

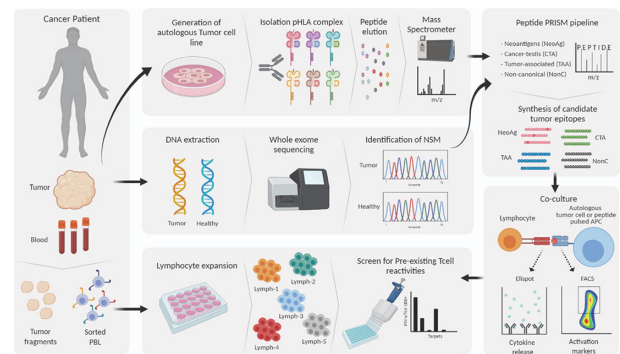
PREFERENTIAL RECOGNITION OF NEOANTIGENS OVER NON-CANONICAL PEPTIDES IN CANCER PATIENTS

¹Maria Lozano-Rabella*, ¹Andrea Garcia-Garjjo, ¹Jara Palomero, ²Florian Erhard, ³Juan Martín-Liberal, ¹Ignacio Matos, ⁴Jared Gartner, ⁴Steven Rosenberg, ⁵Michael Ghosh, ¹Francesc Canals, ⁶August Vidal, ⁷Josep Maria Piulats, ⁶Xavier Matias-Guiu, ¹Irene Braña, ⁸Eva Muñoz, ¹Elena Garralda, ²Andreas Schlosser, ¹Alena Gros. ¹Vall d'Hebrón Institute of Oncology (VHIO), Barcelona, Spain; ²Julius-Maximilians-University Würzburg, Würzburg, Germany; ³Institut Català d'Oncologia (ICO), Barcelona, Spain; ⁴National Cancer Institute (NCI), National Institutes of Health, Bethesda, MD, USA; ⁵University of Tübingen, Interfaculty Institute for Cell Biology, Tübingen, Germany; ⁶Bellvitge University Hospital, Barcelona, Spain; ⁷Catalan Institute of Oncology (ICO), Barcelona, Spain; ⁸Vall d'Hebron Hospital, Barcelona, Spain

Background Despite recent advances in exome and RNA sequencing to identify tumor-rejection antigens including neoantigens, the existing techniques fail to identify the vast majority of antigens targeted by tumor-reactive cells. A growing number of studies suggest that HLA-I peptides derived from non-canonical (nonC) open reading frames or derived from allegedly non-coding regions can contribute to tumor immunogenicity. Here we use proteogenomics to identify personalized candidate canonical and non-canonical tumor-rejection antigens and to evaluate their contribution to cancer immune surveillance in patients.

Methods Whole exome sequencing was performed to identify the non-synonymous somatic mutations (NSM) and immunopeptidomics to identify the HLA-I presented peptides (pHLA) in 9 patient-derived tumor cell lines (TCL). Peptide-PRISM proteogenomics pipeline was used to identify both canonical and non-canonical pHLA, including those derived from NSM in coding regions. All peptides containing mutations and derived from either cancer-testis (CTA) or tumor-associated antigens (TAA) were selected as candidate tumor antigens. For nonC peptides, an immunopeptidomics healthy dataset containing several tissues and HLA-allotypes was used to eliminate those derived from normal ORFs and select nonC peptides preferentially expressed in tumor cells (nonC-TE). The selected candidate peptides were synthesized, pulsed onto autologous APCs and co-cultured with tumor-reactive ex vivo expanded lymphocytes to assess immune recognition (figure 1).

Results NonC-TE peptides were identified in all TCL studied, ranging from 0.5% to 5.4% of the total HLA-I presented peptides (n= 506). As described previously, 5'UTR were the main source. Of note, the tumor type did not have an impact on the frequency of presented nonC peptides, but rather the presence of HLA-A*11:01 and HLA-A*03:01 was a major determinant. T cell responses were detected against at least 13/33 putative neoantigens, 2/24 CTA and 2/61 TAA. On the contrary, none of the 471 nonC-TE candidate peptides tested thus far, including one containing a NSM were able to elicit a recall immune response. Nevertheless, T cells recognizing at least 3 of them were detected through in vitro sensitization of non-autologous PBMCs.



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Tumor biopsies and blood samples are obtained from cancer patients (left panel). Patient-derived tumor cell lines are generated in vitro, the peptides presented on HLA molecules are further isolated and analyzed in a mass-spectrometer (top panel). Whole exome sequencing (WES) from matched tumor and healthy tissue is performed to identify the non-synonymous somatic mutations (NSM) (middle panel). Peptide-PRISM proteogenomics pipeline combines the information from the immunopeptidomics data and WES to identify pHLA sequences from both canonical and non-canonical candidate tumor antigens (top right panel). Lymphocyte populations either TILs or sorted PBMCs are expanded and further screened for pre-existing T cell responses (bottom panel) against the candidate epitopes by co-culturing the T cells with peptide-pulsed autologous APC. The recognition is assessed by measuring IFN γ release by elisot and the upregulation of activation surface markers by FACS (bottom right panel).

Conclusions Our results show that although HLA-I nonC peptides were frequently presented in all TCLs studied and they can be immunogenic, neoantigens derived from mutations in canonical coding regions were preferentially recognized by tumor-reactive lymphocytes, suggesting T cells targeting the latter are primed more efficiently. The identification of mutated nonC antigens using whole genome sequencing to identify mutations in non-coding regions warrants further examination. Still, the specificity of many tumor-reactive TILs remains unknown.

Ethics Approval "This study was approved by the "Comité de Ética de Investigación con Medicamentos del Hospital Universitario Vall d'Hebron" institution's Ethics Board; approval number PR(AG)537/2019."

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