in combination with gemcitabine plus cisplatin for nasopharyngeal carcinoma: results from two single-arm, phase 1 trials. *Lancet Oncol* 2018;19:1338–50.
- Hsu C, Lee S-H, Ejadi S, *et al.* Safety and antitumor activity of pembrolizumab in patients with programmed Death-Ligand 1-Positive nasopharyngeal carcinoma: results of the KEYNOTE-028 study. *JCO* 2017;35:4050–6.
- Ma Y, Fang W, Zhang Y, *et al.* A phase I/II open-label study of nivolumab in previously treated advanced or recurrent nasopharyngeal carcinoma and other solid tumors. *Oncologist* 2019;24:891–e431.
- Hellmann MD, Nathanson T, Rizvi H, *et al.* Genomic features of response to combination immunotherapy in patients with advanced non-small-cell lung cancer. *Cancer Cell* 2018;33:843–52.
- Reck M, Rodríguez-Abreu D, Robinson AG, *et al.* Pembrolizumab versus chemotherapy for PD-L1-Positive Non-Small-Cell lung cancer. *N Engl J Med Overseas Ed* 2016;375:1823–33.
- Voskoboinik I, Whisstock JC, Trapani JA. Perforin and granzymes: function, dysfunction and human pathology. *Nat Rev Immunol* 2015;15:388–400.
- Hou Q, Zhao T, Zhang H, *et al.* Granzyme H induces apoptosis of target tumor cells characterized by DNA fragmentation and Bid-dependent mitochondrial damage. *Mol Immunol* 2008;45:1044–55.
- Larimer BM, Wehrenberg-Klee E, Dubois F, *et al.* Granzyme B PET imaging as a predictive biomarker of immunotherapy response. *Cancer Res* 2017;77:2318–27.
- Hurkmans DP, Basak EA, Schepers N, *et al.* Granzyme B is correlated with clinical outcome after PD-1 blockade in patients with stage IV non-small-cell lung cancer. *J Immunother Cancer* 2020;8:e000586.
- Bolger AM, Lohse M, Usadel B. Trimmomatic: a flexible trimmer for illumina sequence data. *Bioinformatics* 2014;30:2114–20.
- Li H, Durbin R. Fast and accurate short read alignment with Burrows-Wheeler transform. *Bioinformatics* 2009;25:1754–60.
- Picard toolkit. GitHub repository. Broad Institute, 2019. Available: <http://broadinstitute.github.io/picard/>
- Van der Auwera GA, Carneiro MO, Hartl C, *et al.* From FastQ data to high confidence variant calls: the genome analysis toolkit best practices pipeline. *Curr Protoc Bioinformatics* 2013;43:11.10.1–11.10.33.
- Cibulskis K, McKenna A, Fennell T, *et al.* ContEst: estimating cross-contamination of human samples in next-generation sequencing data. *Bioinformatics* 2011;27:2601–2.
- Cibulskis K, Lawrence MS, Carter SL, *et al.* Sensitive detection of somatic point mutations in impure and heterogeneous cancer samples. *Nat Biotechnol* 2013;31:213–9.
- Fang H, Bergmann EA, Arora K, *et al.* Indel variant analysis of short-read sequencing data with scalpel. *Nat Protoc* 2016;11:2529–48.
- Talevich E, Shain AH, Botton T, *et al.* CNVkit: genome-wide copy number detection and visualization from targeted DNA sequencing. *PLoS Comput Biol* 2016;12:e1004873.
- Poniah P, Mohd Zain S, Abdul Razack AH, *et al.* Genome-Wide copy number analysis reveals candidate gene loci that confer susceptibility to high-grade prostate cancer. *Urol Oncol* 2017;35:545.e1–545.e11.
- Zhang L, MacIsaac KD, Zhou T, *et al.* Genomic analysis of nasopharyngeal carcinoma reveals TME-Based subtypes. *Mol Cancer Res* 2017;15:1722–32.
- Airolidi M, Pedani F, Marchionatti S, *et al.* Carboplatin plus taxol is an effective third-line regimen in recurrent undifferentiated nasopharyngeal carcinoma. *Tumori* 2002;88:273–6.

- 32 Foo K-F, Tan E-H, Leong S-S, *et al.* Gemcitabine in metastatic nasopharyngeal carcinoma of the undifferentiated type. *Annals of Oncology* 2002;13:150–6.
- 33 Topalian SL, Hodi FS, Brahmer JR, *et al.* Five-Year survival and correlates among patients with advanced melanoma, renal cell carcinoma, or non-small cell lung cancer treated with nivolumab. *JAMA Oncol* 2019;5:1411.
- 34 Tumei PC, Hellmann MD, Hamid O, *et al.* Liver metastasis and treatment outcome with anti-PD-1 monoclonal antibody in patients with melanoma and NSCLC. *Cancer Immunol Res* 2017;5:417–24.
- 35 Tang H, Li C, Wang L, *et al.* Granzyme H of cytotoxic lymphocytes is required for clearance of the hepatitis B virus through cleavage of the hepatitis B virus X protein. *J.i.* 2012;188:824–31.
- 36 Chen Y-P, Yin J-H, Li W-F, *et al.* Single-Cell transcriptomics reveals regulators underlying immune cell diversity and immune subtypes associated with prognosis in nasopharyngeal carcinoma. *Cell Res* 2020;30:1024–42.
- 37 Jin S, Li R, Chen M-Y, *et al.* Single-Cell transcriptomic analysis defines the interplay between tumor cells, viral infection, and the microenvironment in nasopharyngeal carcinoma. *Cell Res* 2020;30:950–65.
- 38 Gao J, Shi LZ, Zhao H, *et al.* Loss of IFN- γ pathway genes in tumor cells as a mechanism of resistance to anti-CTLA-4 therapy. *Cell* 2016;167:397–404.
- 39 Zhou Z, He H, Wang K, *et al.* Granzyme A from cytotoxic lymphocytes cleaves GSDMB to trigger pyroptosis in target cells. *Science* 2020;368:eaaz7548.
- 40 Zychlinsky A, Zheng LM, Liu CC, *et al.* Cytolytic lymphocytes induce both apoptosis and necrosis in target cells. *J Immunol* 1991;146:393–400.
- 41 Bergsbaken T, Fink SL, Cookson BT. Pyroptosis: host cell death and inflammation. *Nat Rev Microbiol* 2009;7:99–109.
- 42 Zhang Z, Zhang Y, Xia S, *et al.* Gasdermin E suppresses tumour growth by activating anti-tumour immunity. *Nature* 2020;579:415–20.