

**EXPANSION OF TUMOR-INFILTRATING LYMPHOCYTES AND MARROW-INFILTRATING LYMPHOCYTES FROM PEDIATRIC MALIGNANT SOLID TUMORS**

<sup>1</sup>Jonathan Metts\*, <sup>2</sup>Jonathan Hensel, <sup>2</sup>Alejandro Alfaro, <sup>2</sup>Brook Olmo, <sup>2</sup>Shari Pilon-Thomas, <sup>2</sup>John Mullinax, <sup>1</sup>Ivanna Leon. <sup>1</sup>Johns Hopkins All Children's Hospital, St Petersburg, FL, USA; <sup>2</sup>Moffitt Cancer Center, Tampa, FL, USA

**Background** High-risk non-CNS pediatric malignant solid tumors (pMST) have unsatisfactory outcomes, and novel therapies are warranted. Adoptive cellular therapy (ACT) using tumor-infiltrating lymphocytes (TIL) has produced durable responses in melanoma, and improvements in TIL expansion have made ACT-TIL feasible for other solid tumors.<sup>1-3</sup> Preclinical mouse models suggest that T-cells from bone marrow (marrow-infiltrating lymphocytes, MIL) have antitumor reactivity offering another source for ACT.<sup>4, 5</sup> To demonstrate feasibility of ACT in pMST we hypothesized that TIL/MIL can be expanded from these patients.

**Methods** Patients ≤21 years old undergoing standard-of-care pMST resection were enrolled on an IRB approved protocol. Fresh tumor (≥1 cm<sup>3</sup>) was collected and bone marrow (10 mL) was obtained when accessible from standard of care procedures. TIL/MIL were cultured in media containing IL-2 (6000 IU/mL). TIL were expanded from tumor fragment cultures (TFC, >1 mm<sup>3</sup>) or tumor digest. Select TIL samples were further expanded using a rapid expansion protocol (REP). Phenotype of expanded TIL (CD3, CD4, CD8 and CD56) was evaluated using flow cytometry. IFN- $\gamma$  secretion, measured by ELISA assay, measured tumor-specific reactivity after co-culture with autologous tumor and TIL.

**Results** Twenty samples were obtained between March 2019-May 2021. Two samples were ineligible (final pathology not pMST), leaving 18 samples for analysis. Five marrow samples were collected. TIL were expanded from 14/18 samples (78%) through TFC with median  $5.17 \times 10^6$  cells (range  $1.86 \times 10^6$ – $3.21 \times 10^8$ ). Average phenotype (%) of TFC-TIL were CD3 (63.17), CD4 (21.46), CD8 (46.19) and CD56 (32.68). 9/10 (90%) of samples successfully underwent REP with median  $9.35 \times 10^7$  cells (range  $2.49 \times 10^7$ – $5.86 \times 10^8$ ) final viable TIL and average fold-change 718.6 (median 458.6). Average phenotype (%) of post-REP TIL were CD3 (96.04), CD4 (75.04), CD8 (19.17) and CD56 (0.43). TIL were expanded from TFC of therapy-naïve (8/10, 80%) and pretreated (chemotherapy and checkpoint immunotherapy) samples (5/8, 63%). Seven samples had sufficient tissue to test tumor-specific reactivity; all were non-reactive. MIL pre-REP was expanded from four samples with median  $9.55 \times 10^6$  cells (range  $8.00 \times 10^5$ – $1.00 \times 10^7$ ). Average phenotype of expanded MIL (%) were CD3 (45.17), CD4 (24.46), CD8 (36.15) and CD56 (28.21) (table 1).

Results of TIL and MIL expansion from 18 pMST samples. Abbreviations: Dx: diagnosis, pre-REP: pre-rapid expansion protocol, post-REP: post-rapid expansion protocol, PBMC: peripheral blood mononuclear cells, GNB: ganglioneuroblastoma, WT: Wilms tumor, OS: osteosarcoma, NB: neuroblastoma, IMT: inflammatory myofibroblastic tumor; ASPs: alveolar soft part sarcoma, SS: synovial sarcoma, ERMS: embryonal rhabdomyosarcoma, N: no systemic therapy, C: chemotherapy, I: immunotherapy, DNG: did not grow, N/A: not applicable, NR: non-reactive

