BET INHIBITORS SYNERGIZE WITH ANTI-PD1 BY RESCUING TCF1+ PROGENITOR EXHAUSTED T CELLS IN ACUTE MYELOID LEUKEMIA

Kyle Romine, 1Hyun-Jun Cho, 1Yoko Kosaka, 1Kaelan Byrd, 1Jesse Cox, 1Patrick Flynn, 1Matthew Newman, 1Christopher Loo, 1Evan Lind*, 1Oregon Health & Science University, Portland, OR, United States; 2Oregon Health and Sciences University, Portland, OR, United States; 3Oregon Health & Science University, Portland, OR, United States; 4OHSU, Portland, United States

Abstract

Acute Myeloid Leukemia (AML) is the most common adult leukemia and has a very poor prognosis. With a 5-year survival of under 30% (seer.cancer.gov), most people diagnosed with AML will die from the disease. AML is caused by an uncontrolled proliferation of poorly differentiated myeloid precursor cells which results from a combination of three classes of mutations that affect proliferation, differentiation and epigenetic state. For this reason, drugs targeting epigenetic modifications are being actively studied in AML. AML has been shown to avoid immune recognition though inhibiting the function of multiple cell types, especially T cells and therefore immune checkpoint blockade presents a promising therapy for any immune-targeted strategy; however, clinical trials to date have shown very modest efficacy. T cell exhaustion in cancer has been shown to be a regulated process involving transcriptional and epigenetic changes. BRD4 has been shown to be important for maintaining this exhaustion state. It stands to reason that drugs designed to target epigenetic pathways in tumors will have effects on T cell populations present in the tumor microenvironment. In these studies, we investigated the effects of the BET inhibitor (BETi) JQ1 on T cell exhaustion and checkpoint responsiveness in a murine model of AML.

Abstract Figure 1

T cell exhaustion in the AML mouse model. (A) Cytotoxic T cells show an exhausted phenotype in mice with AML. Spleen cultures from mice with AML or WT controls were stained with antibodies to CD3, CD8, TIM3, PD1, and TCF1. Left shows percent of TPEX CD8 T cells. Right panel shows TEX CD8 T cells. N = 12 animals per group. (B) Proliferative defect in T cells in mice with AML. Splenocytes were labeled with the proliferation dye CFSE. Whole spleen suspensions were stimulated with anti-CD3 or anti-CD3 and anti-CD28 for 3 days. FACs plots show proliferation of T cells in each condition

Abstract Figure 2

Treatment with JQ1 results in expansion of T cells with TPEX. (A) Example of proliferation (CFSE dilution) vs TCF-1 expression showing unstimulated, CD3 or CD3+JQ1 120 nM in in vitro 3-day culture. Results gated on CD8 T cells. (B) Summary of T cell proliferation from 4 independent experiments showing the percent proliferation of CD8 T cells with TPEX (PD1+ Tim-3- TCF-1+) (black line) or TEX (PD1+Tim+3+TCF1-) phenotype (red line). Statistics are unpaired T-Test for each treatment condition.

Abstract Figure 3

In vivo treatment of FTL mice with the BETi JQ1. (A) Schematic overview of treatment protocol. (B) White blood cell counts at pre-treatment, 1 week and 2 weeks after JQ1, PD1 blockade or both. (C) Percent of CD8+ T cells of all CD3-gated T cells in the BM of treated animals. (D) A-C Percent of precursor-exhausted CD8+ T cells as a percent of all T cells in the spleen of treated animals. Results combined from 2 separate experiments n=7. D One experiment n=3.
Methods The AML mouse model bears FLT3-ITD and deletion of TET2 restricted to the myeloid lineage. For in vitro studies, splenocytes were stimulated with anti-CD3 and either JQ1, anti-PD1 or both and proliferation and differentiation status were assessed by flow cytometry. For in vivo studies, treatment consisted of 2 weeks with JQ1, anti-PD1 or both.

Results This mouse model of AML exhibits an expansion of terminally exhausted T cells and impaired proliferative capacity after stimulation through the TCR (figure 1). Ex vivo treatment with BETi and anti-PD1 reverses CD8+ T cell exhaustion via rescue of proliferative dysfunction and expansion of more functional precursor exhausted T cells (TPEx-CD8, PD1+, TCF1+, TIM3-) (figure 2). Finally, we show that BETi synergizes with anti-PD1 in vivo leading to a reduction of tumor cells in multiple organ sites, and enrichment of CD8+ T cells in the bone marrow (figure 3).

Conclusions Using an AML mouse model that exhibits leukemia-induced immune exhaustion, we demonstrate the pre-clinical efficacy of combining BETi and anti-PD1 therapy in the treatment of AML.

REFERENCES


Ethics Approval This study has been approved by the OHSU IACUC committee protocol IP00000907 “Immune-based therapeutic approaches for acute myeloid leukemia” Evan Lind PI.

http://dx.doi.org/10.1136/jitc-2021-SITC2021.734