Supplemental figure 1
Supplemental figure S1. (A) Wild-type (WT) parental LLC cell line, WT clones derived from untreated WT LLC cell line or cisplatin-resistant (R) LLC clones were maintained in control conditions or treated with cisplatin (20 µmol/L) for 48 hours. Thereafter, cells were subjected to the cytofluorometric assessment of apoptosis-related parameters upon co-staining with the vital dye propidium iodide (PI) and the mitochondrial membrane potential-sensing dye DiOC6(3). White and black columns illustrate the percentage of dying [DiOC6(3)\textsuperscript{low}PI\textsuperscript{-}] and dead (PI\textsuperscript{*}) cells respectively. Data are reported as means ± SEM (n=4). *P<0.05, **P<0.01, ***P<0.001 (Student t-test), as compared to equally treated WT parental cell line; #P<0.05, ##P<0.01, ###P<0.001 (Student t-test), as compared to equally treated R7 clone. (B) Representative immunoblots showing higher levels of PARylation in R cells compared to WT cells cultured in normal growth medium. Actin levels were monitored to ensure equal loading of lanes. (C) Representative plots of the gating strategy used in flow cytometry analysis of T cell infiltrate from harvested tumor. Lymphocytes were characterized by a low size and granularity. Debris was excluded on SSC versus FSC plot. Then, Yellow Live/Dead dye-positive i.e. non-viable cells were gated out. Live cells were analyzed for expression of CD3 marker and non-single cells were excluded based on FSC-H versus FSC-A. A CD4 versus CD8 plot was used to distinguish CD8\textsuperscript{+}CD4\textsuperscript{-} and CD4\textsuperscript{+}CD8\textsuperscript{-} T cells out of CD3\textsuperscript{+} cells.

Abbreviations: CD: Cluster of differentiation; KDa: KiloDalton, PAR: Poly Adenosine Ribose; PI: Propidium iodide; WT: Wild Type; FSC: Forward scatter; SSC: Side scatter.