**Abstract 178 Table 1 Expansion of TIL from pMST**

Sample	Dx	Age (years)	Staging	Diagnosis	Sampling	Sample Source	Prior Therapy	Tumor Fragment Culture (cells)	post-REP (cells)	Digest (cells)	MIL (cells)	Reactivity
1	GNB	3	Localized	Initial treatment	Uplift resection	Abdomen, primary site	N	DNG	N/A	none available	$1.00 \times 10^7$ pre-REP, $5.20 \times 10^6$ post-REP	NR
2	WT	2	IV, favorable histology	Initial treatment	Uplift resection	Abdomen, primary site	N	$4.68 \times 10^6$	N/A	$1.03 \times 10^8$	N/A	NR
3	WT	1	IV, favorable histology	Initial treatment	Uplift resection	Abdomen, primary site	N	$3.20 \times 10^6$	$4.03 \times 10^7$	$1.60 \times 10^8$	N/A	NR
4	OS	15	Metastatic	Relapse	After salvage chemotherapy	Lung, metastatic site	C	$6.00 \times 10^6$	$5.46 \times 10^7$	$7.25 \times 10^8$	N/A	N/A
5	WT	7	IV, anaplastic	Initial treatment	Uplift resection	Abdomen, lymph node	N	$2.50 \times 10^6$	DNG	$8.63 \times 10^6$	N/A	N/A
6	NB	7	IV, high-risk	Relapse	At resection	CNS, metastatic site	C	$5.63 \times 10^6$	$3.17 \times 10^8$	$4.23 \times 10^8$	N/A	NR
7	IMT	14	Localized	Initial treatment	Uplift resection	Lung, Primary site	N	$3.23 \times 10^6$	N/A	$3.00 \times 10^7$	N/A	N/A
8	NB	16	Stage III	Initial treatment	Uplift resection	Lower Extremity, Primary site	N	$1.88 \times 10^6$	N/A	$1.00 \times 10^8$	N/A	N/A
9	OS	14	Metastatic	Initial treatment	After neoadjuvant chemotherapy	Lower Extremity, Primary site	C	$4.90 \times 10^7$	N/A	none available	$9.12 \times 10^6$ pre-REP	N/A
10	ASPS	20	Metastatic	Relapse	After salvage therapy	CNS, metastatic site	I	DNG	N/A	$2.00 \times 10^8$	N/A	N/A
11	OS	15	Metastatic	Relapse	After salvage therapy	Lung, metastatic site	C	DNG	N/A	$3.20 \times 10^8$	N/A	N/A
12	ASPS	20	Metastatic	Relapse	After salvage therapy	CNS, metastatic site	I	DNG	N/A	$5.53 \times 10^7$	N/A	N/A
13	NB	7	IV, high-risk	Initial treatment	Diagnostic biopsy	Abdomen, primary site	N	$4.00 \times 10^6$	$9.35 \times 10^7$	used in co-culture	$7.26 \times 10^7$ (post-REP only)	NR
14	SS	13	Stage II	Initial treatment	Uplift resection	Lower Extremity, Primary site	N	$3.10 \times 10^6$	$3.95 \times 10^7$	used in co-culture	N/A	NR
15	ERMS	3	Stage III, group 2	Initial treatment	Uplift resection	Abdomen, primary site	N	$1.16 \times 10^6$	$3.22 \times 10^8$	used in co-culture	N/A	NR
16	OS	14	Localized	Initial treatment	After neoadjuvant chemotherapy	Lower Extremity, Primary site	C	$1.37 \times 10^8$	$4.47 \times 10^8$	none available	$8.00 \times 10^6$ pre-REP	N/A
17	WT	4	Stage IV, favorable histology	Initial treatment	Diagnostic biopsy	Abdomen, Primary site	N	$5.40 \times 10^7$	$5.86 \times 10^8$	$8.92 \times 10^7$	N/A	N/A
18	OS	12	Localized	Initial treatment	After neoadjuvant chemotherapy	Lower Extremity, Primary	C	$3.51 \times 10^6$	$2.49 \times 10^7$	$4.00 \times 10^8$	$1.00 \times 10^7$ pre-REP, $1.40 \times 10^8$ post-REP	N/A

**Conclusions** This study demonstrates feasibility of pMST TIL expansion ex vivo. Due to tissue volume constraints inherent in pMST sampling, anti-tumor reactivity testing was not feasible for most patients. Determining optimal strategy for TIL-ACT in pMST will require further investigation regarding techniques for expanding tumor-specific TIL.

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**Ethics Approval** This study was approved by the Johns Hopkins All Children's Hospital IRB (#IRB00193453). Consent was obtained from the patient or parent, as appropriate for age, prior to participating in this study.

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